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Fc receptors: Cell activators of antibody functions

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ABSTRACT

At the onset of an infection early defense systems, such as complement, get into action. Specialized leukocytes (white blood cells) of the innate immune system, including monocytes, macrophages, and neutrophils also participate as a first line of defense against infections. These early responses are rapid but not very specific and are usually not enough to clear completely many infections. The adaptive immune system is also needed to finish the job against many microorganisms. Antibody molecules, produced during the adaptive immune response, are crucial for preventing recurrent infections. Although, IgG antibodies are essential for controlling infections, these molecules do not directly damage the microorganisms they recognize. Today, it is established that leukocytes of the innate immune system are responsible for the protective effects of these antibodies. IgG molecules bind to their cognate antigens and are in turn recognized by specific receptors (Fcy receptors) on the membrane of leukocytes. Crosslinking these receptors on the surface of leukocytes leads to activation of several effector cell functions. These effector functions are geared toward the destruction of microbial pathogens and the induction of an inflammatory state that is beneficial during infections. However, in autoimmune diseases, antibodies can direct these effector functions against normal tissues and cause severe tissue damage. In recent years, several factors that can modulate the IgG-FcyR interaction have been elucidated. In this review, we describe the main types of Fcy receptors, and our current view of how antibody variants interact with these receptors to initiate different cell responses. In addition, new findings on the signaling role of individual Fcy receptors are also discussed.

Keywords: Immunoglobulin; Antibody; Immunoreceptor; Neutrophil; Macrophage

1. INTRODUCTION

At the onset of an infection by different types of microorganisms, including viruses, bacteria, fungi, and protozoa, early defense systems, such as constitutive expression of antimicrobial peptides, and activation of complement get into action. These systems are rapid but not particularly specific. Specialized leukocytes (white blood cells) of the innate immune system, including monocytes, macrophages, and neutrophils also participate as a first line of defense against infections. These leukocytes can bind some microbial molecules, termed danger- and pathogen-associated molecular patterns (DAMPs and PAMPs, respectively) via numerous receptors such as the Toll-like receptor family [1,2]. In this way, leukocytes recognize microorganisms directly and prevent a massive infection [3]. These early responses however, are usually not enough to clear completely many infections. The adaptive immune system is also needed to finish the job against many microorganisms. Antibody molecules, produced during the adaptive immune response, are crucial for preventing recurrent infections [4]. At the beginning of the adaptive response, antibodies belong to the IgM class. These antibodies present low affinity for microbial antigens, but they can easily activate the classical complement pathway. Complement deposited on microorganisms can induce phagocytosis via complement receptors [5,6], or it can induce bacterial lysis via the formation of the membrane attack complex [7]. At later times of the adaptive response, antibodies belong mainly to the IgG class. These antibodies are of higher affinity and of much greater specificity for their particular antigen. Thus, IgG antibodies are key for controlling many microorganisms, as demonstrated by immunodeficiency disorders,



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with low production of this class of antibodies, in which there is increased susceptibility to microbial infections [4]. Although. IgG antibodies are essential for controlling infections, these molecules do not directly damage the microorganisms they recognize. Today, it is established that leukocytes of the innate immune system are responsible for the protective effects of these antibodies. IgG molecules bind to their cognate antigens via their two fragment antigen-binding (Fab) sites, and are in turn recognized by specific receptors on the membrane of leukocytes. These receptors bind the Fc (fragment crystallizable) domain of IgG; thus, they are named Fcy receptors (FcyR) [8,9]. In this way, IgG antibodies are the bridge between the two arms of the immune system. bringing together the specificity of recognition of the adaptive immune system and the destructive potential of the cells of the innate immune system. Crosslinking these receptors on the surface of leukocytes leads to activation of several effector cell functions. Depending on the cell type, and also on the $Fc\gamma$ receptor type, these functions include phagocytosis, cell degranulation, production of various cytokines and chemokines, antibody-dependent cell-mediated cytotoxicity (ADCC), and activation of genes [10]. These effector functions are geared toward the destruction of microbial pathogens and the induction of an inflammatory state that is beneficial during infections. However, in autoimmune diseases, antibodies can direct these effector functions against normal tissues and cause severe tissue damage [11,12]. It is then of great interest to understand how various FcyR are activated to induce these cellular functions. In recent years, several factors that can modulate the IgG-FcyR interaction have been described. These factors include, the particular IgG subclass [13] and the glycosylation pattern of the antibody [14]. In addition, other molecules, such as members of the pentraxin family can bind FcyR [15,16], and certain glycosylation variants of antibodies can bind other cell membrane receptors different from FcyR [17-19]. In this review, we describe the main types of Fcy receptors, and our current view of how antibody variants interact with these receptors to initiate different cell responses. In addition, new findings on the signaling role of individual Fcy receptors are also discussed.

2. Fcy RECEPTORS

2.1. Structure

Antibodies represent an important bridge between the specificity of the adaptive immune system, and the highly destructive mechanisms of cells of the innate immune system. Antibodies bind to microorganisms via their antigen-binding sites, and to Fc receptors on the surface of leukocytes, via their carboxyl terminal Fc portion. Receptors for the Fc portion of various immunoglobulin (Ig) classes have been described [20]. Fc Receptors for IgG (Fc γ R), for IgE (Fc ϵ R), and for IgA (Fc α R) are known [20]. Crosslinking of Fc γ receptors with their IgG antibody ligands triggers various functions in many cells of the immune system. These cell functions include phagocytosis, cell degranulation, production of various cytokines and chemokines, ADCC, and activation of genes [10,21].

Fc γ receptors are a family of glycoproteins, part of the IgG superfamily. They consist of an IgG binding α -subunit, that usually pairs with accessory γ chains, which are important for receptor signaling (**Figure 1**).

In humans, three classes of FcyR have been identified, FcyRI (CD64), FcyRII (CD32), and FcyRIII (CD16) (**Figure 1A**) [22]. FcyR are coded for by different genes and differ in their relative avidity for IgG, molecular structure, and cellular distribution. FcyRI α -subunit presents three Ig-like extracellular domains, and binds monomeric IgG [23]. In contrast, FcyRII and FcyRIII present two Ig-like extracellular domains, and bind only multimeric immune complexes. FcyRI, expressed on monocytes, macrophages, and interferon- γ (IFN- γ)-stimulated neu-



Figure 1. Fcy Receptor family. (A) The human family of receptors for the Fc portion of immunoglobulin G (IgG) molecules comprises three members FcyRI, FcyRII, and FcyRIII. The IgG binding α -subunit in the high affinity FcyRI, possesses three immunoglobulin (Ig)-like extracellular domains. The α subunit in the other low-affinity receptors presents only two Ig-like domains. Activating receptors contain an ITAM (immunoreceptor tyrosine-based activation motif) sequence within the α subunit (for FcyRIIA) or within the accessory γ and ζ chains. In contrast, FcyRIIB is an inhibitory receptor containing an ITIM (immunoreceptor tyrosine-based inhibition motif) sequence. FcyRIIIB is also an activating receptor, which is bound to the cell membrane via a glycosylphosphatidylinositol (GPI) anchor; (B) The murine family of receptors for the Fc portion of immunoglobulin G molecules comprises four members FcyRI, FcyRIIB, FcyRIII, and FcyRIV. Activating receptors (FcyRI, FcyRIII, and FcyRIV) contain an ITAM sequence within the accessory γ chains; while the inhibitory receptor (FcyRIIB) contains an ITIM sequence.

trophils, is associated with a dimer of the common Fc receptor (FcR) gamma-chain (also named FcRy chain). Each γ chain contains tyrosine residues that are phosphorylated upon receptor activation and become docking sites for other signaling molecules. These tyrosine residues are found within a common motif known as ITAM, for immunoreceptor tyrosine-based activation motif [24, 25]. There are several isoforms of FcyRII, derived from its three genes and from alternative splicing. FcyRII isoforms are distributed differently on hematopoietic cells. FcyRIIA and FcyRIIC are found mainly in phagocytic cells (neutrophils, monocytes, and macrophages), whereas FcyRIIB is expressed mainly in B lymphocytes [22]. FcyRIIB expression is inducible in phagocytic leukocytes, for the negative regulation of cell functions, such as phagocytosis [8,26]. The human FcyRIIA is a particular receptor that does not have associated FcRy chains. FcyRIIA contains an ITAM in its cytoplasmic portion, while FcyRIIB has a different tyrosine-containing motif involved in negative signaling. This motif is known as ITIM, for immunoreceptor tyrosine-based inhibition motif [27]. FcyRIII has two isoforms: FcyRIIIA is a receptor with a transmembrane portion and a cytoplasmic tail, associated with an ITAM-containing homodimer of FcRy chains on macrophages, natural killer (NK) cells, and dendritic cells. FcyRIIIA expressed on basophils and mast cells associates with a heterodimer of γ/ζ chains and an extra β chain (Figure 1A) [22,23]. FcyRIIIB is present exclusively on neutrophils and it is a glycosylphosphatidylinositol (GPI)-linked receptor, lacking a cytoplasmic tail. No other subunits are known to associate with it, and its signaling mechanism remains unidentified (Figure 1A). It is also worth noting that human FcyRIIA and FcyRIIIB are exclusive receptors that are not found in other species [28].

In mice, several Fcy receptors have also been described (Figure 1B). These receptors are very similar, yet not identical, to the human FcyR [8]. FcyRI is expressed on monocytes and macrophages, and it is also associated with a dimer of $FcR\gamma$ chains, which contain the ITAM motifs involved in receptor signaling. FcyRIII is a receptor with a transmembrane portion and a cytoplasmic tail, associated with a dimer of FcRy chains, containing ITAMs. This receptor is closer to the human FcyRIIA, as revealed by the genetic structure of FcRs in various species [13]. FcyRIV is also an activating receptor expressed together with a dimer of FcRy chains [29], and it is closer to the human FcyRIIIA (Figure 1B). FcyRIV may be the most relevant activating FcyR in mice, due to its ability to bind IgG2a and IgG2b with higher affinity [13]. FcyRIIB is the negative receptor containing an ITIM motif in its cytoplasmic tail. It is expressed mainly in B lymphocytes but also in phagocytic leukocytes and dendritic cells. FcyRIIB, described first in B lymphocytes, down regulates the activation signals from the B cell antigen receptor (BCR) to inhibit antibody production by the B cell [30]. This inhibitory receptor also helps to regulate initiation of other cell functions in phagocytic leukocytes and dendritic cells by creating, together with activating Fcy receptors, a threshold for cell activation [13,31].

2.2. Cell Expression of Fcy Receptors

Fcy receptors are found on many cells of the immune system, including granulocytes such as neutrophils and eosinophils; phagocytes such as neutrophils, monocytes and macrophages; and lymphocytes such as natural killer cells and B cells [23]. The wide variety of cellular responses regulated by Fcy receptors is consequently not surprising. Monocytes and macrophages express all types of activating Fcy receptors, FcyRI, FcyRII, and FcyRIII in humans, and FcyRI, FcyRIII, and FcyRIV in mice (Table 1). Murine neutrophils express FcyRIII, and FcyRIV, whereas human neutrophils express FcyRIIA and FcyRIIIB. It is noteworthy to mention that human neutrophils have the two unique $Fc\gamma R$, not present in neutrophils of other species. Thus, care should be taken when analyzing data derived from mouse studies of neutrophil FcyR function, since conclusions may not necessarily apply to human neutrophils. NK cells exclusively express FcyRIIIA in humans and FcyRIII in mice. Dendritic cells (DCs) also express various FcyRs, while B lymphocytes express mainly the inhibitory FcyRIIB (Table 1). Some T lymphocyte populations have also been reported to express FcyRs [32-34], but the role for these receptors in T cell function or development remains unclear and requires further studies.

2.3. Fcy Receptor Signaling

Crosslinking of activating Fc γ receptors with their IgG antibody ligands triggers various functions in many cells of the immune system. As mentioned before, all activateing receptors contain ITAM motifs involved in receptor signaling. The exact activation mechanism is not completely clear, but at the initial molecular events involve activation of Src family kinases followed by activation of Syk (spleen tyrosine kinase) family kinases.

The model for the initial steps of activating Fc γ R signaling is as follows: Upon crosslinking, the receptor associates with lipid rafts. Lipid rafts are small regions of the plasma membrane that are enriched in cholesterol and sphingolipids [35]. There, the receptor co-localizes with Src kinases. These kinases phosphorylate tyrosines within the ITAM. Phosphorylated tyrosines then become docking sites for Syk. This kinase then phosphorylates multiple substrates, including phosphatidylinositol 3-kinase (PI 3-K), phospholipase C γ (PLC γ), and the adap-

Human			
Receptor	Principal antibody ligand	Affinity for ligand ^a	Cell distribution
FcyRI (CD64)	IgG1 and IgG3 > IgG4 > IgG2	$\begin{array}{c} High \\ (Kd \sim 10^8 \ M^{-1}) \end{array}$	Macrophages Neutrophils Eosinophils Dendritic cells
FcyRIIA (CD32)	IgG1 > IgG2 and IgG3 > IgG4	$\begin{array}{c} Low \\ (Kd \sim 2 \times 10^6 \ M^{-1}) \end{array}$	Macrophages Neutrophils Mast cells Eosinophils Platelets Dendritic cells
FcyRIIB (CD32)	IgG1 > IgG2 and IgG3 > IgG4	$Low \\ (Kd \sim 2 \times 10^6 \text{ M}^{-1})$	Macrophages Neutrophils Mast cells Eosinophils Dendritic cells B Cells
FcyRIIIA (CD16A)	IgG1 and IgG3	$\begin{array}{c} Low \\ (Kd \sim 5 \times 10^5 M^{-1}) \end{array}$	Macrophages Mast cells Basophils NK cells Dendritic cells
FcyRIIIB (CD16B)	IgG and IgG3	$\frac{Low}{(Kd \sim 2 \times 10^5 \text{ M}^{-1})}$	Neutrophils
Mouse			
Receptor	Principal antibody ligand	Affinity for ligand ^b	Cell distribution
FcyRI	IgG2a	$\begin{array}{c} \text{High} \\ \text{Kd} \sim 1.6 \times 10^8 \text{ M}^{-1} \end{array}$	Monocytes Macrophages Dendritic cells
FcyRIIB	lgG1 IgG2a IgG2b	$\begin{array}{c} Low \\ Kd \sim 3.3 \times 10^6 \ M^{-1} \\ Kd \sim 0.4 \times 10^6 \ M^{-1} \\ Kd \sim 2.2 \times 10^6 \ M^{-1} \end{array}$	B Cells Dendritic cells
FcyRIII	lgG1 IgG2a IgG2b	$\begin{array}{c} Low \\ Kd \sim 0.3 \times 10^6 \ M^{-1} \\ Kd \sim 0.7 \times 10^6 \ M^{-1} \\ Kd \sim 0.6 \times 10^6 \ M^{-1} \end{array}$	Monocytes Macrophages Neutrophils NK cells Dendritic cells
FcyRIV	IgG2a IgG2b	$\begin{array}{c} Low \\ Kd \sim 2.9 \times 10^7 \ M^{-1} \\ Kd \sim 1.7 \times 10^7 \ M^{-1} \end{array}$	Monocytes Macrophages Neutrophils Dendritic cells

Table 1. Fc gamma receptors.

^aData from [64,65]; ^bData from [29,61].

or molecules SLP76 (SH2-domain-containing leukocyte protein of 76 kDa) and LAT (linker for activation of T cells) [36] (Figure 2). These molecules organize and activate several signaling pathways, depending on the cell type, leading to particular cellular responses and to transcriptional changes (Figure 2). For example, SLP76 seems to be important for FcyR signaling in neutrophils [37], while it seems to be dispensable in macrophages and NK cells [38,39]. LAT is another adaptor that seems to participate in many FcyR-mediated functions (Figure 2). LAT was reported to be constitutively associated with the common FcRy chain in monocytes [40], and to be important for efficient phagocytosis in macrophages [40]. Clearly, further studies are needed to identify the particular adaptor used by each type of Fc receptor in the various leukocytes, and to connect this to a unique cell

response.

The inhibitory FcyRIIB is the negative receptor containing an ITIM motif in its cytoplasmic tail instead of an ITAM sequence [41]. It was described first in B lymphocytes, where it down regulates the activation signals from the B cell antigen receptor (BCR) (**Figure 3A**) to inhibit antibody production by the B cell [30]. Contrary to the activating receptors that engage several kinases, this inhibitory receptor signals by activation of phosphatases. Inositol 5-phosphatase (SHIP1) is the main enzyme activated upon crosslinking of FcyRIIB and BCR (**Figure 3B**) [27]. This phosphatase binds via its SH2 domain, to the phosphorylated tyrosines within the ITIM sequence of FcyRIIB. SHIP1 transforms phosphatidylinositol-3,4,5-triphosphate (PIP₃) the main product of PI 3-K, into phosphatidylinositol-3,4-biphosphate (PIP₂); pre-



Figure 2. Activating Fcy receptor signaling. FcyR cross-linking by immunoglobulin (IgG) bound to antigen (Ag), induces activation of Src family kinases and Syk family kinases in lipid rafts. These enzymes associate with phosphorylated tyrosines in the ITAM sequences. Syk then phosphorylates enzymes such as phosphatidylinositol 3-kinase (PI 3-K), phospholipase Cy (PLC γ), and the adaptor molecules SLP76 (SH2-domain-containing leukocyte protein of 76 kDa) and LAT (linker for activation of T cells). PI 3-K produces phosphatidylinositol-3,4,5triphosphate (PIP₃), which leads to activation of Akt and extracellular signal-regulated kinase (ERK). PLCy produces inositoltrisphosphate (IP₃), and diacylglycerol (DAG). These second messengers cause calcium release from the endoplasmic reticulum (ER), and activation of protein kinase C (PKC), respectively. PKC leads to activation of ERK. Vav activates GTPases of the Rho and Rac family, which are involved in the regulation of the actin cytoskeleton. Other enzymes such as Bruton's tyrosine kinase (Btk) also activate the GTPase Rac to induce activation of nuclear factors such as JNK and NF- κ B. P represents a phosphate group. IP₃R, receptor for IP₃.

venting in this way the stimulation of key activating enzymes such as Akt, Btk, and PLC γ (Figure 3B).

2.4. Coexpression of Fcy Receptors and Threshold for Cell Activation

As indicated above, different leukocytes express more than one activating FcyR (Table 1), and most of these cells also express at the same time the inhibitory FcyRIIB. NK cells are particular leukocytes in this respect, because they only express FcyRIIIA in humans and FcyRIII in mice. The coexpression of both activating and inhibitory FcyR results in simultaneous triggering of activating and inhibitory signal transduction pathways (Figure 4A). Thus, a particular cell will respond when the sum of activating and inhibiting signals reach a threshold of activation that is determined by the relative expression of both types of FcyR (Figure 4B). The importance of the inhibitory FcyRIIB in regulating many IgG-mediated responses in different leukocytes was made evident in FcyRIIB-deficient mice, which showed enhanced activity of many IgG-mediated cell responses including: phagocytosis, immune complex-mediated inflammation, IgG-



Figure 3. Inhibitory Fcy RIIB signaling. (A) Upon binding of antigen (Ag) to the B cell antigen receptor (BCR), Src family kinases and Syk family kinases get activated and become associated to the phosphorylated tyrosines in the ITAM sequences of the receptor. Syk then participates in activation of enzymes such as phosphatidylinositol 3-kinase (PI 3-K), Bruton's tyrosine kinase (Btk), and phospholipase $C\gamma$ (PLC γ). Btk leads to activation of Ras and extracellular signal-regulated kinase (ERK), which together with Akt induce cell proliferation. PLCy produces inositoltrisphosphate (IP₃), which causes calcium release, and diacylglycerol (DAG), which causes activation of protein kinase C (PKC). PKC also leads to cell proliferation via ERK; (B) After antigen recognition by both the inhibitory FcyRIIB and the BCR, crosslinking of the receptors leads to phosphorylation of the ITIM sequence. Then, inositol 5-phosphatase (SHIP1) binds to the phosphorylated tyrosines within the ITIM via its SH-2 domain. SHIP1 transforms phosphatidylinositol-3,4,5-triphosphate (PIP₃) the main product of PI 3-K, into phosphatidylinositol-3,4-biphosphate (PIP₂); preventing in this way the stimulation of key activating enzymes (dashed lines). P represents a phosphate group. IP₃R, receptor for IP₃. ER, endoplasmic reticulum.

mediated passive and active anaphylaxis, and IgE-mediated anaphylaxis [42,43]. They also showed enhanced dendritic cell maturation, and antigen presentation [44-46]. These results thus confirmed that FcyRIIB regulates initiation of cell functions by generating, together with activating Fcy receptors, a threshold for cell activation [31, 47].

2.5. Genetic Structure and Polymorphisms

Analysis of FcyR genes in different species has identified



Figure 4. Threshold for activation. (A) The coexpression of both activating and inhibitory $Fc\gamma$ receptors results in simultaneous triggering of activating and inhibitory signaling pathways, which will result in a threshold for activation of a specific cell response such as phagocytosis; (B) A particular cell will respond when the sum of signals from activating receptors (green), and inhibitory receptors (red) reach a threshold of activation that is determined by the relative expression of both types of $Fc\gamma R$.

orthologous receptors between mice and humans. The similarities come from gene localization and also from sequence homology of the extracellular portion of the receptors [8,13]. Therefore, the high affinity receptors FcyRIA and FcRI, the low affinity receptors FcyRIIA and FcyRIII, and also the low affinity receptors FcyRIIIA and FcyRIV cluster in the same area of chromosome 1 (Figure 5). Similarly, the inhibitory FcyRIIB gene maps to the same chromosome region, both in mice and humans (Figure 5). In addition, the human FcyRIIA and mouse FcyRIII, as well as the human FcyRIIIA and mouse FcyRIV, present high sequence homology in their extracellular domains [13]. Despite this similarity, the receptors are not equivalent, since important differences have been detected between mice and human responses to IgG. For example, the human FcyRI binds the IgG1 and IgG3 subclasses with high affinity, while the mouse FcyRI only binds IgG2a with high affinity (Table 1). In addition, the mouse FcyRIV is also able to bind IgE, while the human FcyRIIIA is not [48,49].

In addition, there are several polymorphisms in the human FcyRII and FcyRIII. For FcyRIIA two allelic variants exist expressing either arginine or histidine at position 131 [50,51]. For FcyRIIIA also allelic variants exist expressing either value or phenylalanine at position 158 [52,53]. Similarly, for FcyRIIIB on neutrophils, two isoforms exist, the NA1 and NA2 allotypes [54]. These isoforms differ by five nucleotides and four amino acids, with NA2 containing two additional N-linked glycosylation sites. These differences affect the capacity of FcyRIIIB



Figure 5. Genomic organization of the $Fc\gamma$ -receptor locus for various species. Figure modified from [13]. Data from the ensembl database (<u>http://www.ensembl.org</u>). Cen, centromer.

to interact with human IgG. Therefore, neutrophils from individuals who are homozygous for the NA1 allele have better phagocytosis of IgG-opsonized targets than do neutrophils from NA2-homozygous individuals [55,56]. These multiple Fc γ R and their allelic variants vary greatly in their affinity for different IgG classes [57]. Thus, a great interest exists to understand how different IgG molecules engage different Fc γ R to activate the multiple cell responses associated with antibodies Fc γ receptor signaling.

3. IgG-MEDIATED CELL FUNCTIONS

3.1. Fcy Receptor Affinity for IgG

A single antibody molecule does not bind to Fcy recaptors. However, antigen-antibody complexes promote many low affinity interactions between FcyR on the surface of leukocytes and antibody complexes. The low affinity of antibodies for individual leukocyte FcyR prevents recaptors from binding antibodies in the absence of antigen, thus reducing the chance of immune cell activation when there is not an infection. Immune complexes induce the crosslinking of FcyR to activate the many different antibody-mediated cell responses. Immune complexes are clearly of different kinds, since they are formed by different classes of antibodies, and in vivo studies have suggested that the different IgG classes have different activities. For example, IgG2b/c was better in eliminating B cells [58] and T cell lymphomas [59] than IgG1. Likewise, using class-switch variants of anti-erythrocyte antibodies it was found that IgG2a and IgG2b were better in mediating phagocytosis of opsonized erythrocytes than IgG1 and IgG3 [60,61]. Also, polioencephalomyelitis induced by infection with lactate dehydrogenaseelevating virus (LDV) was delayed much better by IgG2a anti-LDV antibodies than any other IgG class of anti-LDV antibodies [62]. In addition, the severity of glomerular inflammation was greater for IgG2a, followed by

IgG2b and finally IgG1 [63]. All these reports confirmed that different IgG classes mediate different cellular responses *in vivo* through Fcy receptors.

Because, immune complexes of different kinds induce different cell responses, there has been a great interest in determining which type of IgG binds to which FcyR and what particular receptor is involved in mediating the activity of particular IgG classes. Early studies showed that there is a high affinity receptor for IgG (FcyRI), which binds preferably to IgG1 in humans and IgG2a in mice [64,65]. This receptor is saturated with serum IgG on leukocytes in the blood. As mentioned before, the other receptors have only low affinity and can bind to IgG only in the form of immune complexes (Table 1) [64,65]. In addition, it was clearly established that most FcyR have a binding preference for IgG1 and IgG3 over the other classes of IgG (Table 1). Similarly, in mice it was found that IgG1 binds only to FcyRIII, while IgG2a binds to all types of activating FcyR, and IgG2b binds to FcyRIII and FcvRIV. IgG3 does not seem to bind significantly to any of the FcyR (Table 1) [29,61]. In agreement with these data, IgG1 activity was lost in mice deficient in FcyRIII [61,66]. For IgG2a and IgG2b, however the correlation with particular $Fc\gamma$ receptors is not as simple. In some model systems the activity of these IgG classes was lost in FcyRIII-deficient mice, while it was not in others [13]. In a model of autoimmune hemolytic anemia, IgG2amediated response was highly dependent on FcyRIII, but also FcyRI and FcyRIV contributed to the development of severe anemia [67]. In another model of arthritis, mice deficient in FcyRI showed reduced cartilage destruction, and impaired protection from a bacterial infection, indicating the prominent role of FcyRI in IgG2a-dependent immune functions [68]. Thus, IgG2a used all activating Fcy receptors with important contribution from FcyRI and FcyRIII. Similarly, for IgG2b a particular interaction with a particular Fcy receptor cannot be clearly established. In models of IgG2b-dependent B cell depletion [69], and nephrotoxic nephritis [70], inhibition of FcyRIV prevented B cell destruction and kidney inflammation, suggesting a central role for FcyRIV in IgG2b-mediated functions. However, in models of IgG2b-dependent autoimmune hemolytic anemia [67], acute glomerular inflammation [63], or acute lung injury [71], FcyRIV and also FcyRIII were important for the activity of this IgG class. Thus, an order of activity is observed among the different IgG classes, and a preference of engagement with particular $Fc\gamma$ receptors.

Part of the mechanism used to create this IgG-Fc γ R selectivity is revealed by studies that measured the affinities of IgG classes toward both activating Fc γ recaptors and the inhibitory Fc γ RIIB [13]. In this way, it was found that IgG1 has higher affinity for the inhibitory Fc γ RIIB than for the activating Fc γ RIII (**Figure 6A**),

generating a high threshold for activation. In contrast, IgG2a and IgG2b have higher affinity for the activating FcvRIV than for the inhibitory FcvRIIB (Figure 6B). generating a lower threshold for activation. Thus, certain classes of IgG, such as IgG1, are more dependent on the signaling from the inhibitory receptor. In agreement with this view, deletion of the inhibitory FcyRIIB increased IgG1 activity in models of platelet depletion and tumor cell killing [61]. It is important to note that this model for IgG-FcyR selectivity is not static and can be modulated by other factors, such as the pattern of FcyR expression on the different leukocytes (Table 1), and cytokines that can modify FcyR expression (Figure 7). Thus, Th1-type cvtokines such as interferon- ν , and the anaphylatoxin C5a upregulate activating FcyRs expression and downregulate FcyRIIB expression [72,73], whereas Th2-type cytokines, such as interleukin (IL)-4, IL-10, and transforming growth factor-beta (TGF- β) upregulate FcyRIIB expression [8,74].

3.2. Each FcyR Initiates Particular Signaling Pathways That Lead to Unique Cell Responses

All the reports previously described have confirmed that different IgG classes mediate different cellular responses *in vivo* by engaging particular $Fc\gamma$ receptors depending on the relative affinity of these receptors for a particular



Figure 6. Relative preference of IgG classes for murine $Fc\gamma$ receptors. (A) IgG1 has higher affinity for the inhibitory FcγRIIB (thick arrow) than for the activating FcγRIII (thin arrow); (B) In contrast, IgG2a and IgG2b have higher affinity for the activating FcγRIV (thick arrow) than for the inhibitory FcγRIIB (thin arrow). Thus, certain classes of IgG, such as IgG1, are more dependent on signaling from the inhibitory receptor.



Figure 7. Cytokines can modify $Fc\gamma R$ expression. Th1-type cytokines such as interferon- γ IFN- γ , and the anaphylatoxin C5a upregulate activating $Fc\gamma R$ (green) expression and down-regulate $Fc\gamma RIIB$ (red) expression, whereas Th2-type cytokines, such as interleukin (IL)-4, IL-10, and transforming growth factor-beta (TGF- β) upregulate $Fc\gamma RIIB$ expression.

IgG class. However, these findings do not explain what cell response is activated in response to a particular type of Fcy receptor. One possibility is that each leukocyte is already programmed to respond in a certain way when immune complexes crosslink its Fcy receptors. Another possibility is that each FcyR signals differently to initiate a particular cell response. The first possibility is not very likely because a particular leukocyte can respond with various cell functions, and also because the same class of IgG induces different responses in different leukocytes. The idea that each FcyR is able to initiate a particular cell response is supported by recent reports where individual FcyR were crosslinked on human neutrophils. As described above, human neutrophils express only two FcyR, FcyRIIA and FcyRIIIB [28]. These receptors are different in the way they are anchored to the cell membrane. FcyRIIA has a typical transmembrane and cytoplasmic tail containing an ITAM for signaling. In contrast, FcyRIIIB is a GPI-linked receptor, lacking a cytoplasmic tail, and its signaling mechanism remains unidentified. However, early reports indicated that both FcyR are capable of signaling and inducing particular responses [75]. More recently, it was found that FcyRIIA, but not FcyRIIIB could induce an increase in L-selectin expression [76]. Opposite to this, FcyRIIIB, but not FcyRIIA, was able to activate $\beta 1$ integrins [77]. In addition, when the major cell response of neutrophils, arguably phagocytosis, was examined, FcyRIIA was the predominant FcyR mediating this response. FcyRIIIB contribution to phagocytosis was minimal [78]. In complete contrast, FcyRIIIB signaling to the neutrophil nucleus was much more efficient than FcyRIIA signaling. FcyRIIIB, but not FcyRIIA, promoted a robust increase in phosphorylated ERK in the nucleus, and also efficient phosphorylation of the nuclear factor Elk-1 [79]. Interestingly, the FcyRIIA

signaling pathway resembles the classical ITAM-mediated pathway (**Figure 2**) [5], while Fc γ RIIIB signaling pathway remains a mystery (**Figure 8**). Taken together, these reports strongly support the hypothesis that each Fc γ R is capable of initiating particular signaling pathways that lead to unique cell responses.

4. EFFECT OF ANTIBODY GLYCOSYLATION ON Fc RECEPTOR FUNCTION

All antibodies are glycoproteins with various carbohydrate side chains attached to the protein backbone. The immunoglobulin classes IgM, IgA, and IgE have several exposed carbohydrate side chains. In contrast, IgG molecules have one carbohydrate side chain. This carbohydrate (sugar) side chain is important for IgG function. Deletion of the sugar side chain results in an altered conformation of the antibody molecule and in deficient binding to Fcy receptors [80]. This carbohydrate domain is heterogeneous in its sugar composition. More than 30 different glycosylation variants of IgG can be found in serum of a healthy human or mouse individuals [14]. This heterogeneity is formed by variable addition of sugar residues such as sialic acid, N-acetylglucosamine, galactose and fucose in straight or branching patterns. Although heterogeneous, the pattern of glycosylation seems to change in various physiological conditions. For example, terminal galactose and sialic acid residues were reduced in active autoimmune disease [18,81], while they were increased during pregnancy [82,83]. It is not clear what these changes in the glycosylation pattern



Figure 8. Each FcyR initiates particular signaling pathways leading to unique cell responses. In human neutrophils, FcyRIIA activates the classical ITAM-mediated signaling pathway leading to efficient phagocytosis. In contrast, FcyRIIIB promotes a robust increase in phosphorylated ERK in the nucleus, and also efficient phosphorylation of the nuclear factor Elk-1. The FcyRIIIB signaling pathway remains a mystery. P represents a phosphate group; Syk, spleen tyrosine kinase; PI 3-K, phosphatidylinositol 3-kinase; MEK, ERK kinase; ERK, extracellular signal-regulated kinase.

represent, but it seems that they can modulate IgG activity [84]. In contrast, IgG antibodies with reduced fucose residues presented higher affinity for human FcRvIIIA and its mouse ortholog FcyRIV and showed improved antibody-dependent cellular toxicity against various tumor cells [61,85,86]. IgG antibodies with high levels of terminal sialic acid presented lower affinity for Fcy receptors and also reduced inflammatory activity [18,87, 88]. In addition, IgG antibodies with abundant sialic can bind to other cellular receptors different from Fc receptors. SIGNR-1 (specific ICAM-3 grabbing nonintegrin related 1) and its human ortholog DC-SIGN (dendritic cell specific ICAM-3 grabbing nonintegrin) were identified as receptors for sialic acid rich IgG [87]. Moreover, this subpopulation of antibodies was also suggested to be responsible for the anti-inflammatory activity of intravenous Ig (IVIg) therapy, because in SIGNR-1 knockout mice, IVIg did not show an anti-inflammatory effect in a model of rheumatoid arthritis [87]. IVIg therapy that consists on administration of high doses of pooled serum IgG from many different donors has been used for many vears to treat various autoimmune diseases such as rheumatoid arthritis and thrombocytopenia [19]. In addition, it has been reported that IVIg therapy can change the threshold for activation of cells by upregulation of the inhibitory FcyRIIB and downregulation of activating FcyR in some mouse models and in patients with chronic inflammatory demyelinating polyneuropathy [70,89,90]. Thus, glycosylation patterns are critical for binding to particular Fcy receptors and other novel antibody receptors. These reports underline the need of further studies on antibody-Fc receptor interactions to better understand the multiple effects of antibody molecules.

5. NEW LIGANDS FOR Fcy RECEPTORS

Antibodies are the bona fide ligands for Fc receptors. However, some recent reports have identified other molecules different from IgG that can bind Fcy receptors and can also activate the cell functions characteristic of antibodies. Two members of the pentraxin superfamily [91], which are multimeric cyclic proteins, are reported to bind human and mouse Fcy receptors. These pentraxins are C-reactive protein (CRP) and serum amyloid P (SAP) [92-95]. These proteins are usually not found in serum of healthy individuals, but they are rapidly expressed in large amounts during inflammation and microbial infections. CRP and SAP are capable of binding to several microorganisms including bacteria and fungi and thus targeting them for phagocytosis by neutrophils and macrophages [92,96]. These reports suggest that these pentraxin proteins behave like antibodies recognizing foreign antigens on pathogens and directing them to cells of the innate immune system [16]. In support of this idea, it was also found that Fcy receptor uptake of CRP-opsonized

Streptococcus pneumoniae increased the immune response against these bacteria [95]. Moreover, CRP also seems to have an anti-inflammatory effect mediated by Fcy receptors. In FcyR-deficient mice, administration of CRP did not protect from nephrotoxic nephritis or immune thrombocytopenia [94,97].

6. CONCLUSION

Fc Receptors expressed in a wide variety of leukocytes are capable of activating in response to various antibodies different cellular responses of great importance for host defense and for immune regulation. The different subclasses of IgG antibodies are recognized by Fcy receptors with different affinities. Also singular Fcy receptors seem to activate particular cell responses. This provides two ways for modulating cellular responses. In addition, expression of both activating and inhibitory $Fc\gamma$ receptors establishes a threshold for activation of innate immune cells. Thus, Fcy receptors are responsible of controlling the intensity of the immune response and of preventing unnecessary activation of innate immune cells, which might damage normal tissues. Novel glycosylation variants of IgG that bind Fcy receptors with different affinities have been identified and also the anti-inflamematory activity of intravenous IgG therapy. Finally, novel receptors for antibody variants and novel Fcy receptor ligands are been recognized. These new information together with new studies on IgG-Fcy receptor interactions will certainly help us to develop new ways of controlling not only antibody-mediated effector functions directed against pathogens and tumors, but also the exaggerated antibody-mediated cell responses associated with auto-immunity.

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