

Mitochondrial Signaling in Hypoxia

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Received February 21, 2013; revised March 21, 2013; accepted April 21, 2013

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ABSTRACT

This paper focuses on a bioenergetic mechanism responding to hypoxia. This response involves hypoxia-induced reprogramming of respiratory chain function and switching from oxidation of complex I (NAD-related substrates) to complex II (succinate oxidation). Transient, reversible, compensatory activation of respiratory chain complex II is a major mechanism of urgent adaptation to hypoxia, which is necessary for 1) succinate-related energy synthesis in the conditions of oxygen shortage and formation of urgent resistance; 2) succinate-related stabilization of HIF-1 α and initiation of its transcriptional activity related with formation of long-term adaptation; 3) succinate-dependent activation of the succinate-specific receptor GPR91. Thus, mitochondria perform a signaling function with succinate as a signaling molecule. Effects of succinate in hypoxia occur at three levels, intramitochondrial, intracellular and intercellular. In these settings, succinate displays antihypoxic activities. The review is focused on tactics and strategy for development of the antihypoxic defense and antihypoxants with energotropic properties.

Keywords: Respiratory Chain Reprogramming; Mitochondrial Complexes I and II; Adaptation to Hypoxia; Succinate; HIF-1; Energotropic Drugs; Antihypoxic Activities

1. Introduction

Mitochondria play a key role in oxygen homeostasis. This role is determined primarily by the trigger function of mitochondria in the cell, the function of a self-regulating biological machine that uses oxygen for generation of energy in the form of ATP and mitochondrial membrane potential. Inhaled oxygen eventually reflects the state and the oxygen demand of mitochondria because specifically mitochondria are the main oxygen consumer: up to 98% of oxygen delivered to the body work for mitochondrial respiration. This process provides up to 80% - 90% of ATP to cells of different mammalian tissues. Due to this function, which is responsible for vital capacity and function of aerobic organisms, evolution was able to produce most sophisticated physiological systems for oxygen delivery to mitochondria and maintenance of optimal cell oxygenation (act of respiration; lung system of oxygen transport; cardiovascular circulation; blood mass and transportation system; red cells; hemoglobin). Arrangement of the digestive system, including consumption and subsequent stepwise enzymic processing of food, is also determined primarily by the need for supplying substrates to reactions of mitochondrial oxidation and oxidative phosphorylation [1,2]. Mitochondria participate in

regulation of various physiological functions by providing energy for most of intracellular processes necessary for vital functions of the body including, first of all, contractility of the heart and smooth muscles of the gastrointestinal tract, blood vessels and lungs, maintenance of ion gradients in excitable tissues, accumulation of secretory material in vesicles, and support of hormonal and neurotransmitter functions.

Nevertheless, the major function of mitochondria is their **regulatory role in oxygen homeostasis**, which is evident at both systemic and cellular levels.

At the systemic level, mitochondria determine the concentration gradient of oxygen arriving from the environment since specifically mitochondria represent the final step of the interaction with molecular oxygen [1,3,4].

At the cell level, the mitochondrial respiratory chain performs as a signal-transforming metabolic system which activates the functional response and actualizes the physiological response to hypoxia.

Therefore, it is possible to say that *mitochondria are involved in intra- and extracellular signaling mechanisms and that mitochondria function as active signal organelles participating in transmission of information through various intracellular signaling pathways.*

It is known that hypoxia is associated with disturbances of ATP synthesis resulting from depressed functions of electron transport and oxidative phosphorylation in the respiratory chain. The factor determining the hypoxic state is disorders of oxygen delivery from the environment to the cell. In the cell, oxygen participates in reactions of aerobic energy synthesis as a substrate for cytochrome oxidase (COX), the terminal enzyme of mitochondrial respiratory chain. Therefore, oxygen deficiency in the extracellular environment may lead either to reduction or total inhibition of aerobic energy synthesis involving decreased content of high-energy molecules (ATP and PC), the main marker of hypoxic conditions. This results in inhibition of multiple energy-dependent reactions, such as formation of membrane potential, ion transport, electrogenic cell function, muscle contraction, receptor function, etc. Mitochondrial dysfunction inevitably leads to various pathologies and even death.

Therefore, defense of the body against disorders of oxygen homeostasis is primarily related with prevention of disturbances in the work of respiratory chain under oxygen shortage. The antihypoxic defence is a high priority for medicine as many abnormalities, such as ischemia, hypoxia, stroke, traumatic injury, inflammation, etc., involve impaired oxygen delivery to cells.

2. Performance of Respiratory Chain in Hypoxia

In normoxic conditions, performance of the respiratory chain is usually related with oxidation of NAD-related substrates, the major suppliers of reduction equivalents to the respiratory chain through mitochondrial complex I (MtC I—NADH: ubiquinone-oxidoreductase). This complex contribution as estimated by oxygen consumption may reach 55% - 65% in intact cells (**Figure 1A**). At the same time, 25% - 30% of mitochondrial respiration under these conditions is attributed to mitochondrial complex II (MtC II—succinate-ubiquinone reductase), which participates in the citric acid cycle by oxidizing succinate to fumarate. Its function is transferring electrons to the ubiquinone pool without translocating protons. In normoxia, the succinate level in mitochondrial matrix is very low (0.2 - 0.4 mM) [5]. The ratio between the two oxidation pathways depends primarily on properties of the principal enzyme complexes I and II. Kinetic characteristics of two major enzymes of these complexes, NAD-ubiquinon oxidoreductase (MtC I) and succinate dehydrogenase (SDH-MtC II), are tissue-specific and differ in intact animals with different sensitivity to hypoxia [3,6-10].

Hypoxic environments induce reversible depression of

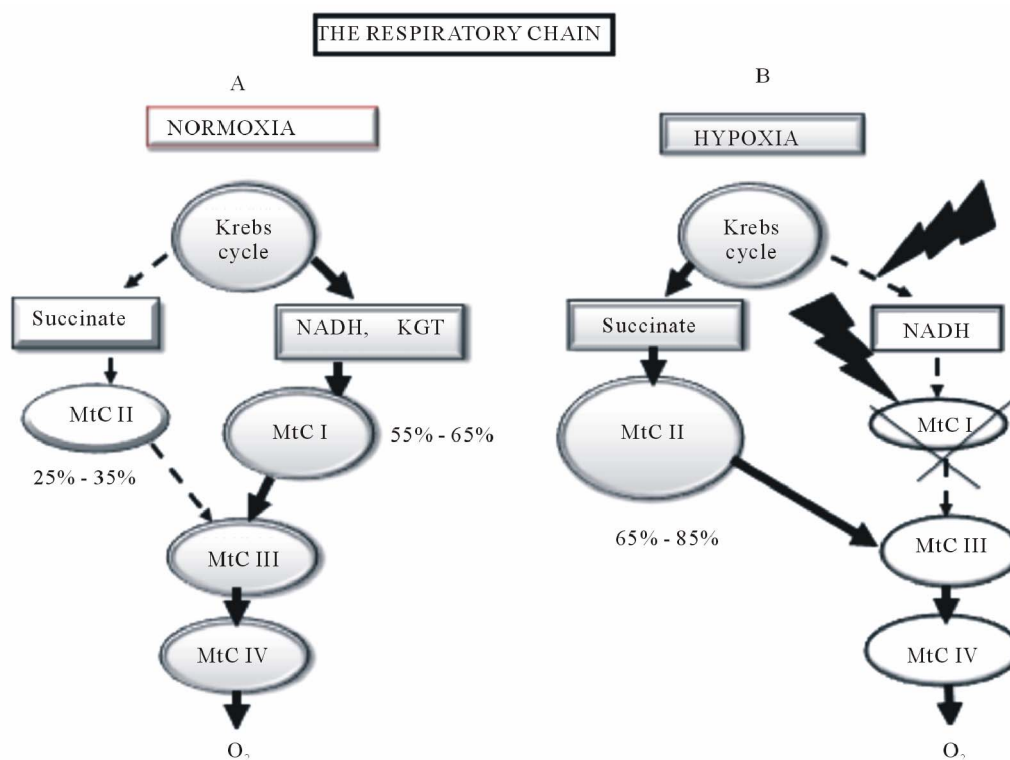


Figure 1. Reprogramming of the respiratory chain function and switching from oxidation of NAD-related substrates (MtC I) in normoxia A to succinate oxidation (MtC II) in hypoxia B. (A) Normoxia; high activity of MtC I (55% - 65% of mitochondrial respiration), low activity of MtC II (25% - 35% of mitochondrial respiration); (B) Hypoxia; depression of MtC I, activation of MtC II (65% - 85% of mitochondrial respiration).

MtC I function. At the present time, a great wealth of experimental evidence confirms the inactivation of MtC I electron transport function in hypoxic conditions, which persists and even increases in the post-hypoxic period (first 30 min - 2 hours of reoxygenation) [6-26].

It has been demonstrated that the hypoxic inactivation of enzyme complex I is only one of three steps in a sophisticated phase process of respiratory chain reprogramming. This process starts during a hypoxic impact and continues in the first hours of posthypoxic period [6-10]. These three steps are the following:

Step 1: Activated electron transport function of MtC I, which provides enhancement of ATP synthesis in early hypoxia. This activation reflects a primary compensatory mechanism for mobilization of basic cell energy resources under the conditions of comparatively mild environmental impacts such as slightly decreased oxygen in the environment.

Step 2: Depressed MtC I activity and, correspondingly, oxidation of NAD-dependent substrates along with *compensatory* activation of MtC II and succinate oxidation, with increased contribution of succinate oxidation to respiration and energy production. This is an alternative pathway for respiratory chain substrates, which provides reduction equivalents to MtC III and MtC IV under progressive hypoxia and plays a special role. In hypoxic conditions, this pathway is thermodynamically superior to oxidation of NAD-dependent substrates and provides a high energetic efficiency of the process as a whole, although only two phosphorylation points are preserved in this process (**Figure 1B**).

Step 3: The step of *decompensation* differs from the first two steps. Decompensation develops under an acute or long-term hypoxic impact. Decompensation is associated with disturbed electron transport function of the respiratory chain in the region of MtC III (cytochromes *b-c*) and then MtC IV (cytochrome oxidase, COX), which leads to general de-energization.

As the severity of hypoxia increases **Step 1** becomes shorter and **Step 2** becomes more pronounced. However when the inhaled O₂ concentration falls below 8%, the capability for MtC II activation decreases.

Therefore the respiratory chain reprogramming in response to hypoxia starts from its substrate region. The respiratory chain terminal region (cytochrome oxidase, COX) is not a limiting step of this process in a broad range of oxygen concentrations up to anoxia, due to COX kinetic properties (low values of K_m (O₂), which determine high affinity of COX to oxygen).

Mitochondrial reprogramming induced by hypoxia correlates with phase changes in the content of adenine nucleotide pool components, ATP, ADP and AMP [3,9,10]. The first step is characterized by a relatively minor, mild (within 6% - 10%) increase in ATP level followed by

transition to a somewhat reduced (by 10% - 15%), steady-state ATP level (second step), which reflects ATP hydrolysis. This reduction is accompanied by a rise of intracellular ADP content and a decrease in ATP/ADP ratio, respectively. However, in parallel with the ongoing slight reduction of intracellular ATP level, the intracellular AMP content gradually raises. The regulatory significance of ATP/ADP ratio gradually decreases and becomes replaced by the control of ATP synthesis through the ATP/AMP ratio. At the last stage of hypoxia, when a linear decrease in ATP content is observed, the regulatory role of ATP/ADP and ATP/AMP ratios becomes minimal, and total adenine nucleotides plus energetic charges decrease [3,9]. The hypoxia-induced changes in adenine nucleotides pool resulting from disturbed electron flow and oxidative phosphorylation reactions precede changes in other functional and metabolic parameters responsible for the cell vital activity. Contrastingly, the compensatory role of glycolysis as an ATP supplier is effective only in early hypoxia. During the terminal period, despite a sharp activation of glycolysis, glycolysis fails to prevent the decrease in ATP content [3,9].

Signs of hypoxic inactivation of MtC I include an initial increase and subsequent decrease in both the intensity of NAD-dependent substrate oxidation and efficiency of coupled oxidative phosphorylation, reduced sensitivity of respiration to specific inhibitors of the respiratory chain NAD-dependent region (ex., rotenone, amytal) in various tissues (brain, heart, liver), and reduction of MtC I respiratory carriers (pyridine nucleotides and flavins), which reflects suppression of reduction equivalent transport through this regions [3,4,6-9].

This process is associated with activation of MtC II (SDH) and increased contribution of succinate oxidation to cell respiration; this contribution may reach 70% - 80% [4,6-11,16-19,21,22,24-41].

Therefore, hypoxic changes are associated with succinate accumulation in tissues and blood. The accumulation of succinate in tissues during hypoxia and the reprogramming of respiratory enzyme performance develop fast and under different levels of oxygenation. For example, Goldberg *et al.* found that after 30 sec of global brain ischemia, succinate level increased 1.5 times in the setting of reduced concentrations of certain NAD-dependent substrates [19]. Similar data were also obtained by other researchers. In the first 30 minutes of anoxic incubation, formation of succinate increases by an order of magnitude, reaches 4 - 7 mM and continues to grow for the first 30 minutes of reoxygenation [17,35,41]. As early as during the first minutes of hypoxic exposure, succinate concentration in blood and tissues increases by an order of magnitude, which allows considering succinate a marker of hypoxia [3-5,10,16,17,31-33,35,41,42]. Any regimen of hypoxic exposure with any level of oxygenation re-

sults in pronounced changes in kinetic properties of MtC I and MtC II including increased K_m values for complex I and decreased K_m values for complex II. This process reflects impaired and enhanced efficiency of enzyme performance, respectively [3,4].

Note that pheno- and genotypic peculiarities of the response to hypoxia are essential; these changes are relatively more pronounced in the brain of low-resistance rats and are not statistically significant for the neocortex of high-resistance rats [3,4,7-9].

Depending on tissue-specific peculiarities of energy metabolism, formation of endogenous succinate in hypoxia uses different pathways. However all these pathways are based on aspartate- and glutamate-dependent aminotransferase reactions, substrate-level α -ketoglutarate phosphorylation during the conversion of α -ketoglutarate to succinate by α -ketoglutarate dehydrogenase, reversal of tricarmonic cycle and oxidative phosphorylation reactions (substrate-level phosphorylation in citric acid cycle via 2-oxoglutarate dehydrogenase and fumarate reductase reactions) [18,25,31,32,35,41,43-46].

Activation of succinate oxidation constitutes a regulatory and compensatory mechanism, which occurs in most tissues (brain, myocardium, liver, kidneys, lymphocytes) in the conditions of oxygen shortage [7,35]. This mechanism prevents or attenuates disorders in ATP synthesis, exerts normalizing effect on the adenylate pool and vital functions of the body, stabilizes and normalizes pH to eliminate hypoxia-characteristic acidosis, and increases the resistance to oxygen shortage. If such switch does not occur in hypoxia, MtC I dysfunction leads to severe deenergization (decreased membrane potential, ATP loss, and changes in the adenine nucleotide pool) and disturbed respiration due to oxidation of NAD-related substrates (electron donors for MtC I) [3,4,7,9,25,26,31,32, 41].

This regulatory reprogramming is designed for using the succinate oxidase pathway of substrate oxidation, which is an energetically more efficient pathway in hypoxic conditions. This reprogramming prevents or alleviates disorders of ATP synthesis, adenylate pool parameters, vitally significant functions of the body, and typical for hypoxia acidosis. As a result, the body resistance to oxygen shortage increases. Hypoxia-induced reprogramming of the respiratory chain substrate region correlates with phase changes observed at the systemic level in hypoxia [9,10,36].

The switch between mitochondrial substrate oxidation pathways accompanies practically all types of hypoxia or ischemia and incorporates into these pathways as a mandatory unit, a *basic molecular (bioenergetics) mechanism*. Thus, the reversible reprogramming of respiratory chain during hypoxia (switching from oxidation of NAD-re-

lated substrates to succinate oxidation is an *obligatory, evolutionary developed, urgent signaling compensatory mechanism*. Due to this mechanism, the energy synthesizing function of respiratory chain is preserved under disturbed oxygen homeostasis.

3. Mitochondrial Signaling in Transcriptional Activity at Hypoxia

The reprogramming of respiratory chain performance in hypoxia is associated with the transcriptional activation of the hypoxic factor, HIF-1 α . The process of adaptation to low oxygen level is known to be regulated mostly by a specific transcription protein factor induced in all tissues under hypoxia, hypoxia-inducible factor 1 (HIF-1) [47-53]. This factor is a heterodimeric transcription protein composed of HIF-1 α and HIF-1 β subunits, which functions as a master regulator of oxygen homeostasis. Regulation of HIF-1 activity is attributed mostly to α subunit.

In normoxic conditions, due to the regulatory interaction between mitochondria and cytosol, prolyl hydroxylase reactions producing proteasomal degradation of HIF-1 α are well supplied with substrates required for activation of these reactions, including aspartate and α -ketoglutarate [20,53,54]. α -Ketoglutarate is synthesized in the tricarmonic acid cycle and is oxidized in substrate phosphorylation reactions and NAD-dependent reactions catalyzed by MtC I. The malate-aspartate bypass can supply α -ketoglutarate to cytosol for prolyl hydroxylase reactions. In normoxic conditions, the involvement of α -ketoglutarate in prolyl hydroxylase reactions is associated with formation of succinate, the prolyl hydroxylase reaction inhibitor in cytosol (α -ketoglutarate + O₂ \rightarrow succinate + CO₂). Apparently this is necessary for slowing these reactions, maintaining the basic HIF- α level, and inducing basic levels of genes encoding the mitochondrial complex protein responsible for energy production.

Hypoxia inhibits both electron transport in the respiratory chain through MtC I and activity of the malate-aspartate bypass. In result, cytosol becomes deficient in α -ketoglutarate and aspartate. At the same time, hypoxia is associated with activation of succinate dehydrogenase (MtC II), and increased formation of succinate in aminotransferase reactions. Succinate is an inhibitor of prolyl hydroxylases. These processes along with O₂ and Fe²⁺ shortage create favorable conditions for inhibition of prolyl hydroxylase reactions, HIF-1 α stabilization, induction of transcriptional processes, including HIF-1 α translocation to the nucleus, HIF-1 α heterodimerization with HIF-1 β , conformational changes, formation of the transcription-active complex (hypoxia response element, HRE), expression of HIF-1-dependent target genes, and synthesis of protective, adaptive proteins in response to hypoxia

[54-58].

By the present time, conclusive evidence has demonstrated a direct correlation between inactivation of MtC I and activation of MtC II and formation of HIF-1 α (**Figure 2**). For instance, it was shown that even a partial (20%) suppression of MtC I activity almost completely inhibited the hypoxic induction of HIF-1 α . However it recovered in the presence of succinate [12-14,58]. We also showed that activation of HIF-1 α synthesis after a single hypoxic exposure in the regimen of preconditioning correlated with increased efficiency of MtC II activity and was associated with a multiple increase in the resistance of animals to acute hypoxia. In the absence of activated succinate oxidation, like, for example, in the neocortex of high-resistant rats, HIF-1 α was undetectable [10,37]. *Therefore, induction of HIF-1 α requires a low MtC I activity and a high MtC II activity, i.e., potentiation of succinate oxidase oxidation.*

Earlier we have shown that on the first day after any hypoxic exposure, a *correlation* is observed between intensity of succinate oxidase oxidation, intensity of HIF-1 α expression, and capability of the body for developing adaptive responses in these conditions [10,18,36, 37]. An inverse relationship also exists; HIF-1 α can influence enzyme activities in the mitochondrial respiratory chain. HIF-1 α suppresses pyruvate oxidation in the tricarboxylic acid cycle, that is, facilitates inhibition of MtC I by activating pyruvate dehydrogenase kinase 1

(PDK-1). After phosphorylation of PDK-1, it inhibits pyruvate dehydrogenase [33,48,59].

Therefore, *an important mechanism of urgent adaptation to hypoxia is the two-way regulatory interaction between the respiratory chain substrate region and expression of the protein factor, HIF-1 α* . After a hypoxic exposure, the switch of metabolic pathways in the respiratory chain to *succinate* oxidation provides HIF-1 α accumulation and induces transcriptional processes. Simultaneously, the succinate-dependent accumulation of HIF-1 α potentiates inhibition of complex 1 and creates conditions for maintaining succinate oxidase oxidation and HIF-1 α accumulation.

In summary, it is obvious that the *reprogramming* of mitochondrial respiratory chain during the initiation phase of urgent compensatory, adaptive responses is the most important regulatory mechanism. This mechanism provides: 1) more efficient succinate-dependent energy production in the conditions of rapidly progressing oxygen shortage; 2) succinate-dependent stabilization of HIF-1 α and subsequent initiation of HIF-1 α transcriptional activity; 3) succinate-dependent formation of urgent resistance to hypoxia in the body.

4. Role of Mitochondria in Cell-to-Cell Interactions

The increased level of succinate in blood and cytoplasm

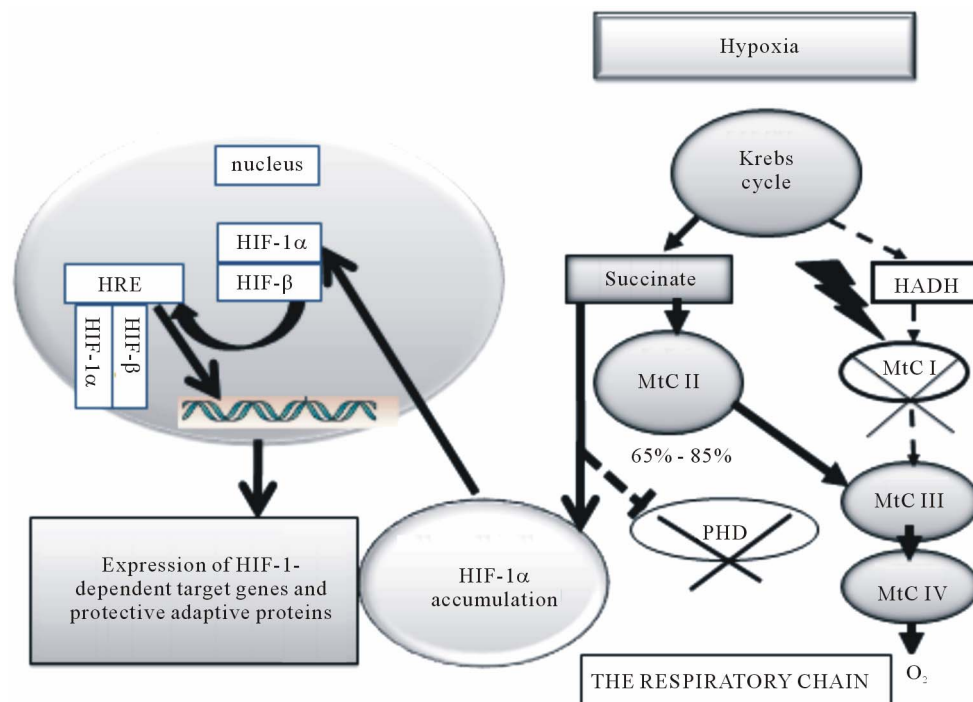


Figure 2. Role of mitochondrial signaling in transcriptional activity at hypoxia. Increased succinate synthesis in hypoxia can provide depression of prolyl hydroxylase activity (PHD), accumulation of HIF-1 α in cytosol and expression of HIF-dependent target genes.

allows considering succinate a marker for hypoxic states [60]. It means that succinate may participate in cell-to-cell interactions and regulatory, functional and metabolic processes at the systemic level. For instance, it has long been known that succinate contributes to regulation of blood pressure. Succinate stimulates renin formation in renal tubules and, thereby, produces a vasodilatory effect in hypoxia [61].

Studies have shown that a succinate-controlled electron bypass exists between peripheral cells and lungs. In hypoxia, blood transports succinate from peripheral hypoxic tissues to lungs, where succinate synthesis is depressed by intensive oxygenation. Delivered succinate is used in lungs as an energy substrate needed for pulmonary vasoconstriction. In this process, succinate is oxidized to fumarate which is again transported with blood to peripheral cells and again forms succinate through the fumarate oxidase reaction [43,62].

In the recent two decades, data on a receptor function of succinate have appeared. Reports have shown that MtC II contributes to gene expression in glutamate- and dopaminergic signaling systems [53].

A sensational discovery was made by a research group in San Francisco in 2004. This study demonstrated that succinate was a specific ligand of GPR91 [63]. This receptor belongs to a group of guanine nucleotide binding protein coupled receptors which form large and diverse gene families translating extracellular signals to intracellular ones. However the sequence similarity among the most distant GPCRs is minimal [64]. GPR91 is localized in plasma membranes. It is found in more than 20 tissue types, and the array of its localizations is continuously expanding. Kidneys, liver, spleen, small intestine, and bladder possess the best capability for expressing this receptor. Only succinate activates GPR91 in a receptor-specific way and also induces GPR91 internalization [65].

Succinate-dependent receptor activation induces:

- accumulation of inositol phosphate and increases $[Ca^{2+}]_i$;
- extracellular signal-regulated kinase (Erk);
- mitogen-activated protein kinase, p38 and cyclooxygenase;
- synthesis and release of prostaglandin E2 and NO;
- Taken together, these results suggest that GPR91 activation by succinate couples to at least several signaling pathways [46,65]. However, the succinate signal is apparently tissue-specific, and these pathways may not work in all tissues.

In kidneys, the succinate-induced expression of GPR91 associated with GPR91 internalization involves the renin-angiotensin system and precipitates renovascular hypertension, a disease closely related to atherosclerosis, diabetes mellitus and renal insufficiency [42,65]. This

process is absent in GPR91-deficient rats [65]. It has been demonstrated recently that this activation is a part of kidney-specific paracrine signaling pathway initiated by high glucose levels. The signaling cascade includes local accumulation of succinate as well as expression and internalization of GPR91 receptors in endothelial cells of renal tubules. This GPR91 signaling cascade can modulate renal function and provide elimination of various metabolic products through renal hyperfiltration. The GPR91 cascade is apparently a connecting link to pathologies such as diabetes mellitus, syndrome of renin-angiotensin system hyperactivity, systemic hypertension, and organ injury [46].

In liver ischemia associated with intensive succinate release, GPR91 is expressed only in hepatic stellate cells. However the fact that succinate does not increase hepatic perfusion pressure suggests that succinate activities in the liver do not contribute to its hypertensive effects. GPR91 transforms the signal related with increased extracellular levels of succinate into an intracellular signal providing activation of stellate cells in response to organ injury. Therefore the effect of succinate should be regarded as a paracrine signal in this instance as well.

Succinate-dependent expression of GPR91 in ischemia was observed in dendritic cells; it results in production of various angiogenic factors, including VEGF. In these cells, succinate triggers mobilization of intracellular calcium, induces migratory responses and acts in synergy with Toll-like receptor ligands to produce pro-inflammatory cytokines [66].

Succinate also enhanced antigen-specific activation of human and mouse helper T cells. GPR91-deficient mice showed less migration of Langerhans cells to draining lymph nodes and impaired tetanus toxoid-specific recall T cell responses [67].

Studies have shown that succinate accumulates in hypoxic retina of rodents and, via GPR91, acts as a potent mediator of vessel growth in the settings of both normal retina development and proliferative ischemic retinopathy. These effects of GPR91 are mediated by retinal ganglion neurons. These neurons respond to increased succinate levels and regulate production of numerous angiogenic factors, including VEGF. Accordingly, succinate did not display any angiogenic effects in RGC-deficient rats [68].

As a GPR91 ligand, succinate can regulate lipolysis in white adipose tissue suggesting that the succinate signaling may regulate energy homeostasis [69]

The receptor function along with the participation of succinate in cell-to-cell signaling and systemic regulation can explain many rapid effects of this metabolite, including the role of succinate in development of urgent adaptation. Data on the interaction of GPR91 with other

receptors help to understand also the regulatory participation of catecholamines and acetylcholine in activation of succinate dehydrogenase.

Therefore, the response to hypoxia initially develops at the subcellular, mitochondrial level through compensatory potentiation of enzyme complex II and involves activation of a specific succinate-dependent receptor (**Figure 3**). This receptor initiates cytoplasmic signaling pathways which coordinate and facilitate formation of adaptive processes, as well as cell-to-cell interactions at the systemic level.

5. Use of Succinate-Containing Drugs as Energotropic Medicines in Clinical Practice

High prevalence of diseases bearing a hypoxic component defines the exceptional importance and social significance of protecting the body from oxygen shortage and related energy deficiency. The above data also show that understanding the role of succinate signaling in the electron transport function of respiratory chain under hypoxia is an essential practical challenge that has defined the tactics and strategy for antihypoxic defense and the development of antihypoxants with energotropic action.

The energotropic therapy implies using as medicines

the substances *that are able to interact with the mitochondrial respiratory chain and to prevent disorders of energy metabolism up to its complete recovery in pathological conditions*. Principles of energotropic therapy are based on the concept of the phase nature of hypoxia-induced mitochondrial dysfunction related with changes in activities of mitochondrial enzymes.

The central role among such medicines belongs to succinate-containing drugs which have been successfully used as effective antihypoxic and energotropic therapy [8,9,18, 35-37]. The most effective of these drugs are structural derivatives of vitamin B₆, which belongs to 1-hydroxypyridine derivatives (ex., mexidole, proxipine). These drugs are used in the early period of acute disorders induced by oxygen shortage [8,10,18,37].

Special studies showed that the *succinate moiety of these drugs is used by the respiratory chain as an energy substrate according to the classic schedule for succinate*. In this process, respiration becomes stimulated, and the electron flow switches from complex I to complex II (succinate monopolization of the respiratory chain as demonstrated by the increased sensitivity of respiration to malonate and the decreased rotenone-sensitive respiration due to oxidation of NAD-dependent substrates) [18, 37,39].

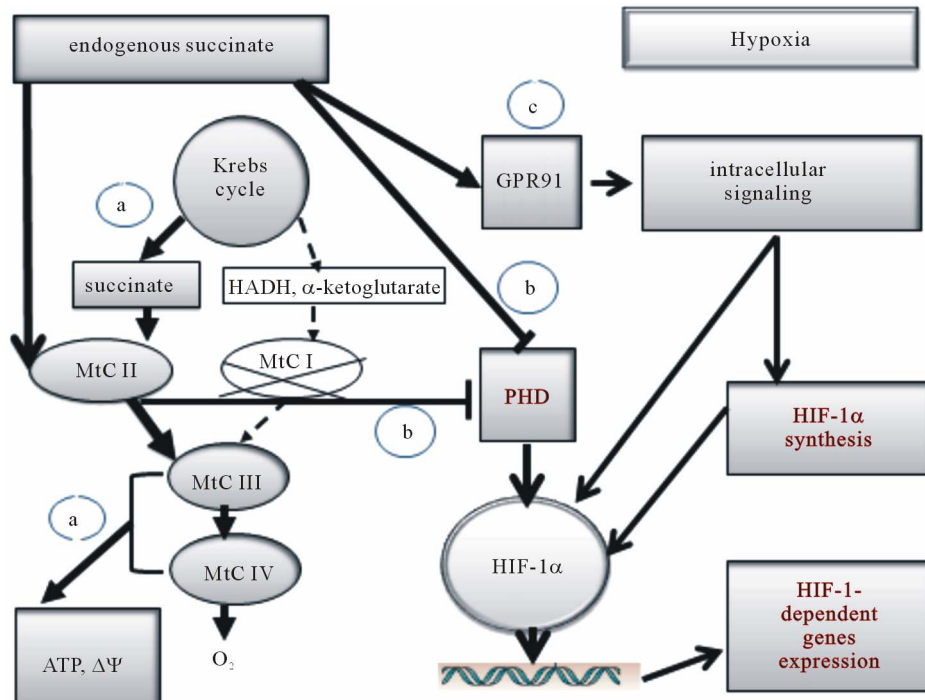


Figure 3. Role of mitochondrial signaling in cell-to-cell interactions in hypoxia. Reprogramming of the mitochondrial respiratory chain in hypoxia is a regulatory and compensatory mechanism, which provides: 1) activation of succinate oxidation, a more efficient pathway of energy production in the conditions of rapidly progressing oxygen shortage (a); 2) succinate-dependent PHD inhibition, subsequent stabilization of HIF-1 α , and initiation of HIF-1 α transcriptional activity (b); 3) succinate-dependent activation of the GPR91 signaling pathway and related signaling pathways, stimulation of HIF-1 α synthesis and transcriptional activity (c).

The obtained results suggest that the succinate-containing drug, proxipine, is a succinate donor for the respiratory chain. In hypoxic conditions, proxipine functions as an antihypoxant potentiating activation of the succinate oxidation pathway to facilitate recovery and normalization of aerobic energy production. Furthermore, the energotropic effect of succinate-containing drugs has been demonstrated to be due to oxidation of succinate incorporated into the structure of 3-oxypyridine by a classic pathway for succinate in the respiratory chain [18]. All succinate-containing drugs are very rapidly absorbed in various types of hypoxia/ischemia. They exert stabilizing or restoring effects on intracellular ATP already 15 minutes after administration.

Energotropic and antihypoxic effects of succinate-containing compounds are associated with 1) modification and resynthesis of phospholipids; 2) normalizing effect on calcium metabolism; 3) catecholamine-mimetic, antiteratogenic, antitoxic, hepatoprotective, antiketogenic, and anticholesterolemic effects; 4) removal of excessive acetyl-Co-A associated with decreases in excessive lipids and their metabolites; 5) reduction and normalization of

pH and elimination of metabolic acidosis [18,37,70].

Proxipine is characterized by a broad range of physiological actions. Succinate-containing drug therapies used during the phase of formation of urgent compensatory adaptive mechanisms, *i.e.*, in the first 1 - 3 days of global cerebral ischemia, stroke, myocardial infarction, acute heart failure, traumatic shock, resuscitation after heart arrest, early postoperative period, after anesthesia, etc., exert pronounced protective, antihypoxic effects and increase ATP in tissues (Figures 4(a)-(c)). Along with the energotropic and antihypoxic effects, proxipine beneficially influence multiple vital functional parameters in hypoxia and ischemia. These drug reduce the death rate, recover the ability of the body for gaining weight, decrease severity of neurological disorders (Figure 3(d)) and aggression typical of hypoxia, exert antistress and normalizing effects on locomotor, exploratory and emotional activities of animals [10,18,36,37].

Timely use of this drug decreases mortality and provides faster and more complete regression of general and focal cerebral symptoms in the majority of patients. Therefore, a single preventive or therapeutic administra-

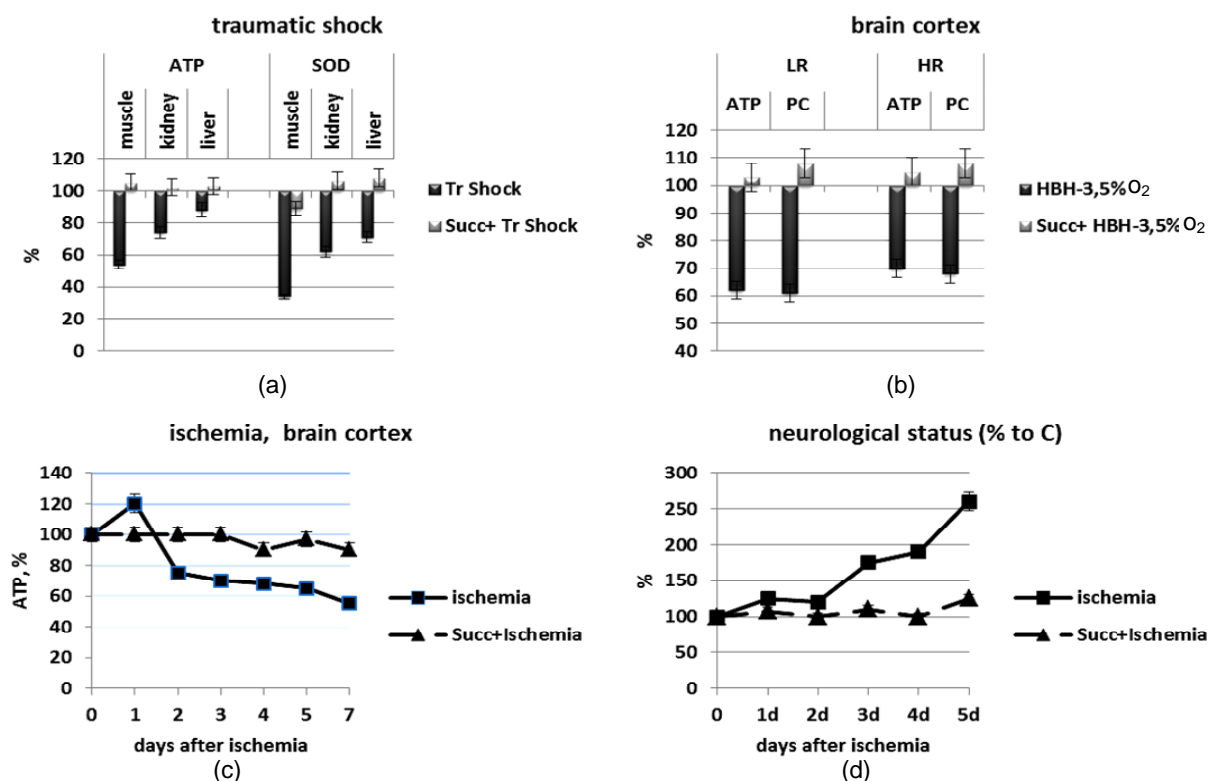


Figure 4. Energotropic and antihypoxic effects of the succinate-containing compound proxipine. Normalizing effect of the succinate-containing drug proxipine (3-hydroxypyridine, 40 mg/kg, i.p.) on the following parameters: (a) ATP level and SOD activity (% of control) in tissues of rats exposed to traumatic shock (Tr Shock). Tr Shock, without proxipine; Succ+ Tr Shock, with proxipine; (b) ATP and phosphocreatine (PC) levels (% of control) in brain cortex of rats exposed to altitude chamber hypoxia (acute hypobaric hypoxia, HBH, 3.5% O₂). HBH, without proxipine; Succ + HBH, with proxipine; LR, low-resistance rats, HR, high-resistance rats; (c) ATP level (% of control) in brain cortex of rats exposed to global ischemia; (d) Neurological status of rats.

tion of succinate-containing drugs in the setting of hypoxic therapy will improve the development of urgent resistance in the body and, therefore, it may be used in practice for activation of urgent adaptive mechanisms [10,37]. However a longer use of succinate-containing drugs loses its protective effects because it began to hamper the formation of long-term mechanisms of adaptation. Therefore the optimal therapeutic effect of succinate can be achieved with no more than three injections at early stages of hypoxic exposure.

6. Conclusions

The role of mitochondria as a cell “power-station” is well known. Participation of mitochondria in essential regulatory processes determining intracellular, intercellular and systemic interactions is presently discussed [19-21, 29,30,32,33,35,49-56,58,60,64,71-75]. However mitochondria also play an important regulatory role in the conditions of hypoxia.

In hypoxia, the mitochondrial respiratory chain not only directly participates in formation of both early and late adaptive signs, but it is also involved in the sophisticated system of intra- and intercellular signaling. This system provides systemic responses to oxygen shortage. At least three regulatory functions of this system can be distinguished: 1) *compensatory* function, which is responsible for development of urgent adaptive responses and resistance to hypoxia; this function is associated with changes in kinetic properties of mitochondrial enzymes and pathways of energy substrate oxidation; 2) *transcriptional* function, which provides HIF-1 dependent expression of genes responsible for development of mechanisms of long-term adaption to low pO₂; 3) *receptor* function, which is related with participation of mitochondria in the system of intercellular signaling (Figure 3).

The tricarboxylic acid cycle intermediate, succinate, plays a special role in these processes. Succinate is a signaling molecule that is involved in this process during various kinds and regimens of hypoxic exposure. In early hypoxia, for example, the ability of respiratory chain for reprogramming (switching from NAD-dependent substrate oxidation to succinate oxidation, which is energetically more efficient in these conditions) maintains both aerobic energy production and development of urgent resistance in the body (Figure 1). If such switch fails urgent mechanisms of adaptation cannot develop.

Increased intracellular succinate provides succinate-dependent stabilization of HIF-1 α and initiation of its transcriptional activity. Therefore, long-term adaptation to hypoxia is a succinate-dependent process, during which HIF-1 α dependent formation of adaptive genes and generation of a new enzyme spectrum (including mitochondrial enzymes) can ensure the vital activity of and the energy supply to cells in conditions of high reduction and

low oxygen concentration (Figure 2).

Another, no less important signaling mechanism in hypoxia is succinate activation of a succinate-specific and very common tissue GPR91 receptor. The receptor signaling function provides initiation and coordination of a broad spectrum of other signaling pathways which may not only facilitate formation of adaptive processes but also provide cell-to-cell interactions at the systemic level (Figure 3).

Understanding of biological mechanisms for adaptation to hypoxia has substantiated the development of a new strategy for antihypoxic defense. A single preventive or therapeutic administration of succinate-containing drugs in the setting of hypoxic therapy enhances development of urgent resistance in the body and, therefore, it may be used in practice for activation of urgent adaptive mechanisms.

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Abbreviations

ATP, adenosine triphosphate;
ADP, adenosine diphosphate;
AMP, adenosine monophosphate;
COX, cytochrome oxidase;
MtC, mitochondrial complex;
GPR91, succinate-related guanine nucleotide-binding protein-coupled receptor;
HBH, acute hypobaric hypoxia;
HIF, hypoxia-inducible factor;
HRE, hypoxia response element;
HR, high-resistance rats;
LR, low-resistance rats;
NAD, NADH, nicotinic adenine nucleotides;
PC, phosphocreatine;
PDK, pyruvate dehydrogenase kinase;
SDH, succinate dehydrogenase.