1	New insights into the effects of support matrix on the removal of organic micro-
2	pollutants and the microbial community in constructed wetlands
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25 Abstract

Constructed wetlands (CWs) are an eco-friendly and cost-effective technology to remove organic 26 micro-pollutants (OMPs) from wastewaters. The support matrix is an important component in 27 CWs as it has a primary role in the growth and development of plants and microbes. However, 28 29 the roles of the support matrix in CWs in removing OMP have not been systematically studied. Therefore, in this study, six common materials (sand, zeolite, blast iron slag, petcoke, polonite 30 31 and crushed autoclaved aerated concrete (CAAC)) as support matrixes were firstly investigated 32 by batch tests to explore their adsorption capacities to selected OMPs (ibuprofen, iohexol, 33 tebuconazole and imazalil). Results showed that the adsorption capacities of the materials were limited (at the level of $\mu g/g$) compared to well-known sorbents (at the level of mg/g), such as 34 35 activated carbon and carbon nanotubes. Column packed with the six materials, respectively, 36 were then built up to study the effects of different materials on microbial community. In the 37 medium-term study (66 days), the removal of four OMPs in all the columns increased by 2-58% from day 25 to day 66, and was mainly attributed to microbial degradation. Furthermore, 38 Community-level physiological profiling (CLPP) analysis indicates that material presence shaped 39 the microbial communities not only in the interstitial water but also in the biofilm. Overall, all the 40 41 findings demonstrate that although the adsorption capacities of the common materials are low, 42 they may be a driver to improve the removal of OMPs by altering the microbial community in CWs. 43

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45 **Keywords**: Pharmaceuticals, Biocides, Biodegradation, Adsorption, Community-level

46 physiological profiling (CLPP), Microbial community

47 **1. Introduction**

Constructed wetlands (CWs) as an eco-friendly and cost-effective biotechnology for wastewater treatment, have been demonstrated to have potential for removing organic micropollutants (OMPs) from wastewater [1, 2]. The removal of OMPs in CWs is typically attributed to substrate sorption, plant uptake and biodegradation processes together [3]. Among these processes, biodegradation has been pointed to be a major process for OMP removal in CWs [4-6]. Plant uptake and biodegradation have been more studied [7-10], but the role of the support matrix (CW substrate) in removing OMPs has not been fully addressed.

55 The support matrix is an important component in CWs as it has a primary role as physical support for the growth and development of plants and biofilm [3]. The material used as support 56 57 matrix may influence the removal of OMPs by affecting the microbial community structures in CWs. To date, the effects of different support matrixes on microbial community structures in CWs 58 59 have not been elucidated yet. In addition, the support matrix can also directly interact with OMPs by sorption processes, depending on the materials employed. These interactions could highly 60 61 influence the performance of CW systems. Thus, an appropriate selection of support matrix may be a determining step to improve the removal of OMPs in CWs. 62

Some well-known good sorbents are generally high priced, limiting their wide application as CW media. Due to the low-cost/low-tech approach used by CW practitioners, the cheaply and locally available materials from natural sources, industrial by-products and manmade products have been mostly used [11, 12]. In fact, for the last decade, large efforts have been made in identifying low cost materials with potential for enhanced phosphorus removal,

68 such as sand [13-15], gravel [16], polonite [17], light expanded clay aggregates (LECA) [18], 69 crushed autoclaved aerated concrete (CAAC) [19], fly ash [20, 21] and slags from steel and power 70 plants [17] etc. However, concerning OMPs, the sorption capacities of different materials have 71 not been well documented. Although some works have reported the sorption of OMPs 72 (pharmaceuticals, pesticides and phenolic compounds, etc.) by materials, such as activated 73 carbon [22-24], clay-based materials [25-27], zeolites and other siliceous materials [28, 29] and industrial and agricultural wastes and by-products [30-32], there is still a number of common 74 75 materials used in CWs that need to be studied. Furthermore, only a preliminary indication of the efficiency or kinetics of the sorption processes can be provided as most work consisted of batch 76 sorption studies at much higher OMP concentration than that observed in untreated or treated 77 78 wastewater [3]. The sorption capacities of common materials (CW substrates) at typical concentration ranges of OMPs in real wastewater (ng to μ g/L) are still unknown. 79

80 Filling the knowledge gaps on the sorption capacities of different materials for OMPs, as well as the effects of materials on microbial community, and consequently biodegradation, will 81 82 allow to better understand the mechanisms behind OMP removal in CWs. As such, a range of substrates typically used in CWs is presently selected: natural materials (sand, zeolite and 83 84 polonite), industrial by-products (blast iron slag, petroleum coke (petcoke) and man-made 85 products (crushed autoclaved aerated concrete (CAAC)). Sand is one of the most common media 86 used in CWs. Zeolite is a low-cost and readily available material, known to be effective to treat ammonium-containing wastewater [33]. Blast-iron slags, polonite and CAAC are often employed 87 for the removal of phosphorus [17, 19]. Petcoke is a carbonaceous hydrophobic black solid 88

material that is delivered from the refinery coker units of crude oil. It is the precursor material
for activated carbon preparation [34].

Two pharmaceuticals (ibuprofen and iohexol) and two pesticides (tebuconazole and imazalil) were used as the target compounds. Ibuprofen and iohexol, widely used in our daily life and hospital respectively, are frequently detected in environmental samples [35-37]. The pesticides tebuconazole and imazalil are relevant in the agricultural setting but also commonly occur in storm water or surface water from urban settings [38, 39]. Physico-chemical properties of the four OMPs are summarized in Table S1.

We aimed to study for the aforementioned four model OMPs: (a) adsorption to the selected materials by short-term isotherm experiments; (b) sorption and biodegradation in packed columns under medium-term operation (66 days) with spiked real wastewater. In addition, the similarity / dissimilarity of the microbial community functional profile in the columns was investigated using community level physiological profiling (CLPP) analysis. CLPP is an easy, accurate and rapid biotechnology, and has been widely employed for studying the functionality of microbial community in CWs [40-44].

104

105 2. Materials and methods

106 **2.1. Chemicals**

107 Methanol (>99.9%) and formic acid (98 %, reagent ACS) were purchased from Merck 108 (Darmstadt, Germany). In the batch adsorption tests, analytical quality standards of ibuprofen

(98% purity), iohexol (95% purity), tebuconazole (99.3% purity) and imazalil (99.8% purity) were supplied by Sigma-Aldrich (Munich, Germany). In the packed column experiment, commercial products of the same compounds were purchased in local stores. The exact concentration of the four compounds in the commercial products were determined prior to performing the study, in order to most adequately dose the compounds in the wastewater.

114 **2.2. Batch adsorption experiment**

Adsorption isotherms were investigated by batch tests at 20 °C for the six selected 115 116 materials (sand, zeolite, blast iron slag, petcoke, polonite and CAAC) (Fig. S1). These materials were purchased from Vestergård (Denmark), Silkem (Slovenia), HACO (Norway), Carbomax 117 (Sweden) Ecofiltration (Sweden), Silikazit (Belgium), respectively. The physical and chemical 118 119 characteristics of the six materials are presented in Table 1. Equilibrium isotherm experiments were performed (n=2) using sealed aliquots of 500 ml of tap water with 30 g of sand, 5g of zeolite, 120 121 petcoke, polonite or CAAC and 10 g of blast iron slag, in brown glass bottles continuously agitated 122 in a shaker (100 rpm) for 24h. Each model compound was studied separately by spiking 10 123 different concentrations (0, 10, 30, 60 µg/L, 0.1, 0.25, 0.5, 1, 5, 10 mg/L). At the end of the equilibrium period (24h), the bottles were placed on a desk for 4 h to allow settling of materials 124 before aliquots of the supernatant were collected for further analysis. Bottles without any 125 adsorbents were used as blanks to monitor the loss of adsorbates during the experiment. The 126 127 amount of model compounds adsorbed onto materials, $q_t (\mu g/g)$ was calculated by mass-balance 128 relationship Eq. (1). Two non-linear isothermal models commonly used to describe the adsorption process, namely Freundlich (Eq. 2) and Langmuir (Eq. 3) isotherm models were 129

130 presently applied.

131
$$q_t = (C_0 - C_t) \frac{V}{W}$$
(1)

where C_0 and C_t are the initial and time t liquid-phase concentration of the model compounds (μ g/L), respectively, *v* is the volume of the solution (L) and *w* is the weight of the dry materials used (g).

135
$$q_e = K_f C_e^{1/N}$$
 (2)

136
$$q_e = \frac{q_m k_l C_e}{1 + k_l C_e}$$
 (3)

where, q_e is the mass of adsorbate per mass unit of adsorbent at equilibrium (µg/g), C_e is the equilibrium adsorbate concentration (mg/L), K_f is the Freundlich adsorption constant (1/(µg/L)^N), N is the degree of non-linearity, q_m is the maximum mass adsorbed at saturation conditions per mass unit of adsorbent (µg/g) and K_i is the Langmuir affinity coefficient, respectively.

142	Table 1. Physical parameters of sand, zeolite, blast iron slag, petcoke, polonite and tobermorite tested in this
143	work (mean ± standard deviation).

Material	Sand	Zeolite	Blast iron	Petcoke	Polonite	CAAC
			slag			
d10 (mm)	0.46	1.38	0.22	0.53	1.43	3.42
d60 (mm)	1.18	2.46	0.47	2.77	3.31	5.65
Uniformity						
coefficient	2.57	1.78	2.14	5.23	2.31	1.65
(d60/ d10)						
Porosity (%)	38 ± 2	52 ± 1	44 ± 3	38 ± 1	52 ± 2	47 ± 2
Density (g/cm3)	2.62 ± 0.01	2.05 ± 0.02	5.72 ± 0.35	1.95 ± 0.09	2.03 ± 0.05	0.75 ± 0.01
pH in water	8.28 ± 0.03	9.79 ± 0.11	7.74 ± 0.02	7.34 ± 0.02	9.11 ± 0.03	8.10 ± 0.03

145 **2.3. Packed column experiment**

146 **2.3.1 Column startup and operation**

147 A total of 18 columns were divided by the same six types of materials (in triplicate): sand, 148 zeolites, blast iron slag, polonite, petcoke and CAAC columns. Each column was set up in a 2.3 L 149 polyvinylchloride (PVC) cylinder (diameter: 10.8 cm, height: 25 cm), equipped with an outlet at a 150 height of 20.5 cm and a perforated hose as inlet in the bottom of the column (Fig. S2). Each 151 column was filled with 23 cm of the respective material. As for the blast iron slag columns, they 152 were filled with a mixture of sand and blast iron slag (10:1, weight/weight) (thereafter referred to as sand+iron columns). The specific volume weight in the sand+iron, zeolite, sand, polonite, 153 154 petcoke and CAAC columns were 1.47 ± 0.01 , 0.63 ± 0.01 , 1.24 ± 0.01 , 0.76 ± 0.01 , 0.74 ± 0.01 155 and 0.41 ± 0.01 g/cm³, respectively.

156 The set of columns was rain protected, but exposed to naturally daily air temperature (minimum -6 °C and maximum 18 °C) variations. The systems were fed with diluted pig manure 157 wastewater, briefly 270 mg/L COD, 140 mg/L BOD₅, 42 mg/L total nitrogen (TN), 6 mg/L total 158 phosphorus (TP), while none of the target compounds was detected in the diluted wastewater 159 160 (full characterization in Table S2). Additionally, ibuprofen, iohexol, tebuconazole and imazalil 161 were spiked in the influent tank to ensure a continuous influent level of 100 μ g/L. The columns were operated at 6.2 cm/d hydraulic loading rate for 66 days using a peristaltic pump with 24 162 163 channels (BT100-1L multi-channels peristaltic pump, Longer pump, China).

164 **2.3.2. Sampling and analysis**

165	During the whole period of the experiment (66 days), two sampling campaigns were
166	conducted at days 25 and 66, respectively. At each time point, influent and effluent of each
167	column were collected for physical-chemical parameters (water temperature, pH, dissolved
168	oxygen (DO) and electrical conductivity (EC)), nutrients (Chemical oxygen demand (COD),
169	biochemical oxygen demand (BOD $_5$), total organic carbon (TOC), total nitrogen (TN), ammonium
170	(NH ₄ ⁺ -N), nitrate (NO ₃ ⁻ -N), and phosphate (PO ₄ ³⁺ -P)) and the four OMPs (ibuprofen, iohexol,
171	tebuconazole and imazalil). Water temperature, pH, DO and EC were analyzed in-situ using
172	portable meters (Multi-Parameter Meter HQ40d, and sensION+ EC5, HACH, USA). In the lab
173	within 24h, COD was colorimetrically determined (DR 3900 Spectrophotometer, Hach) following
174	dichromate digestion according to Standard Methods [45]. The BOD ₅ was measured using
175	respirometric method (WTW OxiTOP $^{\odot;*}$). The TSS was determined following Standard Methods
176	[45]. NH ₄ -N, NO ₃ -N, and PO ₄ -P were measured by QuikChem Methods [®] (10-107-06-3-D, 10-107-
177	04-1-C, and 10-115-01-1-A, respectively) on an automated flow injection analyzer (QuikChem
178	FIA+ 8000 Series, Lachat instruments, Milwaukee, USA). TN and TOC were analyzed by the TNM-
179	1 unit of a TOC-V analyzer (Shimadzu, Japan). The ibuprofen, iohexol, tebuconazole and imazalil
180	concentrations were determined using high-performance liquid chromatography (HPLC) (Thermo
181	Scientific Ultimate 3000) with diode array detection (DAD) after solid phase extraction according
182	to pre-established methods [10, 46]. Briefly, 100 mL of influent samples or 500 mL of effluent
183	samples were extracted using Strata-X cartridges (Phenomenex), eluted with 5 mL of a mixture
184	of methanol and formic acid (v/v, 9:1), further dried and dissolved in 1 mL of a mixture of
185	methanol and water (v/v, 1:1). The concentrated samples were then injected in the HPLC.

At day 66, after collection of water samples, each column was vigorously shaken for one minute, and the interstitial water collected in a 1 L sterilized amber bottle. In addition, columns were destroyed and a substrate sample was taken as a composite mixture of the materials in each column. Substrate was stored in a 50 mL sterilized falcon tubes for biofilm analysis. The collected substrate samples further here named 'biofilm'. All these water and biofilm samples were processed within a 5 h period for CLPP.

The collected substrate was also stored for quantifying the substrate total organic carbon (SOC) according to Standard Methods [45] and the sorbed ibuprofen, iohexol, tebuconazole and imazalil concentration following pre-established methods [10, 46]. Samples were kept at -8 °C, freeze dried and processed for analysis within a two weeks period. Briefly, 2 g of material were mixed with 10 mL of a mixture of methanol:acetone (95:5, v:v) and ultrasonicated for 30 min. The slurry was then centrifuged and the supernatant separated for determination by HPLC-DAD. The analytical figures of merit of the HPLC methodology are presented in Table S3.

199 Pollutants (COD, TOC, BOD_5 , TN, NH_4^+ -N, TP, ibuprofen, iohexol, tebuconazole and 200 imazalil) mass removal efficiency (MRE) was determined using Eq. (4)

201
$$MRE = \frac{C_{in} \times V_{in} - C_e \times V_e}{C_{in} \times V_{in}} \times 100\%$$
(4)

Where C_{in} and C_e represent the influent and effluent concentrations of pollutants (mg/L or μ g/L), respectively. V_{in} and V_e are the total volumes of the influent and effluent water in a certain time (t). The substrate total organic content (SOC) was measured according to the loss of ignition
(LOI) method, described as Eq. (5)

207
$$LOI_{550} = (DW_{105} - DW_{550}) / DW_{550} * 100$$
 (5)

208 Where LOI_{550} means LOI at 550 °C (%). DW_{105} and DW_{550} mean the dry weight of the substrate 209 samples before combustion and after heating to 550 °C, respectively, (g). Substrate 210 concentrations were determined as μ g of OMP per gram of substrate dry weight.

211 One-way Analysis of variance (ANOVA) was used to assess the effects of column type on 212 typically measured water parameters (water temperature, pH, EC and DO), TSS and ibuprofen, 213 iohexol, tebuconazole and imazalil concentrations by material sorption and the pollutants 214 removal (COD, TOC, BOD₅, TN, NH₄⁺-N, TP, ibuprofen, iohexol, tebuconazole and imazalil) at the 215 0.05 significance level using the XLSTAT Pro[®] statistical software (XLSTAT, Paris, France).

216

2.3.3. Community level physiological profiling

The microbial community-level physiological profile (CLPP) of each column was analyzed using BIOLOG[™] Ecoplates (Biolog Inc. Hayward CA, USA). A BIOLOG Ecoplate contains 31 different carbon sources and a blank in three replicates (96 wells in total). Water samples were inoculated directly, while substrate attached biofilm was detached prior to CLPP analysis. The detachment method followed the description by Weber and Legge [47], and more details about CLPP analysis are described in supplementary material.

The CLPP data was analyzed according to the descriptions by Weber et al. [48] and Weber and Legge [49]. The time point selection followed the principle of greatest variance

between well responses and the least number of absorbance values above 2.0 (values above 2.0 225 226 are above the linear absorbance range). Time points selected for the CLPP data analysis in 227 interstitial water, substrate attached biofilm samples and sample types comparison were 54h, 228 36h and 42h, respectively. The average well color development (AWCD) and the number of 229 carbon sources utilized (richness) were calculated [50]. To analyze substrate utilization patterns, 230 the 31 carbon sources were classified into five groups (guilds) as suggested by Weber and Legge [49]: polymers, carbohydrates, carboxylic acids & acetic acids, amino acids and amines/amides. 231 232 To compare the differences of the microbial samples in the different columns, principal 233 component analysis (PCA) were performed on the basis of the differences in carbon source utilization patterns (CSUPs). All the CSUPs data were processed by Taylor transformation after 234 235 the assessment of normality, homoscedasticity and linear correlations [48]. One-way 236 permutational analysis of variance (PERMANOVA) with both Bray-Curtis and Euclidean distance 237 was employed to assess the differences in microbial community metabolic profiles among the columns in the PCA plots. PERMANOVA analysis was carried out using the free paleontological 238 239 statistic software package (PAST) [51]. The relationships between CLPP (AWCD, richness and guild utilization) and the different environmental variables (water temperature, EC, DO, pH, TOC, TN, 240 NH₄⁺-N, TP, ibuprofen, iohexol, tebuconazole and imazalil removal and substrate concentrations 241 242 of ibuprofen, iohexol, tebuconazole and imazalil) were analyzed by canonical correlation analysis 243 (CCorA). This approach was further complemented with Pearson's correlation analysis to test 244 which correlations from the CCA were significant (p < 0.05) [52]. Within the significant results, the correlation coefficient r was interpreted as: strong correlation ($r \ge |0.7|$) and a moderate 245 246 correlation ($|0.5| \le r < |0.7|$) [53, 54]. One-way Analysis of variance (ANOVA) and post hoc Tukey's HSD test were used to assess differences among columns at 95% confidences level (*p*<0.05). One-way ANOVA, PCA and CCorA were conducted using the XLSTAT Pro[®] statistical software (XLSTAT, Paris, France).

250

251 **3. Results**

252 3.1. Batch adsorption

253 The adsorption isotherms, expressed as the amount of each compound adsorbed onto 254 materials at equilibrium (q_e , $\mu g/g$) versus the amount of compound remaining in solution (C_e , μ g/L) is shown in Fig. 1. The adsorption of ibuprofen, tebuconazole and imazalil onto the six 255 256 materials were described by both Freundlich and Langmuir isotherm models with good correlation coefficients (Table 2). As exceptions, ibuprofen did not fit for sand, while iohexol did 257 258 not fit for sand, polonite and CAAC. Regarding ibuprofen, petcoke had the highest adsorption (97 259 $\mu g/g$ of q_m), followed by zeolite (78 $\mu g/g$ of q_m). The q_m for blast iron slag, polonite and CAAC was 260 between 9 and 24 µg/g for ibuprofen. Sand nearly did not adsorb ibuprofen. As for iohexol, only 261 three materials exhibited adsorption, petcoke (84 μ g/g of q_m) > zeolite (57 μ g/g of q_m) > blast iron slag (33 μ g/g of q_m). Petcoke was also the material with the highest adsorption of tebuconazole 262 $(217 \ \mu g/g \text{ of } q_m)$, followed by CAAC (83 $\mu g/g \text{ of } q_m)$ and blast iron slag (53 $\mu g/g \text{ of } q_m)$. The sand, 263 zeolite and polonite had similar q_m for tebuconazole, ranging from 15 to 24 μg/g. For imazalil, the 264 265 petcoke also had the highest adsorption (205 µg/g of qm). Polonite and CAAC had similar qm (81-266 83 μ g/g), which was higher than that of the blast iron slag (60-63 μ g/g). Comparing the adsorption of pharmaceuticals and pesticides, all the materials in general showed higher adsorption of the 267 pesticides (tebuconazole and imazalil) than of the pharmaceuticals (ibuprofen and iohexol). 268



Fig. 1. Equilibrium isotherms for adsorption of ibuporofen, iohexol, tebuconazole and imazalil
onto the blast iron slag, petcoke, polonite, tobermorite sand and zeolite. Comparison between
the experimental data (points) and predictions of Freundlich and Langmuir models (lines).

Table 2. Fitting parameters of Freundlich and Langmuir models to the experimental data (n=2).

Adsorbate	Adsorbent	t Freundlich		h	Langmuir		Adsorbate	Adsorbent	Freundlich			Langmuir			
		K _f	1/n	R ²	q _m	Kı	R ²			K _f	1/n	R ²	q _m	Kı	R ²
					(µg/g)								(µg/g)		
Ibuprofen	Blast iron	5.76	0.32	0.8507	11	0.25	0.8446	Tebuconazole	Blast iron	6.40	0.68	0.9900	53	0.12	0.9884
	slag								slag						
	Petcoke	25.82	0.54	0.9689	97	0.47	0.9556		Petcoke	57.24	0.54	0.9101	217	0.44	0.8957
	Polonite	1.49	0.66	0.7427	9	0.036	0.7272		Polonite	5.12	0.45	0.8242	15	0.79	0.8237
	CAAC	7.78	0.47	0.9371	24	0.66	0.9115		CAAC	18.95	0.54	0.9858	83	0.31	0.9640
	Sand	-	-	-	-	-	-		Sand	2.83	0.63	0.8752	17	0.22	0.8717
	Zeolite	14.26	0.69	0.9376	78	0.26	0.9730		Zeolite	7.04	0.49	0.9532	24	0.50	0.9485
Iohexol	Blast iron	0.13	0.59	0.9821	33	0.00030	0.9848	Imazalil	Blast iron	31.52	0.36	0.9699	63	0.44	0.9345
	slag								slag						
	Petcoke	20.43	0.57	0.9168	84	0.42	0.9114		Petcoke	51.3106	0.56	0.9382	205	0.47	0.9329
	Polonite	-	-	-	-	-	-		Polonite	14.2642	0.62	0.9666	83	0.21	0.9777
	CAAC	-	-	-	-	-	-		CAAC	23.0481	0.49	0.9725	81	0.0012	0.9448
	Sand	-	-	-	-	-	-		Sand	4.55	0.63	0.8869	29	0.18	0.8851
	Zeolite	8.23	0.66	0.9854	57	0.17	0.9880		Zeolite	15.42	0.65	0.9761	60	0.18	0.9828

278 **3.2. Column performance**

279 3.2.1. Removal of OMPs in the columns

280 At day 66, ibuprofen removal was similar among all the columns, ranging from 27-48%, (Fig. 2). Ibuprofen removal significantly increased during time (from day 25 to 66) in the sand+iron, 281 282 zeolite and sand columns (p<0.05). The iohexol removal in the sand+iron, petcoke and CAAC 283 columns (96-98%) was in general higher than in the zeolite, sand and polonite columns (86-89%) at day 66 (Fig. 2). The iohexol removal at day 66 was significantly higher than that at day 25 in all 284 285 the columns (p<0.05). Regarding tebuconazole at day 66 (Fig. 2), the removal in the petcoke columns (85%) was significantly higher than in the remaining columns (42-55%) (p<0.05). 286 Compared to day 25, we also found that tebuconazole removal significantly increased during time 287 (to day 66) (p<0.05). As for imazalil removal at day 66 (Fig. 2), it was highest in the sand+iron and 288 petcoke columns (94-98%), followed by the sand and CAAC columns (both 80%), which was 289 290 significantly higher than in the zeolite columns (42%) at day 66 (p<0.05). Imazalil removal was 291 lowest in the polonite columns (59%). Compared to day 25, all the columns had significantly 292 increased imazalil removal at day 66 except for the polonite columns (p<0.05). In addition, 293 compared the removal efficiencies among the four OMPs in all the columns, it generally followed an order of iohexol > imazalil > tebuconazole > ibuprofen. 294



Fig. 2. The removal of ibuprofen, iohexol, tebuconazole and imazalil in each type of columns along with time. Different letters indicate significant differences between columns (p<0.05). Asterisk above the bars indicate significant differences between day 25 and day 66 for each type of column (p<0.05).

300

The sorption of the four OMPs to the column substrates was measured at the end of experiment (day 66) (Table 3). The sorbed ibuprofen in the sand+iron, zeolite and petcoke columns (0.11-0.22 μ g/g) was similar, which was significantly higher than in the CAAC columns (0.06 μ g/g). Ibuprofen was not detected in the substrates of the sand and polonite columns. Iohexol was not detected in any of the columns except for petcoke columns (0.08 μ g/g). Regarding tebuconazole, its concentration in the substrate of the CAAC columns was the highest (0.99 μ g/g). The tebuconazole concentration in the substrates of the zeolite and petcoke columns 308 (0.50-0.51µg/g) was higher than that in the sand+iron and polonite columns (0.31-0.32 µg/g). The 309 tebuconazole concentration in the substrate of the sand columns was the lowest (0.18 µg/g DW 310 substrate). Like for tebuconazole, the highest imazalil concentration in substrate was found in 311 the CAAC columns (1.26 µg/g), followed by the zeolite and petcoke columns (0.42-0.59 µg/g DW 312 substrate). The sand+iron, sand and polonite columns had the lowest imazalil sorption (0.27-0.31 313 µg/g). The amount of sorbed tebuconazole and imazalil in all the columns was in general higher 314 than that of ibuprofen and iohexol (Table 3).

A comparison between the maximum mass adsorbed per mass of adsorbent (qm) of the 315 four OMPs found in batch tests with the theoretical substrate concentrations calculated based in 316 317 the input mass of the four OMPs in each type of columns was performed (Table 3). The theoretical 318 substrate concentrations values are much lower than the respective q_m , except for the cases 319 where qm could not be determined (no adsorption of ibuprofen and iohexol onto sand, and no 320 adsorption of iohexol onto polonite and CAAC). Furthermore, the percentage of the sorbed ibuprofen, iohexol, tebuconazole and imazalil in comparison with the total input mass of each 321 322 compound was calculated. The present results show that the amount of ibuprofen, iohexol, tebuconazole and imazalil sorbed by the substrates in all the columns were <13%, <36%, <31% 323 324 and <31% of the total input of the respective compound, respectively (Table 3).

Table 3. The maximum mass adsorbed per mass of adsorbent of each material, the theoretical and measured
 substrate concentration, and the percentages of ibuprofen, iohexol, tebuconazole and imazalil removed by
 the substrates in the columns to the total input (mean ± SD, n=3).

Organic Mi	cro-pollutants						
		Sand+	Zeolite	Sand	Polonite	Petcoke	CAAC
		Iron ^g					
Ibuprofen	q _m	1	78	-	9	97	24

	Theoretical	0.86	0.29	2.02	1.67	1.66	3.15
	concentration [#]						
	Measured	0.11±0.01ª	0.16±0.07ª	ND	ND	0.22±0.04ª	0.06±0.01 ^b
	concentration						
	Percentage (%)	12.7±0.9ª	7.8+3.2 ^{ab}	< 4.2±0.3 ^{b,*}	< 2.2±0.3 ^{c,*}	13.1±2.4ª	2.0±0.3 ^c
Iohexol	q _m	3	57	-	-	84	-
	Theoretical	0.12	0.29	0.15	0.24	0.24	0.45
	concentration#						
	Measured	ND	ND	ND	ND	0.08±0.02	ND
	concentration						
	Percentage (%)	<35.7±2.5 ^{a,*}	<13.1±0.3 ^{b,*}	< 30±1.9 ^{c,*}	<15.5±1.0 ^{d,*}	33.2±7.0 ^{ac}	<8.7±0.3 ^e
Tebuconazole	q _m	20	24	17	15	217	83
	Theoretical	1.00	2.34	1.20	1.94	1.93	3.65
	concentration#						
	Measured	0.31±0.04 ^b	0.50±0.04ª	0.18±0.03 ^c	0.32±0.12 ^{abc}	0.51±0.21 ^{abc}	0.99±0.08 ^d
	concentration						
	Percentage (%)	30.8±4.4ª	21.2±1.6 ^b	15.2±2.2°	16.4±6.0 ^{bc}	26.3±10.9 ^{abc}	27.2±2.6ª
Imazalil	q _m	32	60	29	83	205	81
	Theoretical	1.03	2.40	1.20	1.99	1.97	3.74
	concentration#						
	Measured	0.31±0.07ª	0.59±0.11 ^b	0.29±0.02 ^a	0.27±0.05 ^a	0.42±0.14 ^{ab}	1.26±0.09 ^c
	concentration						
	Percentage (%)	30.5±6.5 ^{ac}	24.7±4.5ª	23.3±1.3ª	13.7±2.4 ^b	21.4±7.4 ^{bc}	33.7±3.2ª

#: The theoretical concentration of ibuprofen, iohexol, tebuconzaole and imazalil were calculated based on the assumption that
 the total input mass of the four OMPs were completely adsorbed by the respective materials. The unit for theoretical and
 measured concentration is μg/g DW substrate (DW: dry weight). g: the q_m in the sand+iron columns was calculated based on the
 weight ratio of sand to blast iron slag (10:1). Different letters indicate significant differences between columns (p<0.05). Asterisk
 represents the the percentages of adsorbed compounds calculated based on limits of detection. ND: not detectable.

333

334 **3.2.2. Conventional water parameters**

In order to pollutant removal processes occurred in the column, we further analyzed the conventional water parameters. The effluent temperature during the two sampling campaigns ranged from 14 °C to 17 °C (Table S2). At day 66, the pH in the zeolite (9.8) and polonite (10.0) columns was significantly higher than in the other columns [CAAC (8.7), sand (8.1), petcoke (7.9) and sand+iron (7.9)] (Table S2). The EC ranged between highest values in the zeolite columns (1548 μ S/cm) and lowest in the polonite columns (645 μ S/cm). The DO in all the columns ranged from 0.6 to 3.0 mg/L. At the intermediate sampling (day 25), the differences between columns for pH and EC were already visible (Table S2).

At day 66, the removal of COD tended to be higher in the petcoke (50%), sand+iron (48%) 343 344 and CAAC (40%) columns than in the zeolite (31%), sand (28%) and polonite (26%) columns (Fig. S3). The petcoke columns had the highest TOC removal (59%). The sand+iron, polonite and CAAC 345 346 columns had similar TOC removal (47-49%), which were generally higher than the sand (42%) and zeolite (32%) columns. As for BOD removal, all the columns had similar removal (55-69%) except 347 348 the polonite columns (45%). Compared day 66 to day 25, significantly increased removal was only observed for COD and TOC removal in the sand+iron and zeolite columns (p<0.05). Nitrate was 349 350 barely observed in the influent and effluent at day 66 and also 25 (Table S2). At day 66, the 351 removal of NH₄⁺-N in the zeolite columns was significantly higher than that in the remaining 352 columns (-13%-21%) (p<0.05). TN removal had a similar trend with NH₄⁺-N removal. Compared 353 day 66 to day 25, TN and NH4⁺-N removal was significantly lower in all columns except for zeolite 354 columns (p<0.05). The zeolite columns were the only that had a significantly higher TN and NH₄⁺⁻ 355 N removal at day 66 than day 25 (p<0.05). For total phosphorous at day 66, the sand+iron, 356 polonite and petcoke columns had similar removal (98-99%), higher than that observed in the sand and CAAC columns (68-71%). Zeolite did not remove TP (-61%). Similar trends among 357 columns to those of day 66 were observed for TP removal at day 25. Moreover, TP removal was 358 similar between day 66 and day 25. 359

361 **3.2.3.** Metabolic profiles of interstitial water and biofilm microbial communities

362 In the interstitial water (Fig. 3a), sand+iron, sand and CAAC columns had similar microbial 363 activity and metabolic richness, followed by petcoke columns. Zeolite and polonite columns had 364 the lowest microbial activity and metabolic richness (Fig. 3a). According to the carbon source 365 utilization patterns (CSUPs) of the microbial communities in the interstitial water samples by PCA analysis, three distinct groups (Fig. 4a) were identified: 1) sand+iron columns grouped with CAAC 366 columns, 2) zeolite columns grouped with polonite columns, and 3) sand columns grouped with 367 petcoke columns. The carbon source utilization (guilds) among the different column types (Fig. 368 369 S4) did not reveal any clear trend.

370 In the biofilm (Fig. 3b), the sand columns had the highest microbial activity and metabolic richness. Sand+iron, petcoke and CAAC columns had similar microbial activity and metabolic 371 372 richness, which was higher than that of polonite columns. Zeolite had the lowest microbial 373 activity and metabolic richness. The carbon source utilization patterns (CSUPs) of the microbial 374 communities in the biofilm samples were further analyzed using a PCA ordination (Fig. 3b). Two distinct groups were determined with the exception of an independent point for the zeolite 375 376 columns: (1) petcoke columns were grouped with CAAC columns; (2) sand+iron, sand and polonite columns were grouped together. No significant differences or clear trends can be 377 378 gleaned from the carbon source utilization (guilds) among columns types (Fig. S5).





Fig. 3. Microbial activity based on AWCD and metabolic richness for the interstitial water (a) and biofilm (b) samples at day 66 from the sand+iron (SI), zeolite (Z), sand (S), polonite (PI), petcoke (Pt) and CAAC (C) columns. Different letters indicate significant differences between columns (p<0.05) and PCA plot of the microbial community based on the carbon source utilization pattern in the interstitial water (a) and biofilm (b) samples. The first or the first two letters represent the types of column. The last letters (W and B) represent interstitial water and biofilm, respectively. The different groups are significantly different (p<0.05, PERMANOVA).

388 4. Discussion

All the materials can adsorb ibuprofen, iohexol, tebuconazole, and imazalil with the exceptions of sand for ibuprofen and iohexol, and polonite and CAAC for iohexol. Compared to ibuprofen, tebuconazole and imazalil, iohexol was found to be more difficult to adsorb. It may be attributed to the lower logD (around -2.0) (D: distribution-coefficient) of iohexol in the solutions

during the adsorption experiment (Table S4), denoting its higher hydrophilicity. Although the 393 394 similar logKow for ibuprofen (3.97) and tebuconzaole (4.10) and imazalil (3.89) (Table S1), the three OMPs have different logD in the solutions during the adsorption experiment, presenting 395 0.3-1.4, 3.7 and 3.7-3.8, respectively (Table S4). The results of the adsorption experiment are in 396 397 accordance with the hydrophobicity of the tested OMPs, in which tebuconazole and imazalil were more easily adsorbed by the six materials than ibuprofen. To the best of our knowledge, the 398 adsorption of ibuprofen, iohexol, tebuconazole and imazalil onto the sand, zeolite, blast iron slag, 399 400 petcoke, polonite and CAAC has rarely been investigated by adsorption isotherms, especially in a 401 single experiment. However, the adsorption capacities of the six materials ($\mu g/g$ range) are 402 limited in comparison to well-known sorbents, such as activated carbon and carbon nanotubes 403 (at mg/g range) [55-58]. The matrix in CWs is characterized by being low-cost and locally available [59]. The well-known sorbents are generally also more expensive than the presently studied 404 405 materials, forming a bottleneck for wide application of these high-sorption materials as CWs 406 media. It raises a question whether the materials can play an important role in removing OMPs 407 by enhancing microbial degradation in CWs when their sorption capacities are limited.

Regarding the removal of OMPs in the columns, compared the q_m to the theoretical concentrations of the four OMPs (Table 2), the total input mass of the four OMPs should have been retained in the columns and OMPs in the effluent should be negligible, if only sorption processes had occurred in all the columns. However, the four OMPs were still present in the effluent of all the columns, indicating that the columns were unable to completely remove the OMPs from wastewater. The incomplete removal of the four OMPs by sorption process could be attributed to competitive sorption phenomenon as the OMPs and other pollutants (such as

organics, TN and TP) may compete for preferred binding sites on the materials [60, 61]. In 415 416 addition, it is known that biofilm development decreases the sorption capacity of the packing materials. The percentages of the substrate concentrations of the four OMPs to the total mass 417 418 of the respective OMPs only represent 0-36% of the expected sorption. Thus, the present results 419 indicate that OMPs biodegradation also occurred in all the columns, which is also supported by the removal of organic matter in the columns. Although not always significant, organic removal 420 (COD, TOC, and BOD) in all the columns increased over time (day 25 to 66), suggesting organic 421 422 biodegradation likely occurred. The evidence of biodegradation process is stronger for ibuprofen 423 and iohexol removal, for which OMP sorption was found to be negligible in the batch tests. Although ibuprofen was not detected in the substrates of the sand and polonite columns, and 424 425 Iohexol was also not detected in all the columns except for petcoke columns, the ibuprofen and 426 iohexol were removed from wastewater. Additionally, the low Ibuprofen removal (27-48%) at 427 day 66 in all columns may be due to the fact that ibuprofen is difficult to degrade under anaerobic conditions [62]. Zwiener and Frimmel [63] reported that ibuprofen is easily degraded under oxic 428 429 conditions and poorly under anaerobic conditions in biofilm reactors. Iohexol, tebuconazole and imazlil achieved high removal efficiencies at the end of experiment, indicating that the three 430 OMPs also can easily degraded under anaerobic conditions. It should be noted that iohexol 431 432 removal was more than 86% in all columns, and it is generally considered as a recalcitrant OMP 433 [64]. Thus, the high removal of iohexol in this study reveals that iohexol may be degraded to a larger extent under anaerobic conditions, which warrants further investigation. 434

435 When comparing the removal of OMPs among the different columns, the removal of the 436 respective OMPs were generally similar at the end of experiment, even though the six materials

437 had different sorption capacities. For instance, sand had the lowest sorption capacity compared 438 to other materials (Table 1), but the removal of ibuprofen, iohexol and tebuconazole in the sand columns was similar to the other five types of columns, with the exception of higher tebuconazole 439 440 removal in the petcoke columns. The removal of imazalil in the sand columns was similar with 441 that in the CAAC columns, and even higher than that in the zeolite and polonite columns. Polonite is a similar case with sand, the isotherms revealed a limited sorption capacity, but in the packed 442 columns polonite provided similar removal with the other materials. These results indicate that 443 the tested materials can facilitate OMP removal during wastewater treatment, although the 444 sorption capacities of the six materials were limited. We hypothesized that the sorption is 445 reversible, but the sorption interactions slow the OMPs down in the system providing longer 446 447 contact time with the biofilm so that biodegradation could occur to larger extent.

The microbial community functional profiles in the interstitial water and biofilm 448 449 samples in the six types of columns were further elucidated. To the best of our knowledge, the 450 comparisons of microbial community functional profiles in the interstitial water and biofilm 451 among the different material-packed columns have not been studied before. At the starting 452 period of the column operation, the interstitial water microbial community is known to depend 453 on the microbial communities in the influent [41]. At day 66, the microbial community functional 454 profiles in the interstitial water were different among the different columns, indicating that materials themselves could shape the interstitial water microbial communities even though all 455 456 the columns were fed with the same wastewater. In addition, the microbial community functional profiles in the interstitial water samples of all the columns, except for CAAC, were 457 458 different from that in the influent (Fig. S6). This results are in accordance with the finding by Lv

et al. [65], who found the microbial community in the unplanted mesocosms (sand as the matrix) 459 460 was different from that in the influent. Furthermore, Weber and Legge [41] observed that, in CW mesocosms (gravel as the matrix), after a state of initial difference based on the different 461 462 inoculum community profiles (0-6 days) (different wastewater feeds), the community starts 463 shifting towards a period where the interstitial water microbial community functional profiles in all the mesocosms were similar (days 25-73) before an equilibrium state with unplanted and 464 planted-mesocosm CLPP groupings (after 74 days). However, in the present study, although the 465 466 columns were operated for 66 days, the microbial community functional profiles of the six types 467 of columns were different. This difference can only be explained by the columns packed with different materials. 468

In the biofilm samples, we also observed three different CLPP groupings among the six 469 types of columns, revealing that the different materials also shaped the microbial community 470 471 functional profiles in the biofilm developed during the 66 days in the columns. When comparing 472 the microbial community functional profiles between the interstitial water and biofilm, we found 473 that the microbial activity and metabolic richness were significantly higher in the biofilm than in 474 the interstitial water in all the columns, and the microbial metabolic function profiles of the 475 interstitial water and biofilm were different (Fig. S7). This trend was also observed by Zhang et al. 476 [43] and Weber and Legge [66]. Furthermore, the CLPP grouping patterns in the interstitial water were different from that in the biofilm (see Fig. 3). The present results indicate that, in addition 477 to sand, the other materials as CW matrix also can shape interstitial water and biofilm 478 communities in different directions. Calheiros et al. [67] found that the microbial communities in 479 480 the CWs with expanded clay aggregates and fine gravel for industrial wastewater treatment were

different. Guan *et al.* [68] also observed that bacterial community diversity and structures had pronounced differences between sand and zeolite CWs treating polluted river water. Different materials influenced the establishment and growth of biofilm and microbial community as well as the treatment performance of CWs [69]. The differences in microbial community structures of columns may be attributed to the different characteristics of different materials, such as pH, EC, porosity, organic matter content and solid surface [67, 70].

When studying the correlation between water metrics (temperature, EC, pH, DO, COD, 487 TOC, BOD, TN, NH₄⁺-N, TP, four OMP removal, and substrate concentrations of the four OMPs) 488 489 and microbial metrics (AWCD, richness, and guilds utilization) by CCorA analysis (Table S5 and S6). 490 A larger number of correlations between guild utilization and ibuprofen removal were observed in the biofilm than in the interstitial water, indicating a more crucial role for biofilm microbial 491 492 communities in pollutant removal than interstitial water microbial communities. The present 493 result is consistent with the findings by Lv et al. [44] and Zhang et al. [43], whom reported that 494 biofilm communities in CWs had an important role in tebuconzaole and ibuprofen biodegradation. 495 Kurzbaum et al. [71] also found that higher total cultivable bacteria and specific phenol-degrading 496 bacteria in gravel attached biofilm than in free water when studying the different CW 497 components to understand the relative contribution for phenol biodegradation. In addition, we 498 also observed that pH and DO were the main variables connecting with biofilm communities (Table S6). In examining the pollutants removal results, generally, microbial activity, and 499 500 metabolic richness were determined as the key factors influencing OMPs biodegradation in the columns. 501

To sum up, the presence of materials influences microbial community structures and 502 503 the treatment performance of columns. Even using materials with low adsorption capacities as substrates in biological filters, they can also play an important role in the removal of OMPs. 504 505 Therefore, an appropriate selection of materials for biological filters can improve system's 506 performance via shaping microbial community. However, it is still unknown which material is superior to enrich the functional groups of microbial community for removing OMPs. Moreover, 507 it is well known that plant presence also affect the microbial community structures in CWs [42, 508 509 43, 72-74]. It is yet unclear which effect (material, plant or design) would be greater when using the different tested materials. Such microbial drivers need to be further studied. 510

511

512 5. Conclusions

The adsorption capacities of six representative materials (sand, zeolite, blast iron slag, 513 514 petcoke, polonite and CAAC) to the selected OMPs (ibuprofen, iohexol, tebuconazole and imazalil) in the µg/g range were not promising compared to well-known sorbents, such as activated carbon 515 516 and carbon nanotubes (mg/g). It indicates that none of the tested materials should be especially 517 applied to enhance OMPs by sorption. In the medium-term column study (66 days), the removal 518 of the four OMPs increased with time, during which biodegradation was the main pathway for 519 removal. Although the different materials had different sorption capacities, the removal of the 520 four OMPs in the six types of columns was generally similar. In addition, the CLPP analysis showed that the materials shaped the functionality of microbial communities in interstitial water and 521 biofilm in different directions. Different materials had different biofilms communities with 522

523	distinct functions that resulted in similar removal among columns. Thus, the common materials
524	as CW matrix can also play a key role in OMP removal by strengthening biodegradation processes.
525	However, we presently cannot conclude which material may shape a better microbial community
526	for OMPs removal/degradation.
527	
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532	
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