



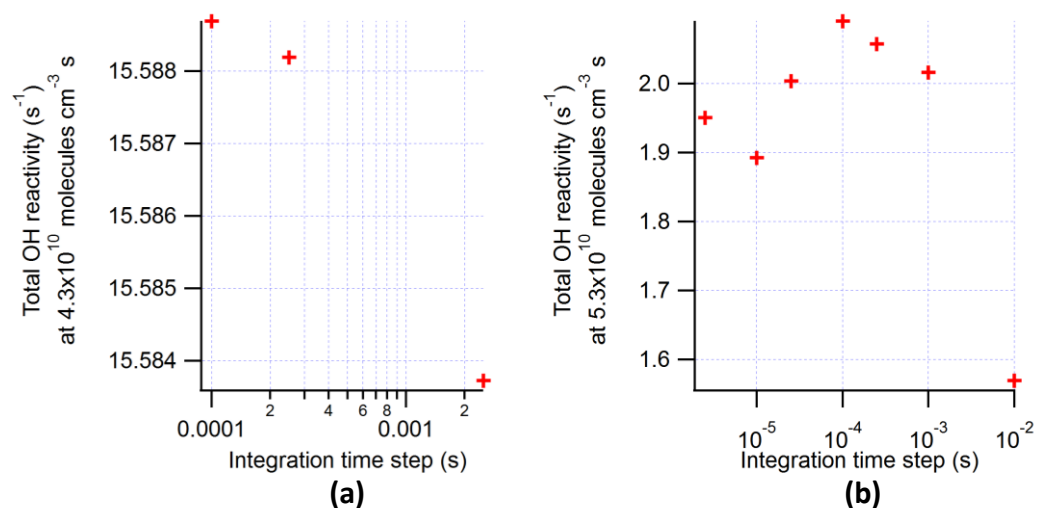
*Supplement of*

## **Evolution of OH reactivity in NO-free volatile organic compound photooxidation investigated by the fully explicit GECKO-A model**

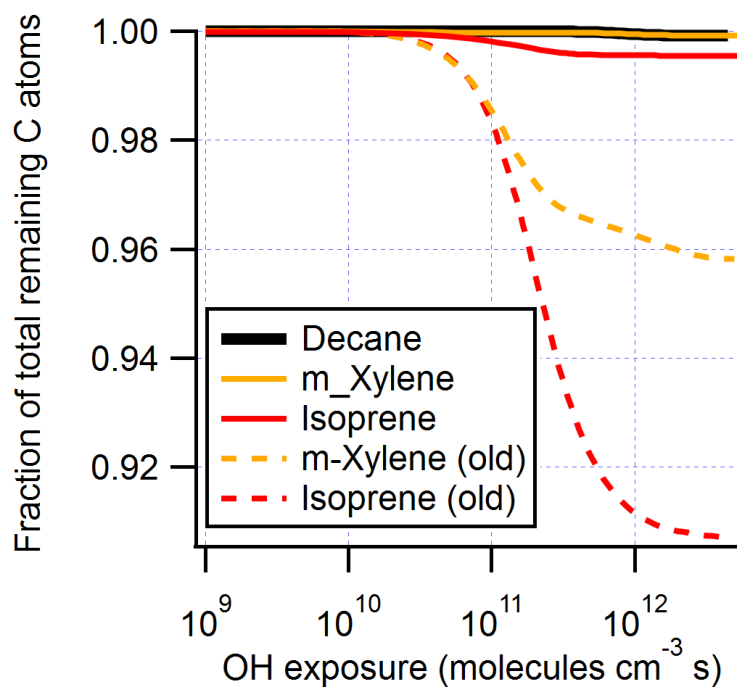
**Zhe Peng et al.**

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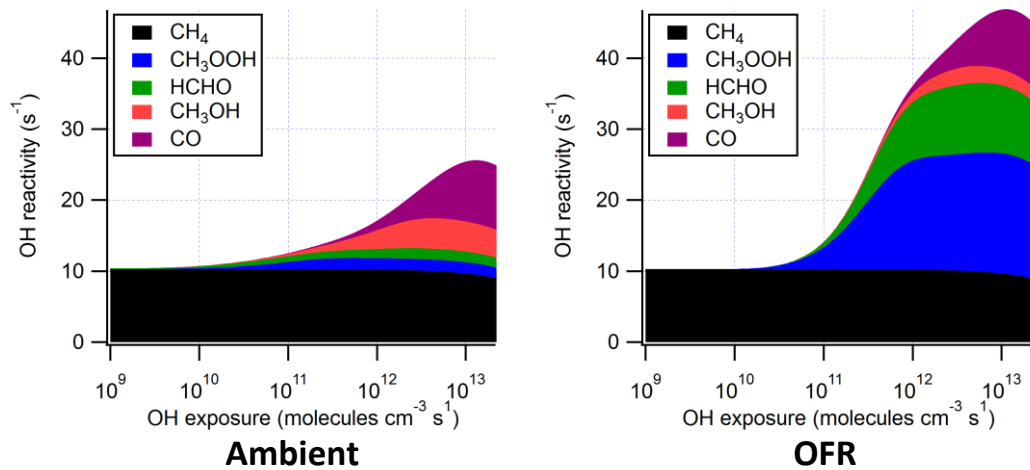
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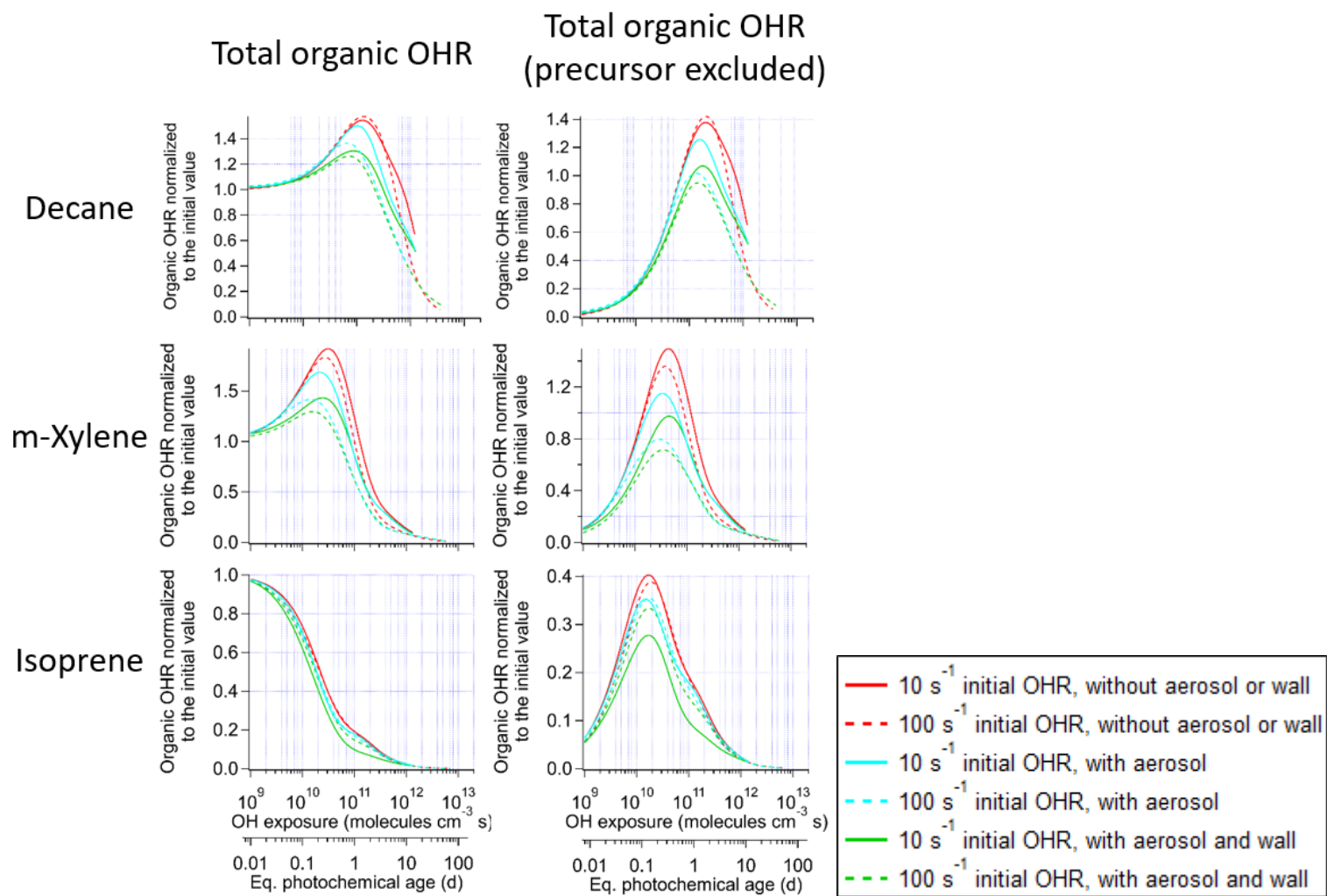
**Figure S1.** Total OH reactivity (OHR) of organics at OH exposure of (a)  $4.3 \times 10^{10}$  and (b)  $5.3 \times 10^{10}$  molecules  $\text{cm}^{-3} \text{s}$  as a function of integration timestep in the test simulations for the (a) m-xylene and (b) isoprene oxidation flow reactor (OFR) cases at relative humidity of 70%, high lamp setting, and initial OHR of  $10 \text{ s}^{-1}$ . Note that the mechanisms in these test simulations are similar but not exactly the same as in the corresponding model cases shown in Table 1.



**Figure S2.** Fractions of total remaining C atoms as a function of OH exposure in the ambient cases with constant sunlight of the photooxidations of decane, m-xylene, and isoprene. Also shown for comparison are the simulations for m-xylene and isoprene with the old mechanisms whose problem of non-conservation of C atoms in some reactions has not been fixed.



**Figure S3.** OH reactivity (OHR) of the organics as a function of OH exposure in the ambient case with constant sunlight; and in the OFR case at relative humidity of 70% and high lamp setting, both with initial OHR of 10 s<sup>-1</sup> of methane photooxidation.



**Figure S4.** Same format as Fig. 1, but for the chamber cases without aerosol or wall, with aerosol (no wall), and with aerosol and wall for decane, m-xylene, and isoprene photooxidation.

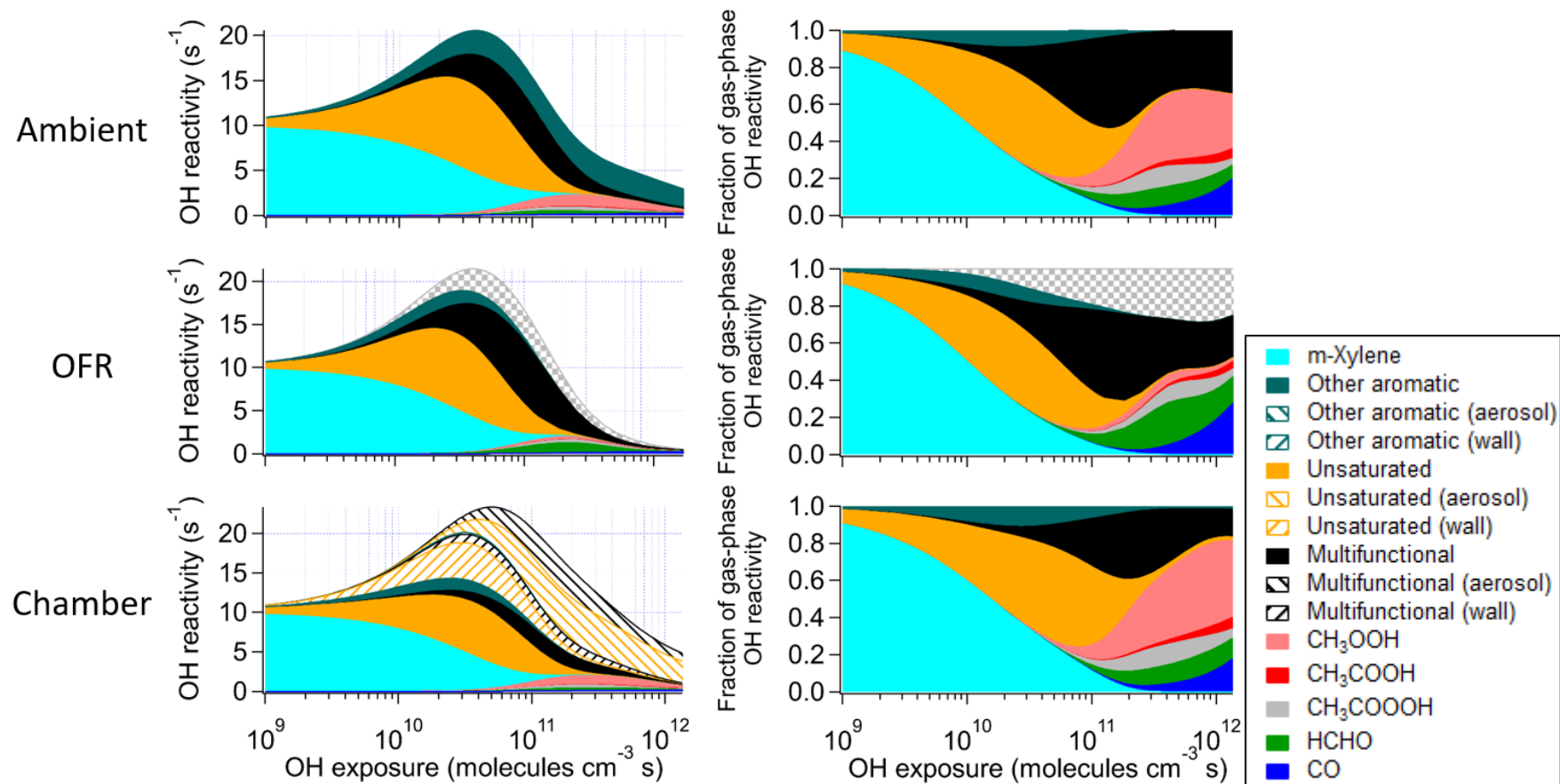
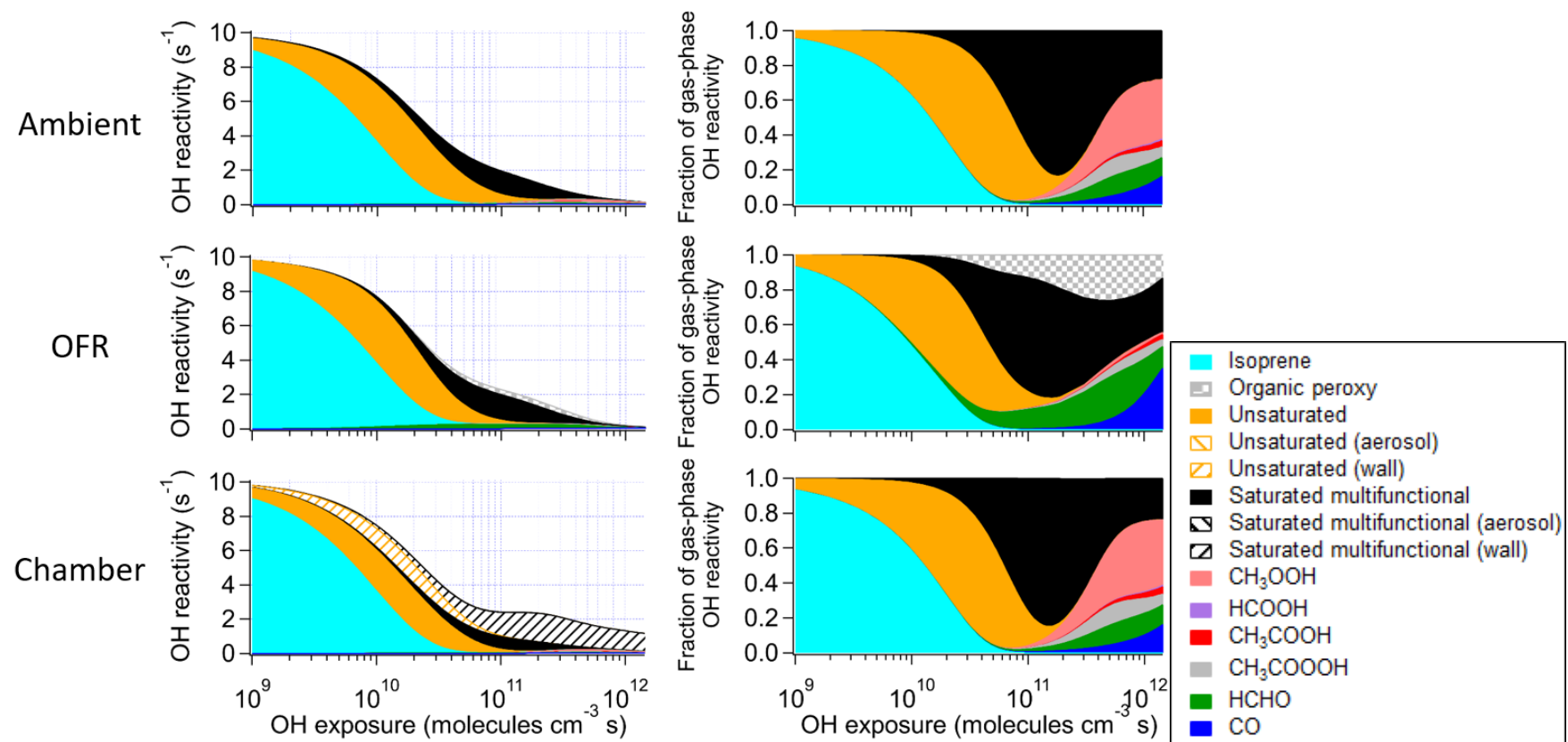
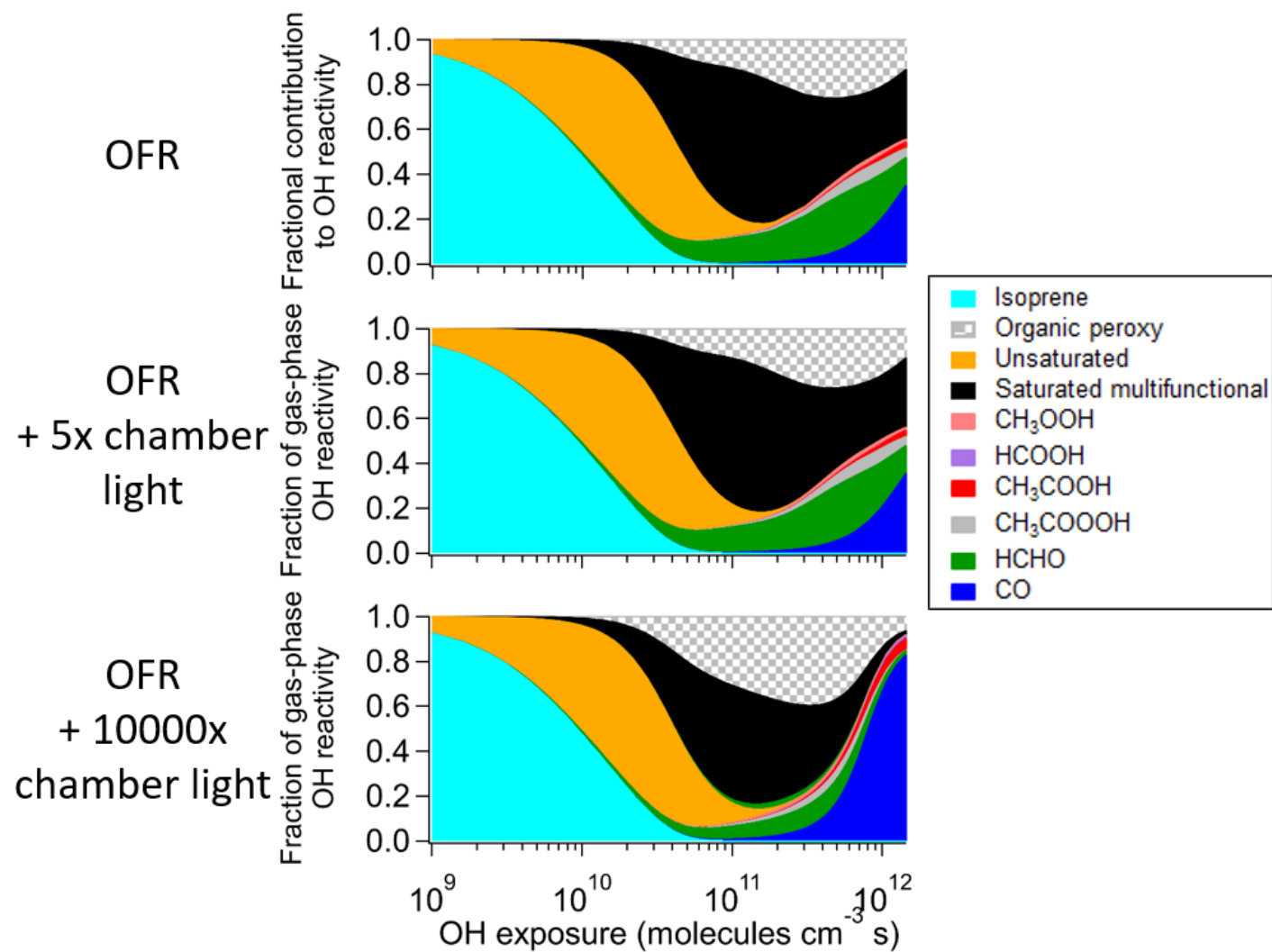


Figure S5. Same format as Fig. 4, but for m-xylene photooxidation.



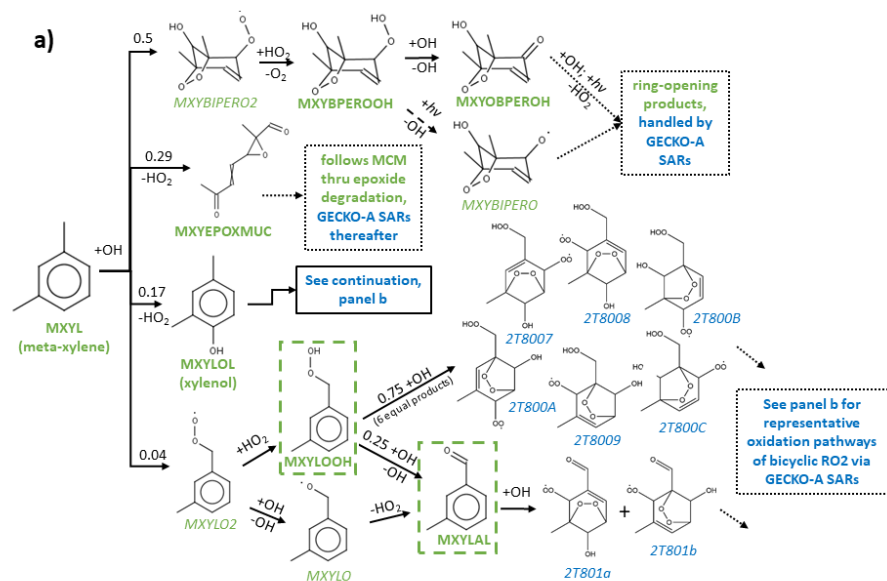
**Figure S6.** Same format as Fig. 4, but for isoprene photooxidation.



**Figure S7.** Fractional contributions of the main species and types of species to the organic OHR as a function of OH exposure in the standard OFR case of isoprene photooxidation with relative humidity of 30%, medium UV lamp setting, and two sensitivity cases based on this OFR case, with additional UV irradiation corresponding to 5 and 10000 times the UV source of the University of Colorado Environmental Chamber Facility, respectively. The types of species shown in this figure exclude the C1 and C2 species listed separately.

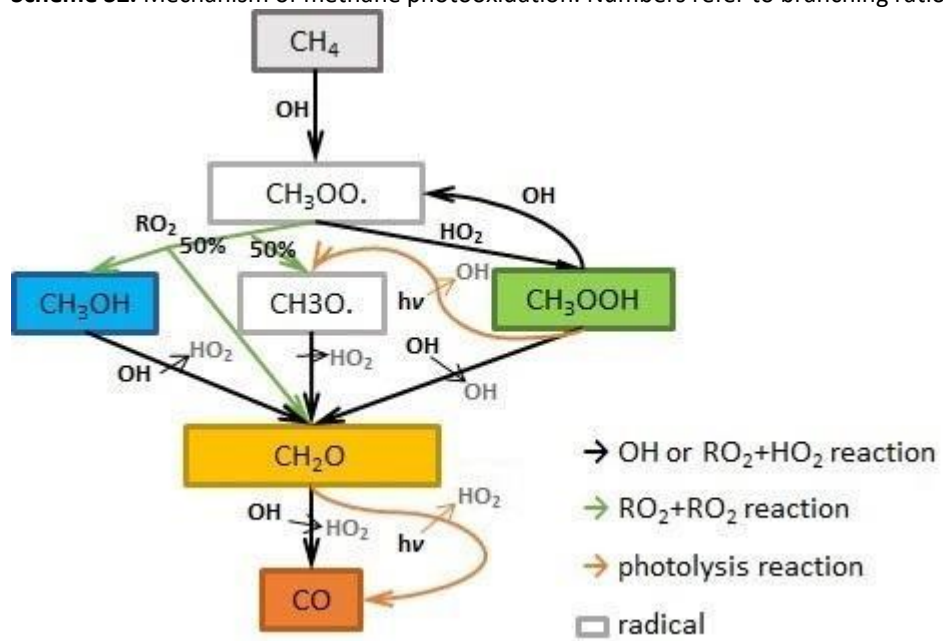


**Scheme S1.** Mechanism of xylenol photooxidation in the absence of NO, derived from MCM v3.2 (Jenkin et al., 2003; Bloss et al., 2005). Panel a): All meta-xylene reaction channels; Panel b): Xylenol reaction channels, expanded. Numbers refer to branching ratios. Green text indicates products from MCM v3.2; blue text indicates products generated by GECKO-A and added in this study; blue-and-green indicates products provided by both MCM3.2 and GECKO-A. Species whose products persist anomalously under zero-NO conditions are indicated with green dashed borders. Short dotted arrows indicate chemistry continuing according to standard SARs. Dashed arrows indicate photolysis pathways (which are usually minor). For illustration, we extend the dominant (HO<sub>2</sub> and higher branching ratio OH) oxidation pathway of one of the most prevalent RO<sub>2</sub> products (2T800J: panel b, lower center to upper right). Its main pathway from initial OH oxidation (lower right, 2nd row up) is also extended to show an example of typical ring-breaking chemistry. Net OH rate constants for MXYLOOH, MXY1OOH, MXYCATECH, and MXYLAL are 1.77x10<sup>-11</sup>, 3.26x10<sup>-11</sup>, 1.56x10<sup>-10</sup> and 8.6x10<sup>-13</sup> cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> respectively. (The MXYLOOH OH rate includes MXYLAL production). Figure adapted from MCM website, <http://mcm.leeds.ac.uk/MCMv3.3.1>, accessed July 3, 2020.





**Scheme S2.** Mechanism of methane photooxidation. Numbers refer to branching ratios.



**Table S1.** Extension of reference absorption cross-sections ( $\sigma$ ) and branching ratios ( $\phi$ ) of organic photolysis to 185 nm and 254 nm

Species, channel	$\sigma_{185\text{nm}}$	$\phi_{185\text{nm}}$ (if $\neq 1$ )
CH <sub>2</sub> O	3.64×10 <sup>-18</sup> Cooper (1996) 184 nm	
→ H <sub>2</sub> + CO		0.44 Formula extrapolation, Röth (2015)
→ HCO + H		0.56 Formula extrapolation, Röth (2015)
CH <sub>3</sub> OOH	9.0×10 <sup>-19</sup> rough extrapolation from 210-280nm data <sup>a</sup>	
CH <sub>2</sub> (OH)OOH	9.0×10 <sup>-19</sup> estimate after CH <sub>3</sub> OOH	
CH <sub>3</sub> CHO	7.84×10 <sup>-20</sup> <sup>a</sup>	
CHOCHO	4.80×10 <sup>-19</sup> Zhu (1996) at 193nm	
→ H <sub>2</sub> + 2 CO		0.81 data extrapolation <sup>b</sup> (225nm)
→ 2 CHO		0.16 data extrapolation <sup>b</sup> (225nm)
→ CH <sub>4</sub> +CO		0.03 data extrapolation <sup>b</sup> (225nm)
CH <sub>2</sub> (OH)CHO	3.85×10 <sup>-18</sup> Karandunandan (2007) at 184.9nm	
CH(O)C(O)OH	4.0×10 <sup>-19</sup> rough extrapolation from Back & Yamamoto (1985) (200nm)	
→ HCHO + CO <sub>2</sub>		0.84
→ HCO + 2 CO		0.16
C(O)(OH)C(O)OH	1.0×10 <sup>-18</sup> estimate after CH(O)C(O)OH and CH <sub>3</sub> C(O)C(O)OH	
→ HC(O)OH + CO <sub>2</sub>		0.72 Yamamoto (1985) (255-309 nm)
→ HC(O)OH + CO <sub>2</sub>		0.27 Yamamoto (1985) (255-309 nm)
CH <sub>3</sub> CH <sub>2</sub> CHO	1.43×10 <sup>-17</sup> Lucazeau & Sandorfy (1970) (184.8 nm)	
CH <sub>3</sub> COCH <sub>3</sub>	2.96×10 <sup>-18</sup> Gierczak (2003) (184.9 nm, 242 K) [note: 296K value is 3.01×10 <sup>-18</sup> ]	
CH <sub>3</sub> COCHO	3.71×10 <sup>-18</sup> data extrapolation <sup>b</sup> (200-215 nm)	
→ CH <sub>3</sub> C(.)O + CHO.		0.90 Raber (1995) (220-320 nm)
→ CH <sub>3</sub> CHO + CO		0.05 Raber (1995) (220-320 nm)
→ CH <sub>4</sub> + 2 CO		0.05 Raber (1995) (220-320 nm)
CH <sub>3</sub> COCH <sub>2</sub> OH	5.40×10 <sup>-18</sup> Dillon (2006) (184.9 nm)	
		0.6 Orlando (1999) (236-340 nm)
CH <sub>3</sub> C(O)C(O)OH	1.0×10 <sup>-17</sup> rough extrapolation from JPL 10-6 (252-280 nm)	
		0.37 Moortgat (1999) (251-400 nm)
<hr/>		
Species, channel	$\sigma_{254\text{nm}}$	$\phi_{254\text{nm}}$
CHOCHO	1.60×10 <sup>-20</sup> Volkamer (2005) at 254nm	
→ H <sub>2</sub> + 2 CO		0.54 data interpolation <sup>b</sup>
→ 2 CHO		0.16 data interpolation <sup>b</sup>
→ CH <sub>4</sub> +CO		0.32 data interpolation <sup>b</sup>
C(O)(OH)C(O)OH	6.55×10 <sup>-20</sup> extrapolation from Yamamoto (1985) (255-260 nm)	
→ HC(O)OH + CO <sub>2</sub>		0.72 Yamamoto (1985) (255-309 nm)
→ CO <sub>2</sub> + CO + H <sub>2</sub> O		0.27 Yamamoto (1985) (255 -309 nm)

<sup>a</sup> Data from online spectral atlas: Keller-Rudek, H., Moortgat, G. K., Sander, R., and Sörensen, R., The MPI-Mainz UV/VIS spectral atlas of gaseous molecules of atmospheric interest, Earth Syst. Sci. Data, 5, 365–373, (2013)

<sup>b</sup> Data from JPL 10-6 (Sander et al, 2011)

**Table S2.** Major contributors (>0.1 s<sup>-1</sup> or the first 20) to the OH reactivity at 1x10<sup>10</sup>, 1x10<sup>11</sup>, and 1x10<sup>12</sup> molecules cm<sup>-3</sup> s in the ambient case with constant UV of the photooxidation of decane, m-xylene, and isoprene. Also shown is the percentage of OH reactivity accounted for by the species reported in this table. Positional isomers with identical rate constant with OH are lumped into a single species. See Scheme S1 for the structures of the special names for m-xylene oxidation in this table. The “C1-O-C1” string in the names represents an epoxide structure. The isoprene case has no species contributing >0.1 s<sup>-1</sup> to the total OH reactivity at 1x10<sup>12</sup> molecules cm<sup>-3</sup> s, and is thus not shown for this OH exposure.

Precursor	OH exposure (molecules cm <sup>-3</sup> s)	Species name	OH reactivity (s <sup>-1</sup> )	Percentage of OH reactivity accounted for
decane	1x10 <sup>10</sup>	decane	8.97	>99
		decyl hydroperoxide	1.95	
		dihydroperoxy decane	0.12	
	1x10 <sup>11</sup>	decyl hydroperoxide	5.06	94
		decane	3.35	
		dihydroperoxy decane	2.17	
		hydroperoxy decanone	1.65	
		decanone	1.18	
		dioxodecyl hydroperoxide	0.36	
		hydroxydecyl hydroperoxide	0.27	
		dioxodecane	0.14	
		hydroxydecyl dihydroperoxide	0.13	
		hydroperoxyhydroxydecanone	0.11	
	1x10 <sup>12</sup>	CH <sub>3</sub> OOH	0.88	53
		HCHO	0.63	
CH <sub>3</sub> CH <sub>2</sub> OOH		0.36		

		CH <sub>3</sub> C(O)OOH	0.31	
		CH <sub>3</sub> CHO	0.31	
		HOOCH <sub>2</sub> C(O)OOH	0.23	
		CH <sub>3</sub> C(O)CH <sub>2</sub> CH <sub>2</sub> OOH	0.23	
		CH <sub>3</sub> C(O)CH <sub>2</sub> OOH	0.21	
		HOOCH <sub>2</sub> CH <sub>2</sub> OOH	0.20	
		HOOCH <sub>2</sub> COOH	0.20	
		CO	0.19	
		HOOCH <sub>2</sub> CHO	0.17	
		CH <sub>3</sub> C(O)CH <sub>2</sub> CHO	0.17	
		HOOCH <sub>2</sub> CH <sub>2</sub> C(O)OOH	0.13	
		HOOCH <sub>2</sub> CH <sub>2</sub> COOH	0.12	
		CH <sub>3</sub> CH <sub>2</sub> C(O)CH <sub>2</sub> CH <sub>2</sub> OOH	0.11	
m-xylene	1x10 <sup>10</sup>	m-xylene	7.92	96
		MXYPEROOH	2.99	
		MXYEPOXMUC	1.41	
		MXYOBPEROH	1.13	
		m-xylenol	0.90	
		CH <sub>3</sub> C(O)CH(OOH)CH(OH)C1H-O-C1(CH <sub>3</sub> )CHO	0.34	
		dimethyl catechol	0.22	
		MXYOLOOH	0.17	

		methylglyoxal	1.04	
		m-xylene	0.96	
		CH <sub>3</sub> OOH	0.91	
		MXYOBEROH	0.81	
		MXYBEROOH	0.59	
		CH <sub>3</sub> C(OOH)(CHO) <sub>2</sub>	0.45	
		CH <sub>3</sub> C1(CHO)-O-C1HCHO	0.43	
		CH <sub>3</sub> C(O)CH=CHC1H-O-C1(CH <sub>3</sub> )CHO	0.39	
		HCHO	0.38	
		CH <sub>3</sub> C(O)CH(OOH)CH(OH)C1H-O-C1(CH <sub>3</sub> )CHO	0.34	
	1x10 <sup>11</sup>	CH <sub>3</sub> C(O)OOH	0.34	69
		HOC(O)C1H-O-C1(CH <sub>3</sub> )CHO	0.31	
		glyoxal	0.22	
		m-xyleneol	0.22	
		HOOC(O)CH=C(CH <sub>3</sub> )CHO	0.21	
		CH <sub>3</sub> C(O)CH=CHC(O)OOH	0.20	
		CH <sub>3</sub> C(O)C(O)CH(OH)C1H-O-C1(CH <sub>3</sub> )CHO	0.18	
		TT801J	0.17	
		TT8004	0.16	
		CH <sub>3</sub> O <sub>2</sub>	0.15	
	1x10 <sup>12</sup>	CH <sub>3</sub> OOH	0.48	56

		CO	0.19	
		HCHO	0.12	
isoprene	1x10 <sup>10</sup>	isoprene	3.68	93
		ISOPOOH	2.80	
		HOOCH <sub>2</sub> CH=C(CH <sub>3</sub> )CHO	0.23	
	1x10 <sup>11</sup>	ISOPOOH	0.37	36
		IEPOX	0.33	



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