

Motor Effects of 1,3-Disubstituted 8-Styrylxanthines as A₁ and A₂ Adenosine-Receptor Antagonists in Rats

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ABSTRACT

A series of 1,3-substituted 8-styrylxanthines (**11a-d**) was synthesized, under chemo- and regioselective conditions, in a good overall yield. The compounds showed affinity towards both A₁ and A_{2A}-adenosine receptors by radioligand binding by means of *in vitro* assays. The (*E*)-3-ethyl-1-propyl-8-styrylxanthine (**11a**) showed the greatest affinity towards the A_{2A} receptor, whereas (*E*)-3-pentyl-1-propyl-8-styrylxanthine (**11d**) showed the greatest affinity for the A₁ receptor. When the 8-styrylxanthines **11a** (**A15Et**) and **11c** (**A15Bu**) were administrated in rats, which were previously injured with 6-hydroxydopamine at the *substantia nigra pars compacta* (SNc), the turning behavior decreased 50%. Based on these results we propose to A15Et as a potential compound to treat some symptoms of Parkinson's disease.

Keywords: Xantines; Adenosine Receptors Antagonists; Turning Behavior; Anti-Parkinsonian Drugs

1. Introduction

The nucleoside adenosine exerts a modulatory influence on the central nervous system by the activation of the G-protein-coupled receptors, through a mechanism involving vasodilatation, inhibition of lipolysis, and insulin release [1,2]. Most of the actions of adenosine are mediated by four extra-cellular receptors [3] termed A₁, A_{2A}, A_{2B} and A₃. The activation of A₁ and A₃ receptors inhibits adenylate cyclase through G_i coupling, and activation of A_{2A} and A_{2B} receptors stimulates adenylate cyclase through G_s coupling [4]. The A₁ receptors are widely distributed throughout the brain, where they regulate neurotransmitter release and neuronal firing by activation of potassium channels. The distribution of A_{2A} is heterogeneous with high levels of expression in the *nucleus accumbens*, olfactory tubercle, *globus pallidus* and striatum [5,6] where A_{2A} receptors are co-localized with dopamine D₂ receptors in the *striato-pallidal* terminal [7]. The A_{2A}-D₂ interaction seems to be more potent in striatal dopamine denervation with supersensitive D₂ receptors [8]. This fact has been confirmed in the course of the

binding assay of the selective agonists and antagonists [6,8,9]. When considered together with the important role of dopamine in the control of motor activity and in the etiology and management of Parkinson's disease (PD), this observation suggested that adenosine A_{2A} receptors could be a novel target for drugs that manage movement disorders [9-11].

Results from recent PD studies on animal models suggest that antagonism of the A_{2A} receptor may also protect against underlying neuronal degenerative processes [9, 12].

The blockade of A_{2A} receptor reverses damage by excitotoxicity [13] and counteracts the motor deficits produced by dopaminergic toxin [14].

Recent preclinical and clinical data suggest that A_{2A} antagonist may provide a complementary therapy for PD [11,15,16]. Furthermore the A_{2A} receptor participates as a neuroprotector agent [12,14,15]. The (*E*)-8-(3-chlorostyryl)caffeine (CSC) has been employed as a selective inhibitor of the A_{2A} subtype of adenosine receptor [17]. However several analogues of CSC act as inhibitors of monoamino oxidase B (MAOB) [18-23]. Because of their role in the metabolism of monoamines and the

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inhibition of MAOB, CSC is being used in preclinical studies for treatment of symptoms of PD [20-23].

Therefore the protective effect may be related to the ability of the A_{2A} antagonists to antagonize the increase of toxicity of glutamate induction by quinolinic acid. Recently, some researchers have proposed that the inhibitors of MAOB, caffeine and their derivatives reduce the risk of developing PD [23] and produce an anxiolytic effect [22]. Current data suggest that adenosine acts on dopamine regulation and dopaminergic function, and that the antagonists of the adenosinergic receptor decrease the dopaminergic activity [23].

For the above mentioned reasons, A_{2A} receptors have become important targets for development of drugs for the treatment of PD [10,23]. Some structures have shown affinity and selectivity towards A_{2A} receptors [9,19,24, 25], as xanthines, adenines, 1,2,4-triazolo[1,5-*a*]quinoxalines and pyrazolo[3,4-*d*]-pyrimidines (**Figure 1**). Pharmacological studies suggested that derivatives of 8-styrylxanthine have biological activity as an anti-Parkinsonian drug [26-28].

In this paper we report the synthesis, binding studies and pharmacology properties of 8-styrylxanthines. We found that (*E*)-3-ethyl-1-propyl-8-styrylxanthine (A15Et, compound **11a**) showed the greatest affinity for the A_{2A} receptor, also the biological experiments showed a decrease in the turning behavior, so we suggest that this compound represent a potential drug for treatment of Parkinson's disease.

2. Materials and Methods

2.1. Synthesis of Chemical Compounds

The melting points were measured with a Melt-temp apparatus and were not corrected. The NMR spectra were

recorded on a Varian spectrometer; ¹H spectra at 400 MHz and ¹³C at 100 MHz. DMSO-d₆ and CDCl₃ were used as solvents. All chemical shifts are quoted in ppm and the coupling constants are expressed in Hertz. The chemical shift of the remaining no deuterated protons of DMSO-d₆ served as an internal standard (δ ¹H: 2.50, ¹³C: 39.7). The IR spectra were obtained from a Shimadzu FT-IR 8400 spectrophotometer, using KBr pellets; wavenumbers of maximum absorption peaks are reported as cm⁻¹. The UV spectra were recorded in a Perkin Elmer Lambda Bio 40 apparatus, using ethanol as solvent; λ_{\max} is described in nm. TLC was performed on 60 F₂₅₄ silica gel plates.

2.1.1. Synthesis of 6-Amino-3-propyluracil (**6**)

A suspension of 6-aminouracil (**5**) (5.08 g, 40 mmol) and (NH₄)₂SO₄ (250 mg) in hexamethyldisilazane (14 mL, 60 mmol), was refluxed for 45 min. In this time the mixture became transparent indicating that all **5** was trisilylated. The crude was allowed to cool to 45°C; then, the *n*-propyl iodide (1.9 mL, 20 mmol) was added. The reaction was refluxed for 2 h. The crude was treated with an aqueous solution of Na₂S₂O₃ (6 g/20 mL). Additionally, a saturated aqueous solution of NaHCO₃ (20 mL) was added drop wise until effervescence ceased. The suspension obtained was filtered, and the white precipitate was washed with cold water, diethyl ether, and hexane. Yield: 92%; mp 267°C - 269°C (lit. 275°C). ¹H-NMR (DMSO-d₆) δ : 6.32 (2H, b, NH₂), 4.53 (1H, s, H-5), 3.59 (2H, t, EtCH₂N, *J* = 7.3 Hz), 1.46 (2H, m, MeCH₂CH₂N), 0.80, (3H, t, CH₃(CH₂)₂N, *J* = 7.3 Hz); ¹³C-NMR (DMSO-d₆) δ : 163.7 (C-4), 154.5 (C-2), 151.9 (C-6), 74.7 (C-5), 40.2 (EtCH₂N), 21.5 (MeCH₂CH₂N), 11.7 (CH₃CH₂CH₂N); IR $\bar{\nu}_{\max}$: 2930, 1741, 1631; UV $\lambda_{\max}(\epsilon)$: 261(3500).

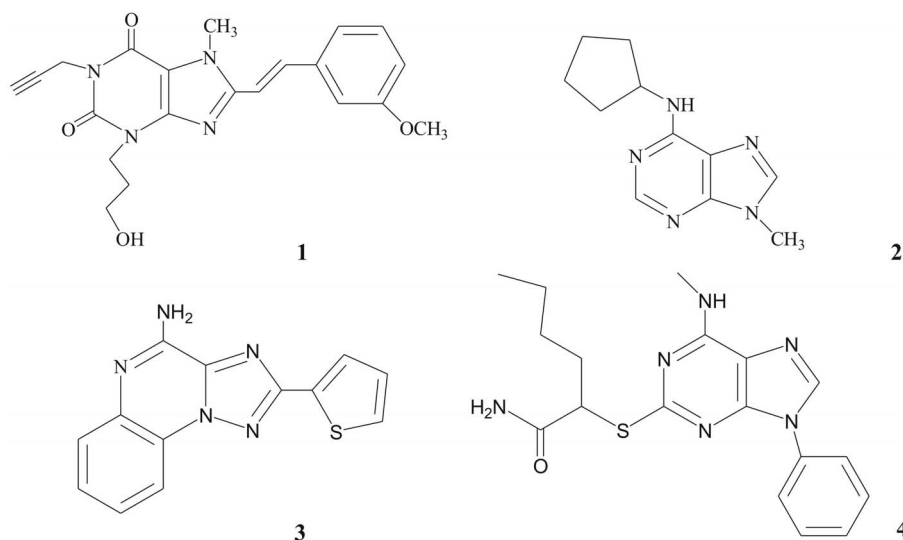


Figure 1. Compounds showing selectivity towards A_{2A} receptors.

2.1.2. Synthesis of 6-Amino-5-nitroso-3-propyluracil (7)

The uracil **6** (0.84 g, 5 mmol) was dissolved in 80% aq AcOH (10 mL)/MeOH (15 mL) warm solution. The solution was cooled down to room temperature and a solution of NaNO₂ (0.69 g, 10 mmol) in water (5 mL) was added drop-wise. The yellow precipitate was collected by filtration and washed with water (2 × 20 mL) and diethyl ether (2 × 20 mL). Yield: 85%; mp > 300°C; ¹H-NMR (DMSO-d₆) δ: 11.37 (1H, b, H-N), 3.79 (2H, t, EtCH₂N, *J* = 7.3), 1.60 (2H, m, MeCH₂CH₂N), 0.89 (3H, t, CH₃CH₂CH₂N, *J* = 7.3); NMR ¹³C (DMSO-d₆) δ: 161.8 (C-4), 150.3 (C-2), 145.5 (C-6), 140.3 (C-5), 40.2 (EtCH₂N), 21.4 (MeCH₂CH₂N), 11.8 (CH₃CH₂CH₂N); IR $\bar{\nu}_{\max}$: 3294, 3185, 1511; UV $\lambda_{\max}(\epsilon)$: 320(5920).

2.1.3. Synthesis of 5,6-Diamino-3-propyluracil (8)

The nitroso compound **7** (10 mmol, 1.98 g) was dissolved in 6% aq NH₄OH (40 mL) at 80°C, then Na₂S₂O₃ (30 mmol, 3.5 g) was added in small portions. The solution decolorizes rapidly. The warm crude was concentrated under reduced pressure; a precipitate is produced. After filtration, the white precipitate was washed with a small amount of cold water (20 mL). Compound **8** is very sensitive to air and to CDCl₃ solutions, so it was immediately used in the following step. Yield: 69%.

2.1.4. Synthesis of (E)-6-amino-3-propyl-5-styrylcarboxamidouracil (9)

A suspension of the diamino compound **8** (7 mmol, 1.28 g), EDCl (7 mmol, 1.34 g), and (E)-cinnamic acid (7 mmol, 1.03 g) in CH₃OH (20 mL) was stirred at room temperature overnight. The precipitate obtained was collected by filtration, washed with MeOH, water, and diethyl ether. Yield: 76%; mp > 300°C; ¹H-NMR (DMSO-d₆) δ: 8.7 (1H, b, HN in styrylamido group), 7.41 (6H, m, aromatics and H-vinyl), 6.85 (1H, d, vinyl, *J*_{trans} = 16), 6.11 (2H, b, -NH₂), 3.6 (2H, t, CH₃CH₂CH₂N, *J* = 7.3), 1.5 (2H, m, CH₃CH₂CH₂N), 0.81 (3H, t, CH₃CH₂CH₂N, *J* = 7.3); ¹³C-NMR (DMSO-d₆) δ: 165.6 (C=O in styrylamido group), 161.2 (C-4), 150.5 (C-6), 150.4 (C-2), 139.4 (Ph-HC=), 135.5 (C-*ipso*), 130.0 (C-*meta*), 129.6 (C-*orto*), 128.0 (C-*para*), 122.9 (=CH-), 87.7 (C-5), 40.2 (EtCH₂N), 21.5 (MeCH₂CH₂N), 11.8 (CH₃CH₂CH₂N); IR $\bar{\nu}_{\max}$: 3318, 3187, 1648; UV $\lambda_{\max}(\epsilon)$: 280(6670).

2.1.5. General Procedure for the Preparation of 1-Substituted (E)-6-amino-3-propyl-5-styrylcarboxamidouracils 10a-d

Compound **9** (1 mmol, 0.314 g) was dissolved in DMF (3.5 mL), then K₂CO₃ (1.1 mmol, 0.16 g) and the alkyl halide (1.1 mmol) were added. The mixture was stirred at

room temperature for 48 h and the progress of the reaction was monitored by TLC (eluent: acetone/hexane, 2:3). After addition of water (30 mL) a precipitate was produced, which was collected by filtration and washed with water (2 × 20 mL) and dried in a vacuum desiccator.

1) (E)-6-amino-1-ethyl-3-propyl-8-styrylcarboxamidouracil (10a)

Yield: 78%; mp 284°C - 286°C; ¹H-NMR (CDCl₃) δ: 8.04 (1H, b, NH), 7.63 (1H, d, -CH=), 7.36 (5H, m, aromatics), 6.70 (1H, d, -CH=), 5.83 (2H, b, -NH₂), 4.28 (2H, q, CH₃CH₂N), 4.12 (2H, t, CH₃CH₂CH₂N), 1.79 (2H, m, CH₃CH₂CH₂N), 1.42 (3H, t, CH₃CH₂N); 0.99 (3H, t, CH₃CH₂CH₂N); ¹³C NMR (CDCl₃) δ: 166.0 (C=O styrylamido), 160.3 (C-4), 152.5 (C-6), 150.7 (C-2), 141.6 and 119.8 (-CH=CH-), 134.3, 129.6, 128.5, 127.7 (aromatics), 95.4 (C-5), 43.4 (CH₃CH₂CH₂N), 39.1 (CH₃CH₂N), 21.1 (CH₃CH₂CH₂N), 13.5 (CH₃CH₂N), 11.7 (CH₃CH₂CH₂N); IR $\bar{\nu}_{\max}$: 3165, 1734, 1651, 968; UV (EtOH) $\lambda_{\max}(\epsilon)$: 278(11620).

2) (E)-6-amino-1,3-dipropyl-8-styrylcarboxamidouracil (10b)

Yield: 75%; mp 272°C - 273°C; ¹H-NMR (CDCl₃) δ: 8.40 (1H, b, NH styrylamido), 7.54 and 6.74 (d, 1H, -CH=CH-, *J*_{trans} = 16.4 Hz), 7.31 (m, 5H, aromatics), 6.11 (b, 2H, -NH₂), 4.14 (m, 4H, both CH₃CH₂CH₂N), 1.86 (m, 2H, CH₃CH₂CH₂N), 1.78 (t, 2H, CH₃CH₂CH₂N, *J* = 7.6 Hz), 1.02 (t, 3H, CH₃CH₂CH₂N, *J* = 7.6 Hz), 0.96 (t, 3H, CH₃CH₂CH₂N, *J* = 7.6 Hz); ¹³C NMR (CDCl₃) δ: 166.0 (C=O styrylamido), 159.8 (C-4), 149.9 (C-6), 147.1 (C-2), 141.7 and 119.9 (-CH=CH-), 134.5, 129.7, 128.8, and 127.8 (aromatics), 89.7 (C-5), 43.8 (CH₃CH₂CH₂N), 43.5 (CH₃CH₂CH₂N), 21.6 (CH₃CH₂CH₂N), 20.2 (CH₃CH₂CH₂N), 13.9 (CH₃CH₂CH₂N), 11.5 (CH₃CH₂CH₂N); IR $\bar{\nu}_{\max}$: 3024, 2962, 1735, 1705, 1651, 972; UV (EtOH) $\lambda_{\max}(\epsilon)$: 279(9130).

3) (E)-6-amino-1-butyl-3-propyl-8-styryluracil (10c)

Yield: 66%; mp 273°C - 274°C; ¹H-NMR (CDCl₃) δ: 8.14 (b, 1H, N-H styrylamido), 7.55 (d, 1H, Ph-CH=, *J* = 15.3 Hz), 6.76 (d, 1H, =CH-, *J* = 15.3 Hz), 7.30 (m, 5H, arom.), 5.82 (b, 2H, -NH₂), 3.78 (m, 4H, CH₃CH₂CH₂N and CH₃CH₂CH₂CH₂N), 1.58 (m, 4H, CH₃CH₂CH₂N and CH₃CH₂CH₂CH₂N), 1.36 (m, 2H, CH₃CH₂CH₂CH₂N), 0.91 (t, 3H, CH₃CH₂CH₂N, *J* = 7.3 Hz), 0.88 (t, 3H, CH₃CH₂CH₂CH₂N, *J* = 7.3 Hz); ¹³C NMR (CDCl₃) δ: 165.9 (C=O styrylamido), 160.1 (C-4), 160.1 (C-4), 150.1 (C-6), 148.5 (C-2), 142.2 and 119.9 (CH=CH), 134.5, 130.1, 128.9, and 128.0 (aromatics), 92.1 (C-5), 43.4 (CH₃CH₂CH₂CH₂N), 43.2 (CH₃CH₂CH₂N), 30.1 (CH₃CH₂CH₂CH₂N), 21.6 (CH₃CH₂CH₂CH₂N), 21.2 (CH₃CH₂CH₂N), 13.8 (CH₃CH₂CH₂N), 11.4 (CH₃CH₂CH₂CH₂N); IR $\bar{\nu}_{\max}$:

3319, 1731, 1652, 969; UV (EtOH) $\lambda_{\max}(\epsilon)$: 279(8525).

4) (E)-6-amino-1-pentyl-3-propyl-8-styryluracil (10d)

Yield: 70%; mp 258°C - 260°C; ¹H-NMR (CDCl₃) δ : 8.04 (b, 1H, N-H styrylamido), 7.63 y 6.70 (d, 1H, CH=CH, J_{trans} = 15.6 Hz), 7.35 (m, 5H, aromatics), 5.08 (b, 2H, -NH₂), 4.28, (t, 2H, BuCH₂N, J = 7.2 Hz), 4.12 (t, 2H, CH₃CH₂CH₂N, J = 7.6 Hz), 1.79 (m, 4H, CH₃CH₂CH₂N and PrCH₂CH₂N, J = 7.6 Hz), 1.42 (t, 3H, CH₃CH₂CH₂N, J = 7.2 Hz); 0.97 (sx, 2H, MeCH₂CH₂CH₂N), 0.87 (t, 3H, CH₃CH₂CH₂CH₂N, J = 7.6 Hz); ¹³C NMR (CDCl₃) δ : 166.0 (C=O styrylamido), 160.35 (C-4), 152.5 (C-6), 150.8 (C-2), 141.7 and 119.8 (CH=CH), 134.3, 129.6, 128.5, and 127.7 (aromatics), 95.4 (C-5), 43.4 (CH₃CH₂CH₂N), 39.2 (BuCH₂N), 21.1 (CH₃CH₂CH₂N), 18.5 (PrCH₂CH₂N), 13.5 (EtCH₃CH₂CH₂N), 11.7 (CH₃CH₂CH₂N) 11.4 (CH₃CH₂CH₂CH₂CH₂N); IR $\bar{\nu}_{\max}$: 3165, 1734 1651, 968; UV (EtOH) $\lambda_{\max}(\epsilon)$: 278(11620).

2.1.6. General Procedure for the Preparation of 3-Substituted (E)-1-propyl-8-styrylxanthines (11a-d)

Compounds **10a-d** (1 mmol) were dissolved in a mixture of CH₃OH (15 mL) and 20% aq KOH (5 mL) and solutions were refluxed for 3 h. The reaction was followed by TLC (eluent: acetone: hexane 2:3) and UV spectroscopy. After addition of conc. HCl (10 mL) a precipitate was obtained. The suspension was concentrated under vacuum. The precipitate was collected by filtration and washed with a small amount of cold water and MeOH.

1) (E)-3-ethyl-1-propyl-8-styrylxanthine (11a)

Yield: 65%; mp 277°C - 278°C (MeOH); ¹H-NMR (CDCl₃) δ : 13.60 (bs, 1H, N-H), 7.67 and 7.07 (d, 1H, CH=CH, J_{trans} = 16.4 Hz), 7.42 (m, 5H, aromatics), 4.07 (q, 2H, MeCH₂N, J = 7.4 Hz), 3.85 (t, 3H, EtCH₂N, J = 7 Hz), 1.58 (m, 2H, MeCH₂CH₂N), 1.25 (t, 3H, CH₃CH₂N, J = 7 Hz), 0.87 (t, 3H, CH₃CH₂CH₂N, J = 7 Hz); ¹³C NMR (CDCl₃) δ : 154.5 (C-6), 151.0 (C-4), 150.2 (C-8), 148.6 (C-2), 135.9 and 116.2 (CH=CH), 135.7, 129.7, 129.6, and 127.7 (aromatics), 107.7 (C-5), 42.7 (EtCH₂N), 38.8 (MeCH₂N), 21.4 (MeCH₂CH₂N), 13.7 (CH₃CH₂N), 11.8 (CH₃CH₂CH₂N); IR $\bar{\nu}_{\max}$: 3318, 3187, 1648; UV $\lambda_{\max}(\epsilon)$: 343(8660). Anal. Calcd for C₁₈H₂₀N₄O₂: C, 66.65; H, 6.21; N, 17.27; Found: C, 66.51; H, 6.09; N, 17.01.

2) (E)-1,3-dipropyl-8-styrylxanthine (11b)

Yield: 72%; mp 267°C - 269°C (MeOH); ¹H-NMR (CDCl₃) δ : 13.10 (bs, 1H, N-H), 7.80 and 7.13 (d, 1H, CH=CH, J_{trans} = 16.4 Hz), 7.40 (m, 5H, aromatics), 4.14 (m, 4H, both CH₃CH₂CH₂N), 1.85 (m, 2H, CH₃CH₂CH₂N¹), 1.76 (m, 2H, CH₃CH₂CH₂N³), 0.99 (m, 6H, both CH₃CH₂CH₂N); ¹³C NMR (CDCl₃) δ : 155.9 (C-6), 151.1 (C-4), 150.2 (C-8), 145.7 (C-2), 137.1 and

115.3 (CH=CH), 135.7, 129.4, 128.9, and 127.3 (aromatics), 107.2 (C-5), 45.5 (CH₃CH₂CH₂N¹), 38.8 (CH₃CH₂CH₂N³), 21.4 (both CH₃CH₂CH₂N), 11.5 (CH₃CH₂CH₂N¹), 11.3 (CH₃CH₂CH₂N³); IR (KBr) $\bar{\nu}_{\max}$: 3318, 3187, 2961, 1702, 1651; UV (EtOH) $\lambda_{\max}(\epsilon)$: 342(3585); Anal. Calcd for C₁₉H₂₂N₄O₂: C, 67.44; H, 6.55; N, 16.56; Found: C, 66.28; H, 6.48; N, 16.37.

3) (E)-3-butyl-1-propyl-8-styrylxanthine (11c)

Yield: 69%; mp 278°C - 279°C (MeOH); ¹H-NMR (CDCl₃) δ : 13.20 (bs, 1H, N-H), 7.77 and 7.09 (d, 1H, CH=CH, J_{trans} = 16.4 Hz), 7.36 (m, 5H, aromatics), 4.19 (t, 2H, PrCH₂N, J = 7.4 Hz), 4.11 (t, 2H, EtCH₂N, J = 7.4 Hz), 1.80 (m, 4H, EtCH₂CH₂N and MeCH₂CH₂N), 1.43 (m, 2H, MeCH₂CH₂CH₂N), 0.99 (m, 6H, both CH₃CH₂CH₂CH₂N); ¹³C NMR (CDCl₃) δ : 156.0 (C-6), 151.4 (C-4), 150.9 (C-8), 149.9 (C-2), 137.0 and 115.4 (CH=CH), 135.7, 129.4, 128.9, and 127.3 (aromatics), 107.4 (C-5), 43.8 (PrCH₂N), 43.6 (EtCH₂N), 30.2 (EtCH₂CH₂N), 21.5 (MeCH₂CH₂CH₂N), 20.1 (MeCH₂CH₂N), 13.9 (CH₃CH₂CH₂CH₂N), 11.5 (CH₃CH₂CH₂N); IR (KBr) $\bar{\nu}_{\max}$: 3318, 2958, 1702, 1649; UV (EtOH) $\lambda_{\max}(\epsilon)$: 342(8785); Anal. Calcd for C₂₀H₂₄N₄O₂: C, 68.16; H, 6.86; N, 15.90; Found: C, 68.03; H, 6.89; N, 15.77.

4) (E)-3-pentyl-1-propyl-8-styrylxanthine (11d)

Yield: 66%; 224°C - 226°C; ¹H-NMR (CDCl₃) δ : 13.57 (bs, 1H, N-H), 7.63 and 7.06 (d, 1H, CH=CH, J_{trans} = 16.4 Hz), 7.40 (m, 5H, aromatics), 4.01 (t, 2H, BuCH₂N, J = 7 Hz), 3.85 (t, 2H, EtCH₂N, J = 7 Hz), 1.70 (m, 2H, PrCH₂CH₂N), 1.56 (m, 2H, MeCH₂CH₂N), 1.31 (m, 4H, MeCH₂CH₂CH₂CH₂N), 0.87 (m, 6H, CH₃(CH₂)₄N and CH₃(CH₂)₂N); ¹³C-NMR (CDCl₃) δ : 154.4 (C-6), 151.2 (C-4), 150.1 (C-8), 148.9 (C-2), 138.9 and 116.4 (CH=CH), 135.7, 129.7, 129.5, and 127.7 (aromatics), 107.6 (C-5), 43.4 (BuCH₂N), 42.7 (EtCH₂N), 28.8 (PrCH₂CH₂N), 27.7 (EtCH₂(CH₂)₂N), 22.3 (MeCH₂(CH₂)₃N), 21.4 (MeCH₂CH₂N), 14.4 (CH₃CH₂CH₂N), 11.7 (CH₃(CH₂)₄N); IR (KBr) $\bar{\nu}_{\max}$: 3318, 2958, 1702, 1649; UV (EtOH) $\lambda_{\max}(\epsilon)$: 347(5520); Anal. Calcd for C₂₁H₂₆N₄O₂: C, 68.82; H, 7.15; N, 15.28; Found: C, 68.70; H, 7.01; N, 15.15.

2.2. Binding Studies

Radioligand binding studies to determine the relative affinity of compounds **11a-d** to A₁ and A_{2A} adenosine receptors were made using crude membrane preparations of rat cortical membrane and rat striatal membrane as previously described [29]. [³H]-2-chloro-N⁶-cyclopentyladenosine ([³H]CCPA) was used as A₁ receptor ligand and [³H]-(-)-3-(3-hydroxypropyl)-8-(3-methoxystyryl)-7-ethyl-1-propargylxanthine (MSX-2) as A_{2A} receptor ligand. Radiolabelled compounds were purchased from NEN Life Sciences, Dreieich, Germany.

2.3. Rotational Behavior in Rats with a Unilateral Lesion Caused by 6-Hydroxydopamine

All procedures described in this study were approved by the BUAP Animal Care Committee and met governmental guidelines (Mexican Council for Animal Care, Norma.

Oficial Mexicana NOM-062-ZOO-1999). Male Wistar rats (from Bioterio Claude Bernard, BUAP) weighed 250 - 350 g at the beginning of the experiment. Animals were housed in pairs in ventilated sound-attenuated rooms under a 12 h light:dark cycle at an ambient temperature with free access to water and food. The animals were deeply anaesthetized with choral hydrate (350 mg/kg ip) and received an injection of 2 μ L 6-hydroxydopamine (6-OHDA) over a period of 10 min (8 μ g/ μ L dissolved in 0.1% ascorbic acid) into the left *substantia nigra pars compacta* (SNc), according to the atlas of Paxinos and Watson 1998 [30] (stereotaxic coordinates; A = 4.0 mm anterior to bregma, L = 1.3 mm side lateral to the midline, V = 6.5 below the skull) to produce a unilateral dopamine depletion.

Two weeks after the lesion caused by the 6-OHDA the rats were evaluated with methamphetamine (METH, 7 mg/kg sc) starting 10 minutes after injection. To measure turning behavior the rats were placed in a plastic chamber and the number of complete (360°) turns, made over a period of 10 minutes, were counted. The mean number of ipsilateral rotations per minute was measured. This model is the one of the most reliable rodent models for the assessment of anti-Parkinsonian activity of the new drugs [31]. Only animals exhibiting at least 10 ipsilateral rotations/minute were used in subsequent experiments.

Ten days after the first evaluation of the turning behavior, the animals were administrated with (*E*)-3-ethyl-1-propyl-8-styrylxanthine (A15Et, **11a**) or (*E*)-3-butyl-1-propyl-8-styrylxanthine (A15Bu, **11c**) at dose of 1 mg/kg sc or with vehicle (mineral oil + DMSO, 0.1 ml/100 g sc), and 10 min later the animals received METH (7 mg/kg sc) and a second turning behavior was evaluated over a period of 80 min.

2.4. Statistical Analysis

The results were expressed as the mean \pm SEM. The statistical analysis for turning behavior was done using *t* of student, with *p* < 0.05 consider significant.

3. Results

3.1. Chemistry

The synthesis of the compounds **11a-d** was accomplished following the general pathway [25,32,33] depicted in the **Scheme 1**, optimizing all involved reactions.

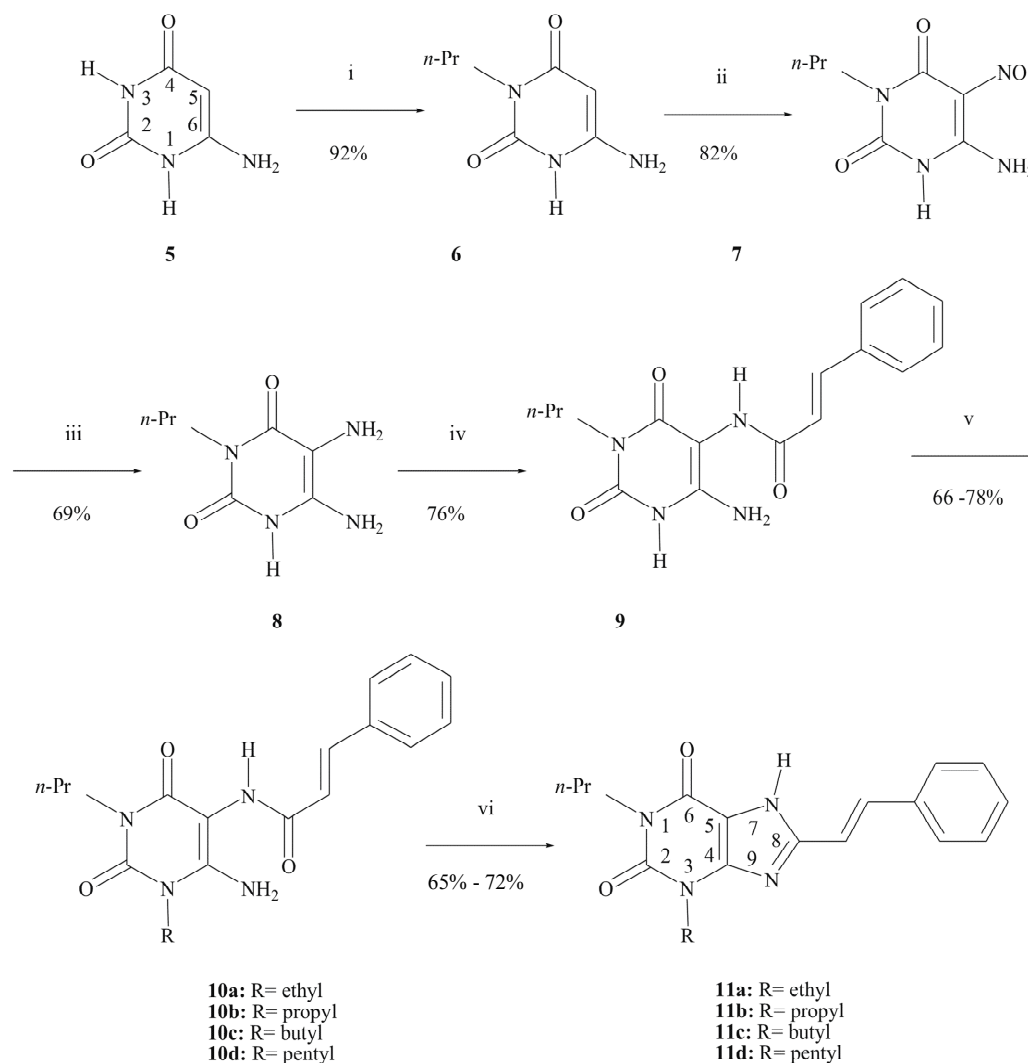
The alkylation of the N atom at position 3 in the 6-aminouracil (**5**) was highly selective using the known "one pot" procedure [32]: trisilylation of **5** by means of hexamethyldisilazane (HMDS), followed by a typical N-alkylation using an alkyl iodide. We have obtained excellent yields using small quantities of HMDS. The identification of the 3-substituted 6-aminouracil was carried out through the analysis of the chemical shift of the signal for NH₂ protons in the ¹H-NMR spectrum: in the 6-amino-1-propyluracil the NH₂ protons are shifted downfield with respect of those in 6-aminouracil; in the 6-amino-3-propyluracil the NH₂ protons are not influenced [34]. The UV spectrum showed the classical absorption of a β -amino α,β -unsaturated carbonyl (λ_{\max} 261 nm).

In the preparation of the 6-amino-5-nitroso-3-propyluracil (**7**) better results were obtained when only 2 eq of sodium nitrite were used, working at room temperature. The yellow compound **7** (λ_{\max} 320 nm) was isolated in 85% yield. In the ¹³C-NMR spectrum the signal for C-5 appears at 140.3 ppm, down-shifted in comparison to that in the starting material (74.6 ppm), caused by the electron-withdrawing effect of the nitroso group. In the IR spectra the characteristic band for the N=O group is shown at 1510 cm⁻¹.

The nitroso compound **6** was chemoselectively reduced by sodium dithionite in alkaline solution to the corresponding 5,6-diamino-3-propyluracil (**8**). It was easily isolated and was used for the next step without further purification because of its instability to light, oxygen and acidic medium. The chemoselective reduction was carried out in the range from 40°C to 80°C; we found that at lower temperatures the reaction needs a huge excess of reducing agent, causing difficulties in the purification of the diamino compound **8**. For this reaction, we additionally examined the influence of the concentration of ammonia. All concentrations used (24%, 12%, 6% and 3%) led successfully to the diamino compound **8** in about 70% yield.

The condensation of the diamino compound **8** with cinnamic acid to form (*E*)-6-amino-3-propyl-5-styrylcarboxamidouracil (**9**) was achieved using N-ethyl-N'-[3-(dimethylamino)propyl]carbodiimide (EDC) as the coupling agent. The configuration of the double bond in the styryl moiety was corroborated by its ¹H-NMR spectrum: the doublet signal at 6.85 ppm, assigned to vinyl proton, has a typical *J*_{trans} = 16 Hz. The alkylation at N¹ of the uracil derivative **9** could easily be performed under mild conditions. A DMF solution of **9**, treated with K₂CO₃ and an alkyl bromide or iodide, yielded the 1-substituted (*E*)-6-amino-3-propyl-5-styrylcarboxamidouracil **10a-d** in good yields. UV absorptions are in the range of 278 - 280 nm. No alkylation at position 7 was observed.

Finally, an alkaline treatment promoted the cyclization



Reagents: i. HMDS, (NH₄)₂SO₄, n-Pr-I; ii. NaNO₂, AcOH; iii. Na₂S₂O₄, NH₄OH; iv. C₉H₈O₂, CH₃OH; v. R-X, DMF, K₂CO₃; vi. KOH, CH₃OH.

Scheme 1. Synthetic pathway for the styrylxanthines 11a-d.

between the amino and carboxamido groups to provide compounds **11a-d**, in around 70% yield. The UV spectra showed bands in the range $\lambda_{\max} = 342 - 347$ nm, characteristic for 8-styrylxanthines.

3.2. Binding Studies

The results of binding studies showed that all the 1,3-disubstituted 8-styrylxanthines synthesized have affinity to both adenosinergic receptors. **Table 1** shows the different affinities of each compound analyzed; it is worth to notice that gradual increase in the length of the carbon chain of the alkyl group at the N atom at position 3 increases the affinity towards A₁ receptors. However, opposite effect is observed for affinity toward A_{2A} receptors.

3.3. Behavioral Test Method

In these conditions the animals showed a clear increase in the turning behavior 14.21 ± 3.206 minutes after METH injection. The difference was stable over a period of 80 minutes with a significant decrease of 1.938 ± 0.6003 minutes respect to group that only received to vehicle (mineral oil + DMSO; 1:1). Animals that only received vehicle did not show deficit in movement initiation or speed of turning (**Figure 2**).

Based on binding studies we evaluated the effect on turning behavior of (*E*)-3-ethyl-1-propyl-8-styrylxanthine (A15Et, **11a**), compound that showed highest affinity towards A_{2A} receptor and also (*E*)-3-butyl-1-propyl-8-styrylxanthine (A15Bu, **11c**) with approximately half affinity to A_{2A} receptor (see **Table 1**).

Table 1. Adenosine receptor affinities of 3-substituted (*E*)-1-propyl-8-styrylxanthines.

Compound	K _i (nM)	
	A ₁ Affinity rat brain cortical membranes [³ H]CCPA	A _{2A} Affinity rat brain striatal membranes [³ H]MSX-2
11a	14.50 ± 1.50 (n = 2)	12.5 ± 0.5 (n = 2)
11b	13.67 ± 0.88 (n = 3)	12.97 ± 1.5 (n = 3)
11c	7.05 ± 0.25 (n = 2)	24.0 ± 8.0 (n = 2)
11d	6.30 ± 1.10 (n = 2)	26.0 ± 2 (n = 2)

K_i [nM] ± SEM or % displacement at concentration indicated.

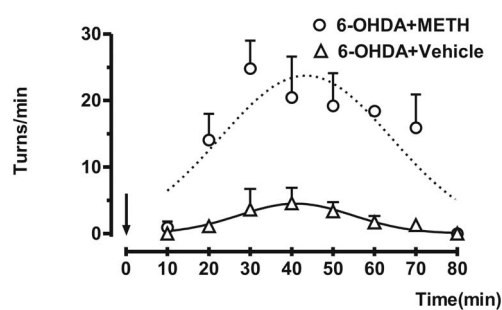


Figure 2. The ipsilateral turn behavior were induced by administration of METH (7 mg/kg sc) in rats with 6-OHDA. The turns/min were counted over 10 minutes. The arrow indicates the administration time. Each point represents the mean number of ipsilateral turns each 10 min ± SEM. Vehicle (Δ) and 6-OHDA (○).

The A15Et decreased the turning behavior and was administrated as a dose of 1 mg/Kg, 30 minutes before the administration of METH (7 mg/kg sc). This significant difference was maintained over a period of 50 minutes respect to the vehicle group (**Figure 3(a)**).

The A15Bu was administrated 30 min before the METH (7 mg/kg sc), decreased significantly the turning behavior and this difference was maintained over a period of 50 - 70 min respect to the vehicle group (**Figure 3(b)**).

4. Discussion

The administration of the A15Et (11a) or A15Bu (11c) effectively reduces the ipsilateral turning behavior caused by METH in rats with unilaterally lesions caused by 6-OHDA. These compounds can exert significant functional effects on motor behavior. Also we have found that rotational asymmetry in rats with unilateral lesions caused by 6-OHDA can be counteract by administration of compounds A15Et and A15Bu (**Figures 3(a)** and **(b)**). These effects were more potent and with more prolonged duration with A15Et, this effect was expected in order

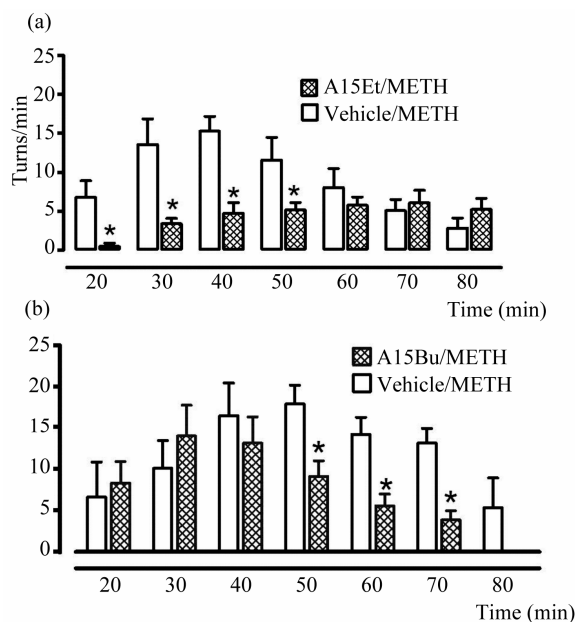


Figure 3. Effect of (a) A15Et and (b) A15Bu on number of ipsilateral turns in rats with unilateral lesions caused by 6-OHDA. Both novel xanthines were administrated at dose of 1 mg/kg sc 10 min before the administration of METH (7 mg/kg sc). Open bars represent ipsilateral turns in the vehicle group and hatched bars show the turns with derivates of xanthines. The data were given as the mean ± SEM. The statistical analysis was made using *t* of student test. * *p* < 0.05 vs Vehicle/METH group.

that A15Et showed highest affinity to A_{2A} receptor and this effect may be related to modulation of A_{2A} receptor over D₂ receptor. Recently it has been reported that other compounds possessing the property of adenosine A_{2A}-receptor antagonism [35,36], improved the motor effect in an experimental model of Parkinson's disease [37]. Because adenosine A_{2A} receptors have a profound influence on motor functions via the modulation of basal ganglia output pathways [37], blockade of this modulatory function by an A_{2A} antagonist could repair striatopallidal abnormal-neuronal activities caused by striatal dopamine depletion in the Parkinsonian state [37].

5. Conclusion

In summary, we describe the synthesis of the 1,3-disubstituted 8-styrylxanthines **11a-d** with optimized conditions and excellent yield. The results of the present study provide evidence that all compounds synthesized have affinity towards A₁ and A_{2A} receptors but compound A15Et showed the highest affinity towards A_{2A} receptor. The administration of the A15Et (11a) or A15Bu (11c) effectively reduces the ipsilateral turning behavior caused by METH in rats with unilaterally lesions caused by 6-OHDA. Based on these results we propose to A15Et as a potential compound to treat some symptoms of Par-

kinson's disease.

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