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# Supplement of

# Constraints on global oceanic emissions of $N_2O$ from observations and models

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# Description of the PlankTOM10.2 equations

#### April 10, 2018

#### Introduction 1

This Supplement presents a full description of the PlankTOM10.2 model, a global marine biogeochemical model based on the representation of ten Plankton Functional Types (PFTs), including six phytoplankton (pPFTs), three zooplankton (zPFTs) and bacteria. PlankTOM10.2 also represents the full cycles of C, O<sub>2</sub>, P and Si and simplified cycles for Fe and N. This version comprises of 40 biogeochemical tracers (Table 1).

#### 1.0.1 Notation

In the following sections, we will show the equations governing tracer and food-web dynamics. These equations are mostly semi-empirical, and have been developed and tested using a multitude of laboratory and field data. As long as not otherwise indicated, both tracers and their respective concentrations will be designated by capital letters, with

- $P_i$ : concentration of pPFT<sub>i</sub> with  $i \in \{1, 6\}$ ,
- $Z_j$ : concentration of zPFT<sub>j</sub>, with  $j \in \{1, 3\}$ ,
- $F_k$ : concentration of food k; where  $F_k$  includes phytoplankton and other food sources
- PRO: proto-zooplankton concentration,
- MES: meso-zooplankton concentration,
- MAC: macro-zooplankton concentration,
- PO<sub>4</sub>: concentration of phosphate,
- NH4: concentration of total ammonium,
- NO3: concentration of nitrate,
- Fe: iron concentration, and
- Si: silicate concentration.

All concentrations are calculated in  $\frac{mol}{L}$  except for chlorophyll whic is in  $\frac{gCHL}{L}$ .

Tables and an index are provided which link the mathematical symbols with the variable names used in

the Fortran code.

Where subscript j includes pico-heterotrophs in addition to the three zoo-plankton types this is stated explicitly.

The ten plankton functional types and the tracers are shown in Figure 1. Figures of this type showing the processes governing the evolution of the PFTs and tracers are included in the following sections.

Table 1: List of biogeochemical Tracers in PlankTOM10.2

Abbreviation	Description	Units
ALK	alkalinity	${ m eq}~{ m L}^{-1}$
BAC	pico-heterotrophs	$\operatorname{mol} L^{-1}$
BFE	Fe in large POM	$\mathrm{mol}\ \mathrm{L}^{-1}$
BSI	biogenic particulate silica	$\mathrm{mol}\ \mathrm{L}^{-1}$
CAL	sinking CaCO <sub>3</sub>	$\mathrm{mol}\ \mathrm{L}^{-1}$
CCH	chlorophyll in calcifiers	${ m g}{ m L}^{-1}$
CFE	Fe in calcifiers	$\mathrm{mol}\ \mathrm{L}^{-1}$
COC	calcifying phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
DCH	chlorophyll in silicifiers	${ m g}{ m L}^{-1}$
DFE	Fe in silicifiers	$\operatorname{mol} L^{-1}$
DIA	silicifying phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
DIC	dissolved inorganic carbon	$\mathrm{mol}\ \mathrm{L}^{-1}$
DOC	dissolved organic carbon	$\mathrm{mol}\ \mathrm{L}^{-1}$
DSI	sinking particulate silica	$\mathrm{mol}\ \mathrm{L}^{-1}$
FER	dissolved iron	$\mathrm{mol}\ \mathrm{L}^{-1}$
FCH	chlorophyll in N <sub>2</sub> fixers	${ m g}{ m L}^{-1}$
FFE	Fe in N <sub>2</sub> fixers	$\mathrm{mol}\ \mathrm{L}^{-1}$
FIX	N <sub>2</sub> fixing phytoplankton	$ m mol~L^{-1}$
GOC	large particulate organic carbon	$\mathrm{mol}\ \mathrm{L}^{-1}$
НСН	chlorophyll in DMSP producers	$\mathrm{mol}\ \mathrm{L}^{-1}$
HFE	Fe in DMSP producers	$\mathrm{mol}\ \mathrm{L}^{-1}$
PIC	pico-phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
MES	meso-zooplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
MIX	mixed phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
N2O	prognostic nitrous oxide	$\mathrm{mol}\ \mathrm{L}^{-1}$
N2S	diagnostic nitrous oxide	$\mathrm{mol}\;\mathrm{L}^{-1}$
NCH	chlorophyll in mixed phytoplankton	${ m g}{ m L}^{-1}$
NFE	Fe in mixed phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
NH4	ammonium + ammonia	$\mathrm{mol}\ \mathrm{L}^{-1}$
NO3	nitrate	$\mathrm{mol}\ \mathrm{L}^{-1}$
OXY	dissolved oxygen	$\mathrm{mol}\ \mathrm{L}^{-1}$
PCH	chlorophyll in pico-phytoplankton	${ m g}{ m L}^{-1}$
PFE	Fe in pico-phytoplankton	$\operatorname{mol} L^{-1}$
PIC	pico-phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
PHA	DMSp producing phytoplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
PO4	generic macronutrient	$\mathrm{mol}\ \mathrm{C}\ \mathrm{L}^{-1}$
POC	small particulate organic carbon	$\mathrm{mol}\ \mathrm{L}^{-1}$
PRO	proto-zooplankton	$\mathrm{mol}\ \mathrm{L}^{-1}$
SFE	Fe in small POM	$\mathrm{mol}\ \mathrm{L}^{-1}$
SIL	dissolved SiO <sub>3</sub>	$\mathrm{mol}\ \mathrm{L}^{-1}$

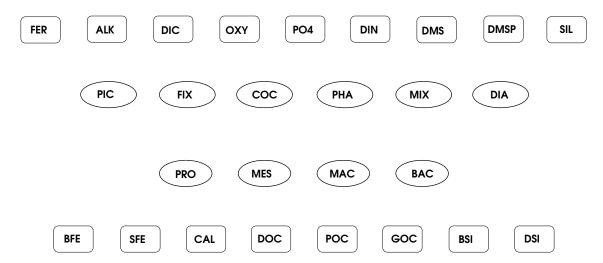


Figure 1: The constituents of PlankTOM10; PFTs are shown as ellipses and tracers as rounded rectangles. There are also tracers for the chlorophyll and iron content of the individual pPFTs but these have been omitted from the figures for clarity.

#### 1.0.2 Tracer Transport

The temporal evolution of all passive tracers T is governed by the balance between its local sources and sinks ('Sources-Minus-Sinks' (SMS), biogeochemical part) and by the physical transport processes (advection and diffusion), hence

$$\frac{dT}{dt} = \nabla \cdot (\vec{u}T) + \nabla \cdot (\vec{K}\nabla T) + SMS,\tag{1}$$

where  $\vec{K}$  is the 3-dimensional tracer diffusion coefficient and  $\vec{u}$  is the fluid velocity, calculated in the physical model. To ensure numerical stability, the sinks processes in SMS are set to zero then the concentration of passive tracers fall below a set threshold (1.e-10).

# 2 Autotrophic PFTs

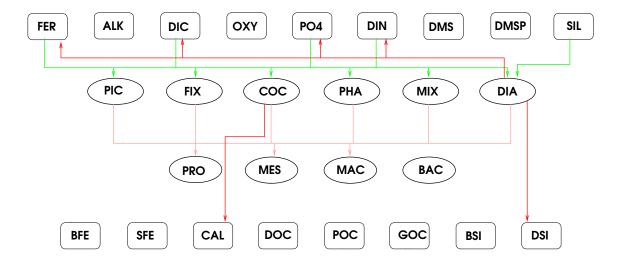
### 2.1 Phytoplankton Biomass - PIC, FIX, COC, PHA, MIX, DIA

The processes governing evolution of phytoplankton biomass for each  $P_i$  is shown in Figure 2. Evolution in terms of carbon is described in this section; chlorophyll (Section 2.2) and iron in phytoplankton (Section 6.1.1) are modelled silimarly. Growth of phytoplankton modifies dissolved organic carbon (Section 4.1), silica (Section 6.2), calcium carbonate (Section 5.1), phosphate, dissolved inorganic nitrogen (Section 6.3), alkalinity (Section 5.3) and oxygen (Section 6.4) in the ocean. The temporal evolution of phytoplankton biomass is given in the equation below:

$$\frac{\partial P_i}{\partial t} = \underbrace{\mu^{P_i} P_i}_{production} - \underbrace{\mu^{P_i}_0 \delta_{P_i} b_{P_i}^T P_i}_{loss} - \underbrace{\sum_j g_{P_i}^{Z_j} Z_j P_i}_{arazina}$$
(2)

 $g_{P_i}^{Z_j}*Z_j*P_i$  describes the amount of biomass lost in grazing by the zPFT  $Z_j, j \in \{1,3\}$  as described in Section 3. In the present configuration of the model all available phytoplankton are grazed so there is no mortality term.

 $\mu_P$  is the phytoplankton growth rate and is a function of temperature, light and nutrient availability:



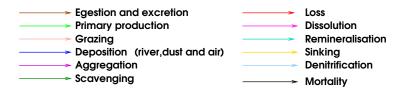


Figure 2: The processes governing the development of the phytoplankton.

$$\mu^{P_i} = \mu_0^{P_i} * (1 + \delta^{P_i}) * f(T) * f(PAR) * f(nut)$$
  
=  $\mu_0^{P_i} * (1 + \delta^{P_i}) * b_{P_i}^T * L_{light}^{P_i} * L_{lim}^{P_i}$  (3)

where  $\mu_0^{P_i}$  is the maximum growth rate at  $0^\circ$  C,  $b_{P_i}$  is the temperature dependence of the growth rate and T is the temperature. For coccolithophorids the growth rate below  $10^\circ$  is reduced to  $(.2 + .8 * \frac{T}{10.}) * b_{coc}^T$ . The radiation available for photosynthesis is dependent on the wavelength and the depth:

$$PAR(z + \Delta z) = .215 * Q_{sr} * e^{-\left(\sum_{i} x_{g} + CHL^{P_{i}} * y_{g}^{P_{i}}\right) \Delta z}$$

$$+ .215 * Q_{sr} * e^{-\left(\sum_{i} x_{r} + CHL^{P_{i}} * y_{r}^{P_{i}}\right) \Delta z}.$$
(4)

where the fraction of available solar radiation  $Q_{sr}$  which is in the photosynthetically active wavelength range has been divided between the blue/green and red wavelengths,  $x_g$ ,  $x_r$  are the extinction coefficients of pure water for blue/green and red wavelengths and  $y_g^{P_i}$ ,  $Y_r^{P_i}$  are the extinction coefficients of chlorophyll. If

$$perfrm = \alpha^{P_i} * \frac{CHL^{P_i}}{P_i} 4.6 * PAR(z)$$
 (5)

and

$$pctnut = \mu_0^{P_i} * (1 + \delta^{P_i}) * b_{P_i}^T * L_{lim}^{P_i}$$
(6)

then

$$L_{light} = 1 - e^{-\frac{perfrm}{pctnut}} \tag{7}$$

The nutrient limitation  $(L_{lim}^{P_i})$  determines the limitation of the growth rate due to the availability of nutrients. It is assumed that nutrient limitation follows Michaelis-Menten kinetics and that growth is determined by the least available nutrient. Hence, for phytoplankton other than silicifiers and nitrogen fixers:

$$L_{lim}^{P_i} = \min\left(\frac{PO_4}{PO_4 + K_{PO_4}^{P_i}}, \frac{\frac{Fe_{P_i}}{P_i} - Fe_{P_i}^{min}}{Fe_{P_i}^{opt} - Fe_{P_i}^{min}}, dinlim\right)$$
(8)

$$dinlim = \frac{NH_4}{NH_4 + K_{NH_4}^{P_i}} + \frac{NO_3(1 - \frac{NH_4}{NH_4 + K_{NH_4}^{P_i}})}{NO_3 + K_{NO_3}^{P_i}}$$
(9)

for silicifiers:

$$L_{lim}^{DIA} = \min\left(\frac{PO_4}{PO_4 + K_{PO_4}^{DIA}}, \frac{\frac{Fe_{DIA}}{DIA} - Fe_{DIA}^{min}}{Fe_{DIA}^{opt} - Fe_{DIA}^{min}}, dinlim, \frac{Si}{Si + K_{Si}^{DIA}}\right). \tag{10}$$

and for nitrogen fixers:

$$L_{lim}^{FIX} = \min \left( \frac{PO_4}{PO_4 + K_{PO_4}^{FIX}}, \frac{\frac{Fe_{FIX}}{FIX} - Fe_{FIX}^{min}}{Fe_{FIX}^{opt} - Fe_{FIX}^{min}}, dinlim + R_{FIX} \left( 1 - dinlim \right) \right)$$
(11)

 $R_{fix}$  is the fraction of the maximum growth rate that can be achieved when growing on  $N_2$ .

# 2.2 Primary Production, Photosynthesis and Chlorophyll - DCH, NCH, CCH, PCH, HCH, FCH

The chlorophyll content of each phytoplankton type (DCH for silicifiers, NCH for mixed-phytoplankton, CCH for calcifiers and PCH for picophytoplankton, HCH for DMS-producers and FCH for  $N_2$ -fixers) is

Table 2: List of Parameters and variables used to compute the evolution of phytoplankton

Term	Variable	Description	Defined in
$\delta_{P_i}$	rn_resphy	respiration as fraction of growth	sms.F90
$\mu_0^{P_i}$	rn_mumpft	maximum growth rate at 0°C	namelist.trc.sms
$\mu^{P_i}$	prophy	productivity of phytoplankton $P_i$	bgcpro.F90
$b_{P_i}$	rn_mutpft	temperature dependence of growth rate	namelist.trc.sms
$\alpha^{P_i}$	rn₋alpphy	initial slope of photosynthesis vs light intensity curve	namelist.trc.sms
PAR	etot	Photosynthetially active radiation	bgcpro.F90
$Q_{sr}$	qsr	surface solar radiation	traqsr.F90
$x_g$	rn₋ekwgrn	absorption coefficient of water for blue-green light	namelist.trc.sms
$x_r$	rn_ekwred	absorption coefficient of water for red light	namelist.trc.sms
$y_g^{P_i}$	rn_kgrphy	light absorption in blue-green	namelist.trc.sms
$y_r^{P_i}$	rn_krdphy	light absorption coefficient for red	namelist.trc.sms
perfrm	perfrm	photosynthetic performance	bgcpro.F90
pctnut	pctnut	macronutrient and temperature defined growth rate	bgcpro.F90
$L_{light}$	xlim8	Light limitation for phytoplankton growth	bgcpro.F90
$Fe_{P_i}^{max}$	rn₋qmaphy	Maximum Fe quota	namelist.trc.sms
$Fe_{P_i}^{min}$ $Fe_{P_i}^{opt}$	rn_qmiphy	Minimum Fe quota	namelist.trc.sms
$Fe_{P_i}^{opt}$	rn_qopphy	Optimum Fe quota	namelist.trc.sms
$K_{NH4}^{P_i}$	rn_kmhphy	half-saturation coefficients for $NH4$	namelist.trc.sms
$K_{NO3}^{P_i}$	rn_kmnphy	half-saturation coefficients for $NO3$	namelist.trc.sms
$K_{PO4}^{P_i}$	rn_kmpphy	half-saturation coefficients for PO4	namelist.trc.sms
$K_{SIL}^{DIA}$	rn_sildia	half-saturation coefficient for SIL in diatoms	namelist.trc.sms
$L_{lim}^{P_i}$	xlimpft	macronutrient limitation for phytoplankton growth	bgcpro.F90

modelled. Chlorophyll evolves in a very similar fashion to phytoplanktonic biomass (see equation 3), as sources and sinks of chlorophyll are of phytoplanktonic origin. The iron-light colimitation model is a dynamical photosynthesis model in which the rate of photosynthesis both controls cellular iron and chlorophyll synthesis and is controlled by their quota (Buitenhuis and Geider, 2010).

$$\frac{\partial Chl^{P_i}}{\partial t} = \underbrace{\rho_{Chl}^{P_i} L_{light} pctnut P_i}_{production} - \underbrace{\mu_0^{P_i} \delta_{P_i} b_{P_i}^T * Chl^{P_i}}_{loss} - \underbrace{\sum_{j} g_{P_i}^{Z_j} Z_j \frac{Chl_{P_i}}{P_i}}_{grazing}, \tag{12}$$

where

$$\rho_{Chl}^{P_i} = \theta_{chl}^{P_i} * pctnut * \frac{L_{light}}{perfrm}$$
 (13)

 $\theta_{chl}^{P_i}$  is the maximum chlorpophyll to carbon ratio for phytoplankton  $P_i$  and perfrm and petnut are defined in equations 5 and 6

Table 3: List of parameters and variables used to calculate the evolution of chlorophyll

Term	Variable	Description	Defined in
$\theta_{Chl}^{P_i}$	rn_thmphy	maximum CHL:C ratio	namelist.trc.sms
$ ho_{Chl}^{P_i}$	rhochl	regulation term of chlorophyll synthesis	bgcpro.F90

## 3 Heterotrophic PFT's

The temporal evolution of zooplankton and the pico-heterotrophs are shown in Figure 3.

#### 3.1 Zooplankton Biomass - PRO, MES and MAC

The temporal evolution of zooplankton concentrations  $Z_j$  in PlankTOM are described as follows (Buitenhuis et al., 2006):

$$\frac{\partial Z_{j}}{\partial t} = \underbrace{\sum_{k} g_{F_{k}}^{Z_{j}} * F_{k} * MGE * Z_{j}}_{growth through grazing} - \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} - \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{mortality through predation} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} - \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} - \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{j}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{k}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{k}}^{Z_{k}} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{k}}^{Z_{k}} * Z_{k} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{k}}^{Z_{k}} * Z_{k} * Z_{k} * Z_{j}}_{loss through grazing} + \underbrace{\sum_{k=j+1}^{3} g_{Z_{k}}^{Z_{k}} * Z_{k} * Z_{k}$$

where  $g_{F_k}^{Z_j}$  is the grazing of zooplankton  $Z_j$  on food source  $F_k$  and MGE is the growth efficiency.  $R_{0^\circ}^{Z_j}$  is the respiration rate at  $0^\circ\mathrm{C}$ ,  $d_{Z_j}$  is the temperature dependence of the respiration ( $d^{10}=Q_{10}$ ).  $m_{0^\circ}^{Z_j}$  is the mortality rate at  $0^\circ\mathrm{C}$ ,  $c_{Z_j}$  is the temperature dependence of the mortality ( $c^{10}=Q_{10}$ ).  $K^{Z_j}$  is the half saturation constant for mortality and is set to  $20*10^{-6}$ . The mortality term for meso- and macrozooplankton is due to predation by top predators for which the total protozooplankton and phytoplankton biomass is used as a proxy. In the presence of ice krill are protected from predation so the macrozooplankton mortality is reduced by a factor of .01.

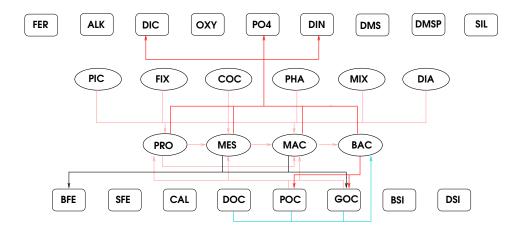




Figure 3: The processes governing the development of the zooplankton and pico-heterortrophs.

Grazing  $g_{F_k}^{Z_j}$ , of zooplankton  $Z_j$  on food source  $F_k$  is dependent on the zooplankton preference,  $p_{F_k}^{Z_j}$ , the concentration of the food source and the temperature.

$$g_{F_k}^{Z_j} = g_{max}^{Z_j}(T) \frac{p_{F_k}^{Z_j}}{K^{Z_j} + \sum_i p_{F_k}^{Z_j} F_k}$$
(15)

The food sources F for zooplankton are summarised in Table 4. For macro-zooplankton they are phytoplankton, meso-zooplankton, proto-zooplankton, pico-heterotrophs, small and large particulate organic matter. The food sources F for meso-zooplankton are phytoplankton, proto-zooplankton are phytoplankton, pico-heterotrophs, small and large particulate organic matter. The food sources for proto-zooplankton are phytoplankton, pico-heterotrophs, small and large particulate organic matter.

Table 4: Food sources for zooplankton and pico-heterotrophs

Food	Macro-zooplankton	Meso-zooplankton	Proto-zooplankton	Pico-heterotrophs
Meso-zooplankton	*			
Proto-zooplankton	*	*		
Phytoplankton	*	*	*	
Pico-heterotrophs	*	*	*	
Large POM	*	*	*	*
Small POM	*	*	*	*
Dissolved OM				*

The temperature dependence of the grazing rate is:

$$g_{max}^{Z_j}(T) = g_{0\circ}^Z b_{Z_j}^T, (16)$$

where  $g_0^Z$  is the maximum grazing rate at  $0^\circ$  C,  $b_{Z_j}$  is the temperature dependence of the grazing rate ( $b^{10} = Q_{10}$ ), T is the local seawater temperature in  ${}^\circ$ Celsius. In shallow water (<600m) in the summer months under ice coverage of between .1 and .3 macrozooplankton experience enhanced recruitment (Wiedenmann et al., 2009). This is included by increasing the growth rate by a factor  $r_{MAC}$  when these conditions apply.

The model growth efficiency MGE, a function of gross growth efficiency (GGE), describes the fraction of grazed food incorporated into zooplankton biomass and basal respiration normalised to all material ingested:

$$MGE_{Z_{j}} = MIN\left(1 - \xi^{Z_{j}}, GGE_{Z_{j}} + \frac{R_{0^{\circ}}^{Z_{j}} * d_{Z_{j}}^{T} * Z_{j}}{\sum_{k} g_{F_{k}}^{Z_{j}}}\right).$$
(17)

Equation 42 shows the possible reduction in  $MGE_{Z_j}$  when zooplankton graze on phytoplankton with a lower  $\frac{Fe}{C}$  ratio than themselves.

#### 3.2 Pico-heterotrophs

The temporal evolution of bacterial concentration is modelled in a similar way to zooplankton:

$$\frac{\partial BAC}{\partial t} = \underbrace{\lambda_{OC}^*(T)BGE * BAC}_{growth \ through \ remineralisation} - \underbrace{R_{0\circ}^{BAC} * d_{BAC}^T * BAC}_{respiration} - \underbrace{\sum_{j} g_{BAC}^{Z_j} * BAC * Z_j}_{grazing}$$
(18)

where BGE is the bacterial growth efficiency. The food sources  $F_k$  for bacteria are DOC and small and large particulate organic carbon. Mineralisation rate  $\lambda_{OC}^*(T)$  is dependent on the temperature and the available food:

$$\lambda_{OC}^{*}(T) = M_{0^{\circ}} * b_{BAC}^{T} \frac{\eta_{O} * \sum_{k} p_{F_{k}}^{BAC} F_{k}}{K_{DOC}^{BAC} + \sum_{k} p_{F_{k}}^{BAC} F_{k}}, \tag{19}$$

where  $M_{0^\circ}$  is the maximum mineralisation rate at  $0^\circ$  C,  $b_{BAC}$  is the temperature dependence of the mineralisation rate ( $b^{10}=Q_{10}$ ) and T is the local seawater temperature in  ${}^\circ$ Celsius. Each food source is associated with a preference  $p_F^{BAC}$ .  $K_{DOC}BAC$  is the half-saturation constant for mineralisation of DOC. Bacterial growth is dependent on the available oxygen:  $\eta_O = \frac{OXY + 3*10^{-6}}{OXY + 10*10^{-6}}$ , which leads to a maximum bacterial growth rate in the absence of oxygen that is 0.3 times the maximum growth rate at high oxygen.

 $R_{0}^{BAC}$  is the respiration rate at 0°C,  $d_{BAC}$  is the temperature dependence of the respiration ( $d^{10} = Q_{10}$ ).

Bacterial growth efficiency BGE, which describes the fraction of mineralised food incorporated into bacterial biomass, is a function temperature and iron availability:

$$BGE = \frac{min(BGE_{0^{\circ}} - e * T, FER_{BAC} + \lambda_{POC}^*Fe + \lambda_{GOC}^*Fe)}{max((\lambda_{DOC}^*DOC + \lambda_{POC}^*POC + \lambda_{GOC}^*GOC) * Fe/C, 1e - 25)}$$
(20)

where  $BGE_{0^\circ}$  is the bacterial growth efficiency at  $0^\circ$  and e is the temperature dependence of bacteria growth,  $FER_{BAC}$  is the uptake of dissolved Fe (see equation 48)and  $\lambda_{GOC}^*$ ,  $\lambda_{DOC}^*$ ,  $\lambda_{POC}^*$  are the remineralisation rates for DOC, GOC and POC respectively as defined above. The remineralisation of iron in POC and GOC is given by:

$$\lambda_{POC}^* Fe = M_{0^{\circ}} * b_{BAC}^T \frac{\eta_O * \sum_k p_{F_k}^{BAC} SFE}{K_{POC}^{BAC} + \sum_k p_E^{BAC} F_k}$$
(21)

and

$$\lambda_{GOC}^* Fe = M_{0^{\circ}} * b_{BAC}^T \frac{\eta_O * \sum_k p_{F_k}^{BAC} BFE}{K_{DOC}^{BAC} + \sum_k p_{F_k}^{BAC} F_k}$$
 (22)

Grazing of bacteria by zooplankton is described in the previous section.

Table 5: List of parameters and variables used to calculate the evolution of zooplankton

Term	Variable	Description	Defined in
$g_0^{Z_j}$	rn₋gramic	maximum grazing rate at $0^{\circ}$	namelist.trc.sms
	rn₋grames	for proto-, meso-	namelist.trc.sms
	rn_gramac	and macrozooplankton	namelist.trc.sms
$g_{max}^{Z_j}$	graze	grazing rate for proto-	bgclos.F90
	graze2	meso- and	bgclos.F90
	graze3	macrozooplankton	bgclos.F90
$b_{Z_i}$	rn_mutpft	Temperature dependence of grazing	namelist.trc.sms
,		for proto, meso- and	namelist.trc.sms
		macro-zooplankton	namelist.trc.sms
$r_{MAC}$	rn_icemac	enhanced recruitment factor under ice	namelist.trc.sms
$p_F^Z$	rn_gmibac	proto-zoo. grazing preference for bacteria	namelist.trc.sms
	rn₋gmigoc	proto-zoo. grazing preference for GOC	namelist.trc.sms
	rn₋gmipoc	proto-zoo. grazing preference for POC	namelist.trc.sms
	rn_gmiphy	proto-zoo. grazing preference for phyto.	namelist.trc.sms
	rn₋gmebac	meso-zoo preference for bacteria	namelist.trc.sms
	rn_gmegoc	meso-zoo. grazing preference for GOC	namelist.trc.sms
	rn₋gmepoc	meso-zoo. grazing preference for POC	namelist.trc.sms
	rn₋gmemic	meso-zoo. grazing preference for proto-zoo.	namelist.trc.sms
	rn_gmephy	meso-zoo. grazing preferencefor phyto	namelist.trc.sms
	rn₋gmabac	macro-zoo preference for bacteria	namelist.trc.sms
	rn_gmagoc	macro-zoo preference for GOC	namelist.trc.sms
	rn_gmames	macro-zoo preference for meso-zoo	namelist.trc.sms
	rn_gmamic	macro-zoo preference for proto-zoo	namelist.trc.sms
	rn_gmapoc	macro-zoo preference for POC	namelist.trc.sms
	rn_gmaphy	macro-zoo preference for each phyto. type	namelist.trc.sms
$K^{Z_j}$	rn_grkmic	half-saturation constant for	namelist.trc.sms
	rn₋grkmes	proto-, meso-	namelist.trc.sms
7	rn_grkmac	and macro-zooplankton	namelist.trc.sms
$\sigma^{Z_j}$	rn₋sigmic	Fraction of zooplankton	namelist.trc.sms
	rn₋sigmes	excretion as DIC	
. 7	rn_sigmac		
$\xi^{Z_j}$	rn₋unamic	Fraction of unassimilated	namelist.trc.sms
	rn₋unames	food by proto-, meso-	
1.00	rn₋unamac	and macro-zooplankton	
$MGE_{Z_j}$	micrge	model growth of efficiency	bgcbio.F90
	mesoge	of proto-, meso- and	
-Z:	macrge	macro-zooplankton	
$R_{0^{\circ}}^{Z_{j}}$	rn₋resmic	Respiration at 0°C of	namelist.trc.sms
	rn_resmes	proto-, meso-	namelist.trc.sms
	rn_resmac	and macro-zooplankton	namelist.trc.sms
$d_{Z_j}$	rn_retmic	Temperature dependence of resipration of	namelist.trc.sms
	rn_retmes	proto-, meso-	namelist.trc.sms
Z	rn_retmac	and macro-zooplankton	namelist.trc.sms
$m_{0}^{Z}$	rn_mormes	mortality at $0^{\circ}$ C of meso-zoo.	namelist.trc.sms
	rn_mormac	and macro-zooplankton	namelist.trc.sms
$c_{Z_j}$	rn_motmes	temperature dependence of mortality	namelist.trc.sms
CCF	rn_motmac	for meso and macro-zooplankton	namelist.trc.sms
$GGE_{Z_j}$	rn_ggemic	Growth efficiency	namelist.trc.sms namelist.trc.sms
	rn_ggemes	of proto-, meso- and	namelist.trc.sms
	rn_ggemac	macro-zooplankton	namensi.trc.sms

Table 6: List of parameters and variables used to calculate the evolution of pico-heterotrophs

Term	Variable	Description	Defined in
$M_0$ °	rn₋grabac	Maximum growth rate for bacteria	namelist.trc.sms
$K_{DOC}^{BAC}$	rn_kmobac	DOC half saturation constant of bacteria	namelist.trc.sms
$p_F^{BAC}$	rn_gbadoc	bacterial preference for DOC	namelist.trc.sms
	rn_gbapoc	bacterial preference for POC	namelist.trc.sms
	rn₋gbagoc	bacterial preference for GOC	namelist.trc.sms
$BGE_{0}^{\circ}$	rn_ggebac	Bacterial growth efficiency at 0°	namelist.trc.sms
$R_{0}^{BAC}$	rn₋resbac	respiration at 0°C	namelist.trc.sms
$d_{BAC}$	rn₋retbac	Temperature dependence of respiration	namelist.trc.sms
e	rn_ggtbac	Temperature dependence of bacterial growth efficiency	namelist.trc.sms
$FER_{BAC}$	ubafer	Uptake of dissolved Fe by bacteria	bgcsnk.F90
$\eta_O$	$\frac{OXY+3*10^{-6}}{OXY+10*10^{-6}}$	oxygen limitation to bacteria growth	
$\lambda_{POC}^* Fe$	ofer	remineralisation of Fe in POC	bgcsnk.F90
$\lambda_{GOC}^*Fe$	ofer2	remineralisation of Fe in GOC	bgcsnk.F90
$\lambda_{DOC}^{\star}DOC$	olimi	remineralisation of DOC	bgcnul.F90 ,bgcsnk.F90
$\lambda_{POC}^{\star}POC$	orem	remineralisation of POC	bgcnul.F90 ,bgcsnk.F90
$\lambda_{GOC}^{\star}GOC$	orem2	remineralisation of GOC	bgcnul.F90 ,bgcsnk.F90

### 3.2.1 Denitrification

When waters become suboxic, bacteria can also use nitrate in order to gain oxidative power for DOC remineralization. Hence, there is a (bacterial) denitrification term in the model (Eq. 63).

#### Organic matter and bacterial remineralisation 4

The source and sinks for dissolved organic carbon (DOC) and small (POC) and large (GOC) particulate carbon are shown in Figure 4

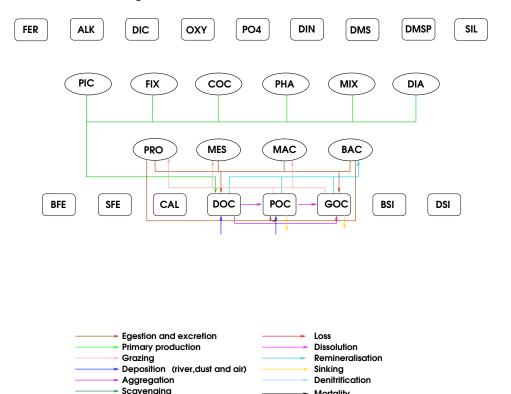


Figure 4: The source and sinks for dissolved organic carbon (DOC) and small (POC) and large (GOC) particulate carbon.

Mortality

#### Dissolved organic carbon - DOC

The evolution of DOC is calculated in the following way:

$$\frac{\partial DOC}{\partial t} = \underbrace{\sum_{production} \nu_{P_i}^{tot} \mu^{P_i} P_i}_{production} + \underbrace{\sum_{j} \left[ (1 - \sigma^{Z_j})(1 - \xi^{Z_j} - MGE^{Z_j}) \sum_{k} g_{F_k}^{Z_j} * Z_j * F_k \right]}_{egestion} + \underbrace{\underbrace{333R_{0^{\circ}}^{BAC} d_{BAC}^{T} BAC}_{excretion} - \underbrace{\lambda_{DOC}^{\star} DOC}_{remineralisation} - \underbrace{\Phi_{agg}^{DOC \to POC} - \Phi_{agg}^{DOC \to GOC}}_{aggregation}}_{aggregation} + \underbrace{DOC_{riv}}_{river\ input}, \tag{23}$$

where  $\nu_{P_i}^{tot} = \nu_{P_i} + (1 - L_{lim}^{P_i})\nu_{P_i}^{max}$  is the fraction of phytoplankton growth which forms DOC. Bacterial degradation of DOC is given by equation 19 for DOC i.e.:

$$\lambda_{DOC}^{\star}DOC = M_{0^{\circ}} * b_{BAC}^{T} \frac{\eta_{0} * p_{DOC}^{BAC} * DOC}{K_{DOC}^{BAC} + p_{DOC}^{BAC}DOC}.$$

$$(24)$$

 $\eta_O = rac{3*10^{-6} + OXY}{OXY + 10^{-6}}$  leads to a maximum bacterial growth rate in the absence of oxygen that is 0.3 times the maximum growth rate at high oxygen. The aggregation functions  $\Phi_{agg}^{X o Y}$  are described in Section 4.2.

Table 7: List of Parameters used in bacterial remineralisation of DOC

Term	Variable	Description	Defined in
$\overline{\nu_{P_i}}$	rn₋docphy	minimum DOC excretion ratio	namelist.trc.sms
$ u_{p_i}^{max}$	rn_domphy	maximum DOC excretion ratio	namelist.trc.sms
$egin{array}{l}  u_{p_i}^{max} \ g_{F_i}^{Z_j} Z_j \end{array}$	gramit	Total grazing by	bgclos.F90
·	gramet	proto,meso and	
	gramat	macro-zooplankton	
$d_{BAC}$	rn₋retbac	temperature dependence of bacterial respiration	namelist.trc.sms
$\lambda_{OC}^{\star}OC$	olimi	Remineralisation rate of DOC	
	orem	Remineralisation rate of POC	
	orem2	Remineralisation rate of GOC	bgcnul.F90 ,bgcsnk.F90
$K_{PO4}^{BAC}$	rn_kmpbac	PO4 half saturation constant	namelist.trc.sms
$K_{FER}^{BAC}$	rn_kmfbac	FER half saturation constant	namelist.trc.sms
$b_{BAC}$	rn_mutpft	temp. dependence of growth rate	namelist.trc.sms
$DOC_{riv}$	depdoc	River input of DOC	trcini.dgom

#### 4.2 Particulate aggregation

Particle aggregation through either differential sinking or turbulent coagulation is calculated by:

$$\Phi_{agg}^{DOC \to POC} = \phi_1^{DOC} \epsilon DOC^2 + \phi_2^{DOC} \epsilon DOC POC 
\Phi_{agg}^{DOC \to GOC} = \phi_3^{DOC} \epsilon DOC GOC 
\Phi_{agg}^{POC \to GOC} = \phi_1^{POC} \epsilon POC^2 + \phi_2^{POC} \epsilon GOC POC 
+ \phi_3^{POC} POC GOC + \phi_4^{POC} POC^2$$
(25)

In which  $\epsilon$  is the shear rate. The coefficients  $\phi$  were obtained by integrating the standard curvilinear kernels for collisions over the size range of each organic matter pool.

Table 8: List of Parameters used in particulate aggregation

Term	Variable	Description	Defined in
$\Phi_{agg}^{DOC \to POC}$	xaggdoc	DOC-POC aggregation	bgcsnk.F90
$\Phi_{agg}^{DOC  o GOC}$	xaggdoc2	DOC-GOC aggregation	bgcsnk.F90
$\Phi_{aaa}^{POC \to GOC}$	xagg	POC-GOC aggregation	bgcsnk.F90
$\phi_1^{DOC}$	rn₋ag5doc	DOC-POC aggregation	namelist.trc.sms
$\phi_2^{DOC}$	1000.		
$\phi_3^{DOC}$	rn_ag6doc	DOC-GOC aggregation	namelist.trc.sms
$\phi_1^{POC}$	rn_ag1poc	POC-GOC aggregation	namelist.trc.sms
$\phi_2^{POC}$	rn_ag2poc	POC-GOC aggregation	namelist.trc.sms
$\phi_3^{POC}$	rn_ag3poc	POC-GOC aggregation	namelist.trc.sms
$\phi_4^{POC}$	rn₋ag4poc	POC-GOC aggregation	namelist.trc.sms

#### 4.3 Sinking

Using the data in Ploug et al. (2008) and applying the drag equations of Buitenhuis et al. (2001) results in a new function describing the relationship between particle density and sinking speed (Buitenhuis et al., 2012):

$$V_{sink} = k_{GOC} * MAX(\rho_{particle} - \rho_{seawater}, \rho_{min})^{S_{GOC}},$$
 (26)

where, if  $\rho_{GOC}$  (=1.08),  $\rho_{CAL}$  (=1.34) and  $\rho_{DSI}$  are the densities of the organic matter, CaCO<sub>3</sub>, and SiO<sub>2</sub> respectively, the particle density  $\rho_{particle}$  is calculated by:

$$\rho_{particle} = \frac{(GOC * 240. + CAL * 100. + DSI * 60.)}{\max(\frac{GOC * 240.}{\rho_{GOC}} + \frac{CAL * 100.}{\rho_{CAL}} + \frac{DSI * 60.}{\rho_{DSI}}, 10^{-15})}$$
(27)

and

$$\rho_{min} = \left(\frac{S_{POC}}{k_{GOC}}\right)^{S_{GOC}} \tag{28}$$

Table 9: List of Parameters used in sinking

Term	Variable	Description	Defined in
$S_{POC}$	rn_snkpoc	sinking speed of POC	namelist.trc.sms
$S_{GOC}$	rn_snkgoc	sinking speed parameter for GOC	namelist.trc.sms
$k_{GOC}$	rn₋singoc	second sinking speed parameter for GOC	namelist.trc.sms
$ ho_{min}$	dnsmin	density at which GOC sinking speed is rn_snkpoc	trclsm.dgom.h90
$ ho_{seawater}$	rhop	density of sea-water	
$ \rho_{particle} - \rho_{seawater} $	xdens	density of particle	bgcsnk.F90
$V_{sink}$	xvsink	sinking speed of particle	bgcsnk.F90

#### 4.4 Small particulate organic carbon - POC

The temporal evolution of small particulate organic carbon, POC, is calculated as

$$\frac{\partial POC}{\partial t} = \underbrace{\xi^{PRO} * \sum_{F_i} g_{F_i}^{PRO} PRO}_{F_i} - \underbrace{\sum_{Z_j} g_{POC}^{Z_j} * Z_j * POC}_{grazing \ on \ POC} + \underbrace{0.333 * R_{0^{\circ}}^{BAC} * d_{BAC}^{T} * BAC}_{excretion} - \underbrace{\lambda_{POC}^{\star} POC}_{POC \ remineralisation} - \underbrace{\sum_{POC} \frac{\partial POC}{\partial z}}_{POC \ sinking} + \underbrace{\Phi_{agg}^{DOC \to POC}}_{aggregation \ to \ POC} - \underbrace{\Phi_{agg}^{POC \to GOC}}_{aggregation \ to \ GOC} + \underbrace{POC_{riv}}_{river \ input}.$$
(29)

Here,  $\xi^{mic}$  is the unassimilated fraction of grazed material,  $g_{F_i}^{mic}$  are the grazing coefficients of proto-zooplankton on food sources F as specified in equation 14, and all others variables are as above.

Table 10: List of parameters and variables used to calculate the evolution of POC

Term	Variable	Description	Defined in
$K_{P_i}$	rn₋mokpft	half saturation constant for	namelist.trc.sms
		mortality	
$POC_{riv}$	deppoc	river input of POC	trcini.dgom.h

## 4.5 Large particulate organic carbon - GOC

The temporal derivative of large particulate organic carbon (GOC) is calculated as

$$\frac{\partial GOC}{\partial t} = \sum_{j} \xi^{Z_{j}} \sum_{k} g_{F_{k}}^{Z_{j}} * Z_{j} * F_{k} - \sum_{j} g_{GOC}^{Z_{j}} * Z_{j} * GOC + \sum_{j} m_{0^{\circ}}^{Z_{j}} * c^{T} * Z_{j}$$

$$zooplankton unassimilated food loss through grazing MES,MAC mortality$$

$$+ \underbrace{\Phi_{agg}^{DOC \to GOC} + \Phi_{agg}^{POC \to GOC}PHA}_{agg} - \underbrace{\lambda_{GOC}^{\star}GOC}_{GOC remineralisation} - \underbrace{V_{sink} \frac{\partial GOC}{\partial z}}_{GOC sinking}. (30)$$

 $\xi^{Z_j}$  is unassimilated fraction of material grazed by meso- and macro-zooplankton and  $m^{Z_j}$  is mesoand macro-zooplankton mortality as in equation (14).  $V_{sink}$  is the sinking rate of GOC and is calculated as equation (26).

# 5 Carbonate chemistry

#### 5.1 Calcite - CAL

Calcification in the model is performed only by phytoplankton calcifiers, COC. Losses of calcifiers result in detached/sinking CaCO<sub>3</sub>, and enters the tracer CAL. Attached CaCO<sub>3</sub> is produced in a fixed ratio to organic matter and therefore there is no tracer for its concentration. It does however reduces alkalinity, ALK, and dissolved inorganic carbon, DIC. The source and sinks for detached carbonate (CAL), dissolved inorganic carbon (DIC) and alkalinity (ALK) are shown in Figure 5

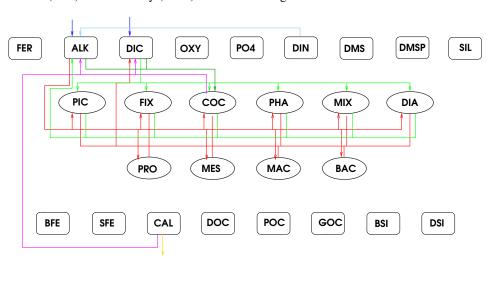




Figure 5: The source and sinks for detached carbonate (CAL), dissolved inorganic carbon (DIC) and alkalinity (ALK).

$$\frac{\partial CaCO_{3attached}}{\partial t} = R_{CAL} \underbrace{\mu^{COC}COC}_{production \ by \ COC}$$
(31)

For detached CaCO<sub>3</sub>, CAL:

$$\frac{\partial CAL}{\partial t} = R_{CAL}(1 - R_{diss}) \left( \underbrace{\mu_0^{COC} \delta_{COC} b_{COC}^T COC}_{COC \ loss} + \underbrace{\sum_{j} g_{COC}^{Z_j} Z_j * COC}_{grazing \ by \ zooplankton} \right) - \underbrace{V_{sink} \frac{\partial CAL}{\partial z}}_{sinking} - \underbrace{\beta_{CO_3} CAL}_{dissolution},$$
(32)

where  $R_{CAL}$  is the calcification to particulate primary production ratio,  $R_{diss}$  is the fraction of attached coccoliths that is dissolved during losses of coccolithophores,  $V_{sink}$  is the sinking speed and is described in section 4.3, and  $\beta_{CO_3}$  is the dissolution rate:

$$\beta_{CO_3} = MIN\left(1, \frac{1 - \delta_{sat}}{K_{CAL} + \delta_{sat}}\right) \tag{33}$$

where  $\delta_{sat}$  is the deviation from saturation and  $K_{CAL}$  is the half saturation constant for calcite dissolution.  $\beta_{CO_3}$  is  $0.25\ month^{-1}$  at the sea surface, and  $1\ month^{-1}$  at and below saturation.

CAL is calculated in bgcbio and reduced by the fraction dissolved in bgclys.

Table 11: List of parameters and variables used to calculate the evolution of calcite

Term	Variable	Description	Defined in
$R_{CAL}$	rn₋coccal	CaCO <sub>3</sub> to Carbon ratio	namelist.trc.sms
$\mu^{COC}$	prophy	coccolithophorid productivity	bgcpro.F90
$R_{diss}$	rn₋discal	Fraction of CaCO <sub>3</sub> dissolved	namelist.trc.sms
		during coccolithophorid death	
$K_{CAL}$	rn_lyscal	half saturation constant for calcite dissolution	namelist.trc.sms
$\delta_{sat}$	delco3	deviation from saturation	bgclys.F90
$\beta_{CO_3}CAL$	remco3	dissolved CaCO <sub>3</sub>	bgclys.F90
$V_{sink}CAL$	sinkcal	sinking speed of CaCO <sub>3</sub>	bgcsnk.F90

#### 5.2 Dissolved inorganic carbon - DIC

The temporal evolution of dissolved inorganic carbon, DIC, is calculated as

$$\frac{\partial DIC}{\partial t} = \underbrace{-\sum_{i} \mu^{P_{i}} * (1 + \nu^{TOT}_{P_{i}}) P_{i}}_{primary \ production} + \underbrace{R_{diss} R_{CAL} \left( \underbrace{\mu^{P_{i}}_{0} \delta_{COC} b^{T}_{COC} COC}_{COC} + \sum_{j} g^{Z_{j}}_{COC} Z_{j} COC \right)}_{grazing \ by \ zooplankton} + \underbrace{DIC_{riv}}_{river \ input} + \underbrace{\beta_{CO_{3}} CAL}_{dissolution} + F^{CO_{2}}_{air-sea} . \tag{34}$$

In addition to the inclusion of grazing by zooplankton remineralistion by bacteria is included as a function of their growth efficiency and respiration (in this case subscript j includes the pico-heterotrophs):

$$\begin{array}{ll}
\operatorname{consum} &= \underbrace{\sum_{j} \sigma^{Z_{j}} * (1 - \xi^{Z_{j}} - MGE^{Z_{j}}) \sum_{k} g_{F_{k}}^{Z_{j}} * Z_{j} * F_{k}}_{foodrespiration} \\
&+ \underbrace{(1 - BGE) * (\lambda_{DOC}^{\star} DOC + \lambda_{POC}^{\star} POC + \lambda_{GOC}^{\star} GOC)}_{remineralisation} \\
&+ \underbrace{\sum_{j=1}^{3} R_{0\circ}^{Z_{j}} d_{Z_{j}}^{T} Z_{j} + \underbrace{.333R_{0\circ}^{BAC} d_{BAC}^{T} BAC}_{respiration} + \underbrace{\sum_{i} \delta_{P_{i}} b_{P_{i}}^{T} \mu_{0}^{P_{i}} P_{i}}_{loss}.
\end{array} \tag{35}$$

The bacterial growth efficiency, BGE, is given by Equation 20. The terms for attached CaCO<sub>3</sub> and production of DIC by dissolution are described in Section 5.1. River deposition  $DIC_{riv}$  is the input of DIC from rivers, see Section 8.6. The air-to-sea flux is described in section 7.

Dissolved inorganic carbon is calculated in bgcbio; in bgclys the CaCO<sub>3</sub> dissolution to DIC is included while in bgcflx the air-sea flux of DIC is added.

Table 12: List of Parameters used in the evolution of DIC and ALK

Term	Variable	Description	Defined in
BGE	bactge	bacteria growth efficiency	bgcbio,bgcsnk.F90
$DIC_{riv} depdic$		river input of DIC	river.nc
$R_{\frac{N}{C}}$	alknut	N+S+P to Carbon ratio	trcini.dgom.F90
$DIC_{riv}$	depdic	River deposition of DIC	trcini.dgom.F90

#### 5.3 Alkalinity - ALK

The temporal evolution of alkalinity is calculated as:

$$\frac{\partial ALK}{\partial t} = R_{C}^{N} \left( \sum_{i} \mu^{P_{i}} P_{i} (1 + \nu_{P_{i}}^{tot}) - \underbrace{consum}_{remineralisation} \right) - \underbrace{2 * R_{CAL} \mu^{coc}COC}_{calcification} \\
+ 2R_{CAL} R_{diss} \left( \mu_{0}^{COC} \delta_{coc} b_{COC}^{T}COC + \sum_{j} g_{coc}^{Z_{j}} Z_{j}COC \right) \\
+ \underbrace{DIC_{riv}}_{river\ input} + \underbrace{N_{denit}}_{denitrification} + \underbrace{2 * \beta_{CO_{3}} CaCO_{3}}_{dissolution} \tag{36}$$

where  $R_{\frac{N}{C}} = \frac{N+S+P}{C} = \frac{16+6+1}{122}$  is the effect of nutrient uptake and remineralisation on alkalinity (Wolf-Gladrow et al., 2007). The terms for the production of attached CaCO<sub>3</sub>, dissolved COC and dissolved CaCO<sub>3</sub> are described in Section 5.1. River deposition,  $DIC_{riv}$  is described in Section 8.6 and denitrification,  $N_{denit}$  in Section 6.3.

# 6 Nutrients and gases

The processes governing the evolution of dissolved iron (FER), large (BFE) and small (SFE) particulate iron, dissolved silica (SIL), biogenic silica (BSI) and detrital silica (DSI) are shown in Figure 6.

The processes governing the evolution of phosphate (PO4), dissolved inorganic nitrogen (NO3 and NH4) and gases (OXY and optionally N2S, N2O and DMS) are shown in Figure 7.

#### 6.1 The Iron Cycle

### 6.1.1 Fe in PFTs

The iron content of phytoplankton (DFE for silicifiers, NFE for mixed-phytoplankton, CFE for calcifiers, PFE for picophytoplankton, HFE for DMS producers and FFE for  $N_2$ -fixers) is given by:

$$\frac{\partial Fe^{P_i}}{\partial t} = \underbrace{\mu_0^{P_i} (1 + \delta^{P_i}) L_{Q_{Fe}}^{P_i} L_{limFe}^{P_i} b_{P_i}^T P_i}_{production} - \underbrace{\sum_{j} g_{P_i}^{Z_j} Z_j * Fe^{P_i}}_{arazing} \tag{37}$$

 $\rho_{Fe}^{P_i}$  describes the iron-light colimitation to phytoplankton growth (Buitenhuis and Geider, 2010) and is given by:

$$L_{Q_{Fe}}^{P_{i}} = \left(\frac{(\frac{\rho max}{\rho min}Fe_{P_{i}}^{max} - Fe_{P_{i}}^{max})(Fe_{P_{i}}^{max} - \frac{Fe_{P_{i}}}{P_{i}})}{(Fe_{P_{i}}^{max} - Fe_{P_{i}}^{min})} + Fe_{P_{i}}^{max}\right) * L_{light}$$
(38)

For phytoplankton other than nitrogen fixers and silicifiers the nutrient limitation is given by:

$$L_{limFe}^{P_{i}} = \min\left(\frac{PO_{4}}{PO_{4} + K_{PO_{4}}^{P_{i}}}, \frac{FER}{FER + K_{FER}^{P_{I}}}, dinlim\right)$$
(39)

in which dinlim is defined in Eq. 9, for silicifiers

$$L_{limFe}^{DIA} = \min\left(\frac{PO_4}{PO_4 + K_{PO_4}^{DIA}}, \frac{FER}{FER + K_{FER}^{DIA}}, dinlim, \frac{Si}{Si + K_{Si}^{DIA}}\right). \tag{40}$$

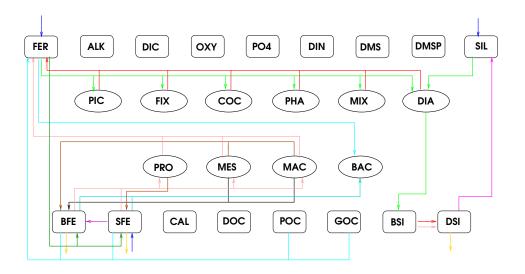
and for nitrogen fixers:

$$L_{limFe}^{FIX} = \min\left(\frac{PO_4}{PO_4 + K_{PO_4}^{FIX}}, \frac{FER}{FER + K_{FER}^{FIX}}, dinlim + R_{FIX} \left(1 - dinlim\right)\right)$$
(41)

The Fe/C ratio of zooplankton is fixed. If zooplankton graze on phytoplankton that have a higher Fe:C ratio than themselves, the excess is remineralised to dissolved iron. If the phytoplankton Fe/C ratio is lower than zooplankton Fe:C, the model growth efficiency (MGE) is decreased:

$$MGE^{Z_{j}} = MIN \left( 1 - \xi^{Z_{j}}, GGE_{Z_{j}} + \frac{R_{0}^{Z_{j}}^{Z_{j}} d_{Z_{j}}^{T} Z_{j}}{\sum_{k} g_{F_{k}}^{Z_{j}}}, \frac{\sum_{k} g_{F_{k}}^{Z_{j}} \frac{Fe_{F_{k}}}{F_{k}} (1 - \xi^{Z_{j}})}{MAX \left( \sum_{k} g_{F_{k}}^{Z_{j}} \left( \frac{Fe}{C} \right)_{Z}, 1e - 25 \right)} \right)$$
(42)

#### 6.1.2 Fe in detrital matter - BFE, SFE



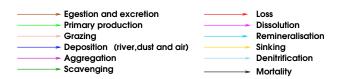


Figure 6: The sources and sinks for dissolved iron (FER), large (BFE) and small (SFE) particulate iron, dissolved silica (SIL), biogenic silica (BSI) and detrital silica (DSI).

Iron in detrital matter is divided into BFE in large organic particles (GOC) and SFE in small organic particles (POC). Production terms of particulate organic iron follow the Fe/C ratio of the source organisms.

There is no iron in DOM, but iron is added from dissolved iron to particulate organic iron during degradation of DOM. Degradation of POM conserves the Fe:C ratio of POM. The bottom correction removes as much carbon from the bottom water layers as is added by rivers (Section 8.6). Because iron is scavenged, the Fe/C ratio of POM sometimes becomes excessive. It is therefore set to a maximum, currently 2<sup>-6</sup> mol:mol.

$$\frac{\partial BFE}{\partial t} = \underbrace{Fe_{scave}(POC + GOC + DSI + CAL)GOC}_{scavenging} - \underbrace{\sum_{j} g_{GOC}^{Z_{j}} * Z_{j} * GOC}_{grazing \ loss}^{BFE} + \underbrace{\left(\frac{Fe}{C}\right)_{Z} \sum_{j=MES,MAC} m_{0^{\circ}}^{Z_{j}} c^{T} z^{j}}_{mortality} + \underbrace{\sum_{j=MES,MAC} \xi^{Z_{j}} \sum_{k} g_{F_{k}}^{Z_{j}} * Z_{j} * F_{k} \frac{Fe_{F_{k}}}{F_{k}}}_{unassimilated \ food} + \underbrace{\phi_{agg}^{POC \to GOC} \frac{SFE}{POC}}_{Fe \ aggregation} - \underbrace{\lambda_{GOC}^{*}Fe}_{remineralisation} - \underbrace{V_{sink}}_{sinking \ of \ BFE}^{OC} \underbrace{\partial BFE}_{sinking \ of \ BFE}$$

$$(43)$$

$$\frac{\partial SFE}{\partial t} = \underbrace{Fe_{scave} * (POC + GOC + DSI + CAL) * POC}_{scavenging} \\
- \underbrace{\sum_{j} g_{POC}^{Z_{j}} * Z_{j} * POC}_{grazing \ loss}^{SFE} + \xi^{MIC} \sum_{k} g_{F_{k}}^{MIC} * MIC * F_{k} \frac{Fe_{F_{k}}}{F_{k}}}_{unassimilated \ food} \\
- \underbrace{\phi_{agg}^{POC \to GOC} \frac{SFE}{POC}}_{Fe \ aggregation} - \underbrace{\lambda_{POC}^{*}Fe}_{remineralisation} - \underbrace{\lambda_{POC}^{*}Fe}_{sinking \ of \ SFE} + \underbrace{\left(\frac{Fe}{C}\right)_{Z} POC_{riv}}_{river \ invut} \tag{44}$$

The remineralisation  $\lambda_{POC}^* Fe$  is given by equation 19.  $Fe_{scav}$  is described below.

#### 6.1.3 Dissolved Fe - FER

The temporal evolution of dissolved iron, FER, is calculated as follows:

$$\frac{\partial FER}{\partial t} = -\underbrace{\mu_0^{P_i}(1+\delta^{P_i})\rho_{Fe}^{P_i}L_{limFe}^{P_i}b_{P_i}^TP_i}_{production} + \underbrace{\sum_{j}\left(\sum_{k}g_{f_k}^{z_j}*Z_j*F_k\frac{Fe_{F_k}}{F_k}(1-\xi^{Z_j}) - \left(\frac{Fe}{C}\right)_{Z}\sum_{k}g_{F_k}^{Z_j}*Z_j*F_k*MGE^{Z_j}\right)}_{grazing} + \underbrace{FER_{remin\_POC\_GOC} + FER_{remin\_BFE\_SFE} - \underbrace{FER_{BAC}}_{bacterial\ demand\ for\ FER} - \underbrace{Fe_{scav}}_{scavenging\ dust\ deposition\ river\ input} + \underbrace{Fe_{riv}}_{scavenging\ dust\ deposition\ river\ input} \tag{45}$$

Iron is input from rivers, see Section 8.6, and the dissolution of dust from the atmosphere, see Section 8.5. Iron is taken up by phytoplankton during primary production (see above). When iron concentration is above 0.6 nM, it is scavenged by POM: the evolution of scavenged iron, Fe<sub>scav</sub> is calculated as:

$$Fe_{scav} = k_{scm} + k_{sc} * (POC + GOC + CAL + DSI) * 1e6$$

$$* \frac{-(1 + l_{Fe}k_{eq} - FERk_{eq}) + ((1 + l_{Fe}k_{eq} - FERk_{eq})^2 + 4FERk_{eq})^{0.5}}{2k_{eq}}$$
(46)

where  $k_{scm}$  and  $k_{sc}$  are scavenging parameters and  $k_{eq}$  is given by:

$$k_{eq} = 10^{17.27 - \frac{1565.7}{T - 19}}. (47)$$

The iron ligand,  $l_{Fe}$  is set to a value of  $.6*10^-9$  at latitudes North of 30S and below 200m depth,  $.3*10^-9$  South of 40S and below 200 m, 0 above 100m depth, and linearly interpolated in between. Part of the scavenged iron is added to POM, and part is removed from the model.

Bacteria demand for Fe can be supplied from the remineralisation of BFE and SFE and from dissolved iron. The net effect on FER may be an increase - if remineralisation exceeds the bacterial demand or a decrease if demand exceeds that supplied by remineralisation. Bacterial demand for FER, FER $_{BAC}$  is:

$$FER_{BAC} = \frac{BGE\left(\frac{Fe}{C}\right)_Z * \left(\lambda_{DOC}^*DOC + \lambda_{POC}^*POC + \lambda_{GOC}^*GOC - \lambda_{POC}^*Fe - \lambda_{GOC}^*Fe\right)FER}{K_{FER}^{BAC} + FER} \tag{48}$$

or zero if this is negative.

The contribution to FER from remineralisation of POC and GOC is:

$$FER_{remin\_POC\_GOC} = \lambda_{POC}^* Fe + \lambda_{GOC}^* Fe$$
(49)

The remineralisation of BFE and SFE contributes to FER by:

$$FER_{remin\_BFE\_SFE} = -BGE \left(\frac{Fe}{C}\right)_{Z} * (\lambda_{DOC}^{*}DOC + \lambda_{POC}^{*}POC + \lambda_{GOC}^{*}GOC - \lambda_{POC}^{*}Fe - \lambda_{GOC}^{*}Fe)$$

$$(50)$$

or zero if this is negative.

Table 13: List of parameters and variables used to calculate the evolution of iron

Term	Variable	Description	Defined in
$Fe_{th}$	rn₋rhfphy	maximum/minimum Fe uptake rate	namelist.trc.sms
$FER_{remin\_BFE\_SFE}$	rbafer	Release of dissolved Fe by bacteria	bgcsnk.F90
$Fe_{scav}$	xscave	Iron scavenged by particulate organic matter	bgcsnk.F90
$Fe_{riv}$	depfer	River deposition	trcini.dgom.F90
$Fe_{dep}$	irondep	Dust deposition	bgcbio.F90
$k_{sc}$	rn_scofer	Scavenging rate for iron by particles	namelist.trc.sms
$k_{scm}$	rn_scmfer	Minimum scavenging rate for iron	namelist.trc.sms
$k_{eq}$	xkeq	Scavenging rate parameter	bgcsnk.F90
$l_{Fe}$	ligfer	iron ligand concentration	bgcsnk.F90

#### 6.2 The Silicate cycle

Silica is input from rivers and the dissolution of dust from the atmosphere. Growth of diatoms consumes dissolved silica (SIL) from the water to produce hydrated silica (biogenic silica BSI). Loss processes of diatoms produce sinking particulate silica (DSI).

#### 6.2.1 Dissolved SiO<sub>3</sub> - SIL

The temporal evolution of dissolved silica is calculated as:

$$\frac{\partial SIL}{\partial t} = \underbrace{-0.15 \min\left(1, \frac{SIL}{K_{SIL}}\right) \left(\frac{Si}{C}\right)_{DIA} \mu^{DIA}DIA}_{production} + \underbrace{SIL_{riv}}_{river\ input} + \underbrace{SIL_{dep}}_{dust deposition}$$
(51)

where  $\mu^{DIA}DIA$  is the primary production, in terms of carbon, of diatoms,  $K_{SIL}$  is the half saturation constant for SiO<sub>3</sub> in diatoms,  $\beta_{Si}$  is the remineralisation rate of silica which is dependent on temperature,

T and oxygen OXY:

$$\beta_{Si} = \min\left(1.32 * 10^{16} e^{\frac{-11200}{(273.15+T)}}, .1\right) \eta_O.$$
 (52)

 $\left(\frac{Si}{C}\right)_{DIA}$  increases with iron stress and silicate availability:

$$\left(\frac{Si}{C}\right)_{DIA} = 4. - 3 * min\left(\frac{max(0, FER)}{K_{FER}^{DIA}}, 1\right).$$
(53)

Observations in the Southern Ocean show a high  $\left(\frac{Si}{C}\right)_{DIA}$  ratio in areas with very high Si concentration so  $\left(\frac{Si}{C}\right)_{DIA}$  is arbitrarily increased throughout the ocean to reflect this:

$$\left(\frac{Si}{C}\right)_{DIA} = \frac{6.*SIL}{SIL + K_{BSI}}.$$
(54)

 $\left(rac{Si}{C}
ight)_{DIA}$  is set to the higher of these two ratios.  $SIL_{dep}$  is described in 8.5 and  $SIL_{riv}$  in 8.6.

Table 14: List of parameters and variables used to calculate the evolution of silica

Term	Variable	Description	Defined in
$\beta_{Si}$	siremin	remineraliation rate of silica	bgcsnk.F90
$\mu_{DIA}$	prophy	primary production of diatoms	bgcpro.F90,bgcnul.F90
$\overset{Si}{C}_{PIA}^{IA}_{KFER}$	silfac	Si/C ratio of diatoms	bgcpro.F90
$K_{FER}^{DIA}$	rn_kmfphy	half saturation constant of Fe	namelist.trc.sms
$K_{BSI}$	rn_kmsbsi	half saturation constant for $\left(\frac{Si}{C}\right)$	namelist.trc.sms
$SIL_{riv}$	depsil	river input of SiO <sub>3</sub>	trcini.dgom.F90
$SIL_{atm}$	sidep	input of atmospheric silica to the water column	bgcbio.F90

#### 6.2.2 Biogenic particulate silica - BSI

The temporal evolution of biogenic silica is calculated as:

$$\frac{\partial BSI}{\partial t} = \underbrace{0.15 \min\left(1, \frac{SIL}{K_{SIL}}\right) \left(\frac{Si}{C}\right)_{DIA} \mu^{DIA}DIA}_{production} - \underbrace{\sum_{j} g_{DIA}^{Z_{j}} * Z_{j} * DIA \frac{BSI}{DIA}}_{grazing} - \underbrace{\delta_{DIA} \mu_{0}^{DIA} b^{T} \frac{BSI}{DIA}}_{loss} \tag{55}$$

where  $\delta^{DIA}$  is the excretion ratio for diatoms and  $\left(\frac{Si}{C}\right)_{DIA}$  is described above.

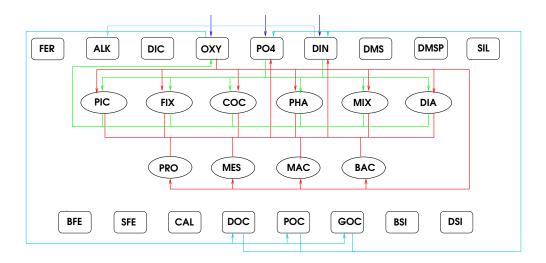
#### 6.2.3 Sinking particulate silica - DSI

The temporal evolution of sinking particulate silica is calculated as:

$$\frac{\partial DSI}{\partial t} = \underbrace{\delta_{DIA}\mu_0^{DIA}b^T \frac{BSI}{DIA}}_{loss} - \underbrace{\beta_{Si}DSI}_{dissolution} + \underbrace{\sum_{j} g_{DIA}^{Z_j} * Z_j * DIA \frac{BSI}{DIA}}_{grazing} + \underbrace{V_{sink} \frac{\partial DSI}{\partial z}}_{sinking DSI}$$
(56)

where  $\delta^{DIA}$  is the excretion ratio for diatoms as above.

#### 6.3 Phosphorus and Nitrogen - PO4, NH4 and NO3



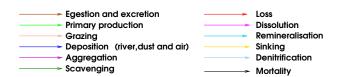


Figure 7: The sources and sinks for phophate (PO4), nitrogen (DIN=NH4+NO3), oxygen (OXY)).

Phosphate is input to the ocean by river deposition; it is consumed during phytoplankton growth and produced during respiration.

$$\frac{\partial PO4}{\partial t} = \underbrace{\sum -\mu^{P_i} P_i \left(1 + \nu^{tot}_{P_i}\right) \frac{P}{C}}_{production} + \underbrace{consum \frac{P}{C}}_{remineralisation} + \underbrace{PO4_{riv}}_{river\ input}$$
(57)

consum is defined in equation 35.

Dissolved ammonium evolves as:

$$\frac{\partial NH4}{\partial t} = \underbrace{\sum -\mu^{P_i} P_i (1 + \nu^{tot}_{p_i}) \frac{N}{C} DIN_{NH4}}_{production} + \underbrace{\underbrace{consum \frac{N}{C}}_{remineralisation}}_{production} - nitrification + \underbrace{\underbrace{NHy_{riv} \frac{N}{C}}_{atmosphere deposition}}_{river input} + \underbrace{\underbrace{NHy_{atm}}_{atmosphere deposition}} \tag{58}$$

For phytoplankton other than nitrogen fixers:

$$DIN_{NH4} = \frac{NH_4}{(NH_4 + K_{NH4}^{P_i})dinlim}$$
 (59)

and for nitrogen fixers:

$$DIN_{NH4} = \frac{NH_4}{(NH_4 + K_{NH4}^{P_i})(dinlim + R_{fix}(1 - dinlim))}$$
(60)

dinlim is defined in Eq. 9.

$$nitrification = r_{nitrif} * max((1 - log(OXY * 1e6) * 0.159)(1 - resp_{BAC}^{NO_3}), 0)$$

$$* \frac{NH4}{NH4 + K_{nitrif}} * d_{BAC}^T * NH4$$
(61)

Dissolved nitrate evolves as:

$$\frac{\partial NO3}{\partial t} = \underbrace{\sum -\mu^{P_i} P_i (1 + \nu^{tot}_{p_i}) \frac{N}{C} DIN_{NO3}}_{production} - \underbrace{\underbrace{N_{denit}}_{denitrification}}_{production} + \underbrace{NOx_{riv} \frac{N}{C}}_{river\ input} + \underbrace{NOx_{atm}}_{atmosphere\ deposition}$$
(62)

where

$$N_{denit} = 0.8 \left( \frac{O}{C} * consum * resp_{BAC}^{NO_3} \right).$$
 (63)

 $\frac{O}{C}=\frac{172}{122}$  and  $resp_{BAC}^{NO_3}$  is the fraction of bacterial respiration that uses NO<sub>3</sub> rather than O<sub>2</sub> and is described in Section 6.4. For phytoplankton other than nitrogen fixers:

$$DIN_{NO3} = \frac{NO_3(1 - \frac{NH_4}{NH_4 + K_{NH_4}^{P_i}})}{(NO_3 + K_{NO3}^{P_i})dinlim}$$
(64)

and for nitrogen fixers:

$$DIN_{NO3} = \frac{NO_3(1 - \frac{NH_4}{NH_4 + K_{NH4}^{P_i}})}{(NO_3 + K_{NO3}^{P_i})(dinlim + R_{fix}(1 - dinlim))}$$
(65)

Table 15: List of Parameters used in the evolution of phosphate and nitrogen

Term	Variable	Description	Defined in
$DIN_{NH4}$	1-dinpft	fraction of phyto growth that is supported by NH4	bgcpro.F90
$DIN_{NO3}$	dinpft	fraction of phyto growth that is supported by NO3	bgcpro.F90
$K_{NH4}^{P_i}$	rn_kmhphy	NH <sub>4</sub> half saturation constants for phytoplankton	namelist.trc.sms
$K_{nitrif}$	rn_kmhnit	NH <sub>4</sub> half saturation constant nitrification	namelist.trc.sms
$K_{NO3}^{P_i} \ rac{N}{C} \ N_{denit}$	rn_kmnphy	NO <sub>3</sub> half saturation constants for phytoplankton	namelist.trc.sms
$\frac{N}{C}$	ratn2c	N:C ratio organic matter = 16:122	trcini.dgom.F90
$N_{denit}$	denitr	denitrification	bgcbio.F90
$NHy_{atm}$	atmamm	Atmosphere input of $NH_y$	trcini.dgom
$NHy_{riv}$	depamm	River input of $NH_y$	trcini.dgom
$NOx_{atm}$	atmnit	Atmosphere input of $NO_x$	trcini.dgom
$NOx_{riv}$	depnit	River input of $NO_x$	trcini.dgom
$PO4_{riv}$	deppo4	River input of phosphate	trcini.dgom
$R_{FIX}$	rn_munfix	Fraction of growth rate during N <sub>2</sub> fixation	namelist.trc.sms
		relative to growth on fixed N	
$r_{nitrif}$	rn_nitnh4	NH4 saturated nitrification rate at 0 C	namelist.trc.sms
$resp_{BAC}^{NO_3}$	nitrfac	fraction of bacterial respiration	bgcnul.F90
2.10		using NO <sub>3</sub> rather than O <sub>2</sub>	

#### 6.4 Oxygen - OXY

Oxygen is produced during the growth of phytoplankton. It is consumed during the growth of  $N_2$  fixers on  $N_2$  and during the remineralisation described by the term consum in Section 5.2. There is also an exchange of oxygen with the atmosphere.

$$\frac{\partial OXY}{\partial t} = \underbrace{\frac{O}{C} \sum \mu^{P_i} P_i \left(1 + \nu^{tot}_{P_i}\right)}_{phytoplankton \ growth} - \underbrace{\frac{N}{C} \mu^{P_{fix}} P_{fix} \left(1 + \nu^{tot}_{FIX}\right) 1.25 (1 - DIN_{nit})}_{growth \ of \ N_2 \ fixers \ onN_2} - \underbrace{\frac{O}{C} consum (1 - resp_{BAC}^{NO_3})}_{remineralisation} + \underbrace{\frac{F_{air-sea}^{O_2}}{O_2 \ flux \ from \ air \ to \ sea}}_{O_2 \ flux \ from \ air \ to \ sea}$$
(66)

Eq. 66 is not used because OXY is fixed at observed concentrations. The fraction of bacterial respiration that uses  $NO_3$  rather than  $O_2$ ,  $resp_{BAC}^{NO_3}$  is given by:

$$resp_{BAC}^{NO_3} = \frac{\sin\left(\max\left(-.5, \frac{8.5E - 6 - OXY}{17E - 6 + OXY}\right) * \pi\right) + 1}{2}$$
 (67)

The air-sea exchange of oxygen,  $F_{air-sea}^{O_2}$ , is given by

$$F_{air-sea}^{O_2} = \left(\frac{O}{N_{pi}} sol_{O_2} \left(1. - e^{20.1050 - 0.0097982 * sstk - 6163.10/sstk}\right) - OXY\right) 0.27v^2 (1 - \gamma) (68)$$

Eq. 68 is not used because OXY is fixed at observed concentrations. The terms are described described in Section 7. It is calculated in bgcflx.

#### 6.5 Diagnostic nitrous oxide - N2S

The diagnostic formulation of nitrous oxide production is a function of  $O_2$  consumption, with a yield that depends on the oxygen concentration. Under oxic conditions, there is a constant yield, while under suboxic conditions the yield increases as oxygen decreases:

$$\frac{\partial N2S}{\partial t} = (\alpha + \beta * exp(-0.1 * \frac{OXY - 1e - 6}{1e - 6})) * \frac{O}{C}consum(1 - resp_{BAC}^{NO_3})$$
 (69)

#### 6.6 Prognostic nitrous oxide - N2O

The prognostic formulation of nitrous oxide production is a function of redox reactions in the nitrogen cycle.

$$\frac{\partial N2O}{\partial t} = (y_{nitrif} * nitrification + y_{denitr} * N_{denit} - y_{N2Ocons} * N_{N2Ocons})$$
 (70)

$$N_{N2Ocons} = 0.8 \left( \frac{O}{C} * consum * \frac{\sin\left(\max\left(-.5, \frac{7E - 6 - OXY}{14E - 6 + OXY}\right) * \pi\right) + 1}{2} \right)$$
 (71)

# 7 Air-sea exchange of gases

The air-sea flux of gases ( $CO_2$ ,  $O_2$ , and optionally  $N_2O$  and/or DMS) is given by the product of gas exchange coefficient and the difference in concentration of the gas across the sea-air interface:

$$F_{air-sea} = k_w * (1 - \gamma) * (pC_{gas}^{air} - pC_{gas}^{sea})$$

$$\tag{72}$$

where  $k_w$  is the gas exchange coefficient,  $\gamma$  is the fraction of the ocean covered by ice,  $pC_{gas}^{air}$  is the concentration of the gas in the air directly above the water, and  $pC_{gas}^{sea}$  is the sea surface concentration of the gas.

Table 16: List of Parameters used in the evolution of N2S and N2O

Term	Variable	Description	Defined in
$\alpha$	rn₋aoun2s	yield of oxic N2O production	namelist.trc.sms
$\beta$	rn_betn2s	yield of suboxic N2O production	namelist.trc.sms
consum	consum	remineralisation rate	Eq. 35, bgcnul.F90
$N_{denit}$	denitr	denitrification	Eq. 63, bgcbio.F90
$N_{N2Ocons}$	degn2o	N2O consumption rate/ $y_{N2Ocons}$	Eq. 71, bgcbio.F90
nitrification	nitrif	nitrification rate	Eq. 61, bgcbio.F90
$\frac{O}{C}$	rato2c	-O2:C ratio = 172:122	trcini.dgom.F90
$resp_{BAC}^{NO_3}$	nitrfac	fraction of bacterial respiration	Eq. 67, bgcnul.F90
- Bile		using NO <sub>3</sub> rather than O <sub>2</sub>	
$y_{N2Ocons}$	rn₋degn2o	yield of N2O consumption	namelist.trc.sms
$y_{denitr}$	rn_denn2o	N2O yield of denitrification	namelist.trc.sms
$y_{nitrif}$	rn₋aoun2o	N2O yield of nitrification	namelist.trc.sms

The gas exchange coefficient is calculated according to Wanninkhof (1992) (eq. 3):

$$k_w = 0.27 * v^2 * \sqrt{660./Schmidt_{gas}}$$
 (73)

where v is the amplitude of the winds (m/s), sst is the sea surface temperature, and Schmidt $_{gas}$  is the Schmidt number for each gas Wanninkhof (1992).

#### **7.1 CO**<sub>2</sub>

For the gas exchange coefficient CO<sub>2</sub> Wanninkhof (1992) include a chemical enhancement term:

$$k_w^{CO_2} = 0.27 * v^2 + 2.5 * (0.5246 + 0.016256 * sst + 0.00049946 * sst^2)$$
 (74)

For  $CO_2$ ,  $pC_{CO_2}^{air}$  is calculated from the measured mixing ratio of  $CO_2$  in the atmosphere ( $C_{CO_2}^{air}$ , in ppm) times the solubility of  $CO_2$  in sea water and corrected for 100% water vapor Sarmiento et al. (1992):

$$pC_{CO_2}^{air} = C_{CO_2}^{air} * sol_{CO_2} * (1. - e^{20.1050 - 0.0097982 * sstk - 6163.10/sstk})$$
(75)

where sstk is sea surface temperature in degree Kelvin. The solubility of  $CO_2$  is given by:

$$sol_{CO_2} = e^{c00 + c01/(sstk*.01) + c02*\ln(sstk*.01) + sal*(c03 + c04*qtt + c05*(sstk*.01)^2)} * smicr$$
 (76)

where sal is the salinity and the coefficents c00, c01, c02, c03, c04, c05 and smicr are given by Wanninkhof (1992). The Schmidt number for  $CO_2$  is given by:

$$Schmidt_{CO_2} = 2073.1 - 125.62 * sst + 3.6276 * sst^2 - 0.043126 * sst^3$$
(77)

 $C_{CO_2}^{sea}$  is the concentration of  $CO_2$  in the model, calculated based on the state variables DIC and TALK.

#### **7.2 O**<sub>2</sub>

For  $O_2$ ,  $pC_{O_2}^{air}$  is calculated from the measured mixing ratio of  $O_2$  in the atmosphere ( $C_{O_2}^{air}$ , times the solubility of  $O_2$  in seawater, also corrected for 100% water vapor as for  $CO_2$  Sarmiento et al. (1992):

$$pC_{O_2}^{air} = C_{O_2}^{air} * sol_{O_2} * (1. - e^{20.1050 - 0.0097982 * sstk - 6163.10/sstk})$$
(78)

The solubility of  $O_2$  is calculated as follows:

$$sol_{O_2} = e^{ox0+ox1/(sstk*.01)+ox2*\ln(sstk*.01)+sal*(ox3+ox4*(sstk*.01)+ox5*(sstk*.01)^2)}$$

$$* oxyco$$
(79)

The Schmidt number for  $O_2$  is given by:

$$Schmidt_{O_2} = 1953.4 - 128.0 * sst + 3.9918 * sst^2 - 0.050091 * sst^3$$
 (80)

where sal is the salinity and the coefficients ox0, ox1, ox2, ox3, ox4, ox5, and oxyco are given by Wanninkhof (1992).

Table 17: List of parameters and variables used to calculate the evolution of air-sea fluxes

Term	Variable	Description	Defined in
v	wndm	wind speed	
sal	<b>sn</b> (1)	salinity of sea surface layer	
sst	tn (1)	temperature of sea surface (°C)	
co1	c00	and other chemical constants	trcini.dgom.F90
$Schmidt_{CO_2}$	schmico2	Schmidt number for CO <sub>2</sub>	bgcflx.F90
$Schmidt_{O_2}$	schmio2	Schmidt number for O <sub>2</sub>	bgcflx.F90
$\gamma$	freeze	fraction of ocean covered by ice	limflx.F90
$\frac{O}{N}$ $mi$	atcox	pre-industrial ratio of oxygen to nitrogen	trcini.dgom.F90
$F_{air-sea}^{O}$	flu16	air-sea oxygen flux	bgcflx.F90

# 8 Model Setup

#### 8.1 Ocean General Circulation Model

The physical model NEMO v3.1 (Madec (2008),

http://www.nemo-ocean.eu/About-NEMO/Reference-manuals) was developed by the Laboratoire d' Océanographie Dynamique et de Climatologie (LODYC) to study large scale ocean circulation and its interaction with atmosphere and sea-ice. NEMO is based on the Navier-Stokes equations describing the motions of the fluid and on a non-linear equation of state, which couples the two tracers salinity and temperature to the fluid velocity.

### 8.2 Sea-Ice Model

NEMO is coupled to the Louvain-La-Neuve Sea-Ice Model (LIM, Timmermann et al., 2005), developed by Fichefet and Morales-Maqueda (1999). LIM has been thoroughly validated for both Arctic and Antarctic conditions, and has been used in a wide range of process studies. Due to the use of an elaborate technique for solving the continuity equations (Prather, 1986), LIM is particularly suited to describing the ice-edge in coarse grid resolutions, which are typically used for climate modelling studies. The physical fields that are advected in LIM are the ice concentration, the snow volume per unit area, the ice volume per unit area, the snow enthalpy per unit area, the ice enthalpy per unit area, and the brine reservoir per unit area. A full model description and details of the coupling to OPA-ORCA can be found in Timmermann et al. (2005).

#### 8.3 Forcing

#### 8.3.1 Physical Forcing

The model is forced by daily wind stress, cloud cover and precipitation from the NCEP/ NCAR reanalysed fields (Kalnay et al., 1996). Sensible and latent heat fluxes are calculated with bulk formulae using the differences between the surface temperature calculated by OPA and the observed air temperature, taking into account local humidity. At the end of each year a water balance is calculated and a uniform water flux correction is applied during the following year to conserve the water mass.

#### 8.4 Initialisation

All model simulations are initialized with observations from the World Ocean Atlas 2009 for temperature (Locarnini et al., 2010), salinity (Antonov et al., 2010)  $PO_4^{3-}$ ,  $NO_3^-$ ,  $SiO_3^-$ , (Garcia et al., 2010b) and  $O_2$  (Garcia et al., 2010a). DIC, alkalinity (GLODAP) observations were from Key et al. (2004). The biological state variables are initialised with the output from previous model runs.

#### 8.5 Dust input

The model is forced with Fe and Si input from monthly dust fluxes taken from Jickells et al. (2005) and interpolated to daily values in bgcint.F90. The input is total dust rather than in units of Fe. We assume 0.035g Fe per g of dust and either 8.8g Si per g Fe or, the equivalent, 0.308 g Si per g dust. The solubility of Fe in dust is generally taken to be 2 % and may be set in rn\_fersol . The solubility of Si in dust is 7.5 %. Using these values the dust is converted to equivalent Fe,  $Fe_{dep}$  and Si,  $Si_{dep}$  in units of mol/L/timestep in bgcbio.F90.

#### 8.6 River input

Annual fluxes of riverine carbon and nutrient (N, Si, Fe) to the ocean were computed following a global river drainage direction map (DDM30), considering population and basin area (Döll and Lehner, 2002), and river runoff (Kourzoun, 1977; Ludwig and Probst, 1998) at 0.5° increments of latitude and longitude as in da Cunha et al. (2007). This map represents the drainage directions of surface water on all continents, except Antarctica. Cells of the map are connected by their drainage directions and are thus organized into drainage basins. We use the cells corresponding to basin outlets to the ocean as input data for PlankTOM.

Values for  $DIC_{riv}$ ,  $DOC_{riv}$ ,  $POC_{riv}$ ,  $NHy_{riv}$ ,  $NOx_{riv}$ ,  $PO4_{riv}$ ,  $SIL_{riv}$  and  $Fe_{riv}$  as used in the preceding Sections are obtained by multiplying the input by the relevant parameter in Table 18. Thus all riverine inputs may be switched off by setting their parameter to zero.

In order to close the N, Si, and alkalinity cycles of the ocean, as much POM, DOM, SiO2 and CaCO3 is removed from the bottom water layer as is added by rivers and Si in dust.

#### 8.6.1 Dissolved Inorganic Nitrogen (DIN)

To calculate riverine DIN inputs we used a regression model originally developed by Smith et al. (2003):

$$\log DIN = 3.99 + 0.35 \log POP + 0.75 \log R \tag{81}$$

where (DIN) is in mol N km $^{-2}$  y $^{-1}$ , (POP) is population density in people km $^{-2}$ , and (R) is runoff in m y $^{-1}$ . The model describes DIN export by the analysis of 165 systems for which DIN flux data is available (Meybeck and A., 1997), S. Smith and F. Wulff (Eds.), LOICZ-Biogeochemical modelling node, 2000, available at http://data.ecology.su.se/MNODE/]. In this model, riverine DIN export to the coastal zone is a function of basin population density and runoff: On the basis of basin area, basin population (for the year 1990) and runoff provided by the DDM30 map, 16.3 Tg DIN y $^{-1}$  (1.16 Tmol N y $^{-1}$ ) are transported to the coastal zone by rivers. In the Smith et al. 2003 model, the average N:P ratio of riverine export is 18:1, which is close to the PISCES-T N:P ratio of 16:1. Nitrogen retention in estuarine areas was not included owing to lack of global data.

#### 8.6.2 Dissolved Silica (Si)

Rivers are responsible for 80% of the inputs of Si to the ocean (Treguer et al., 1995). For an estimate of riverine input of dissolved Si we used the runoff data from the DDM30 map, and applied an average concentration of Si in river waters of 4.2 mg Si/L (Treguer et al., 1995). Si concentration in river water is variable according to basin geology but regional data is not available. Our estimate leads to a dissolved Si river input of 187 Tg Si  $y^{-1}$  to the ocean. This value is comparable to the range of 140  $\pm$  30 Tg Si  $y^{-1}$  for a net riverine dissolved Si input to the ocean proposed by Treguer et al. (1995), considering estuarine retention of Si.

#### 8.6.3 Dissolved Iron (Fe)

Rivers and continental shelf sediments supply Fe to surface waters. Because it is extensively removed from the dissolved phase in estuaries, rivers are thought to be a minor source for the open ocean, but not for coastal zones. We used the runoff data from the DDM30 map and applied an average concentration of dissolved Fe in river waters of 40 mg  $L^{-1}$  (Martin and Meybeck, 1979; Martin and Whitfield, 1983). As for Si, river basin geology influences Fe concentration in river water, but there is no available global database on riverine Fe. Our estimate leads to a gross dissolved Fe input of 1.75 Tg Fe  $y^{-1}$ , comparable to the estimate of 1.45 Tg Fe  $y^{-1}$  by Chester (1990). During estuarine mixing, flocculation of colloidal Fe and organic matter forms particulate Fe because of the major change in ionic strength upon mixing of fresh water and seawater (de Baar and Jong, 2001). This removal has been well documented in many estuaries. Literature values show that approximately 80 to 99% of the gross dissolved Fe input is lost to the particulate phase in estuaries at low salinities (Boyle et al., 1977; Chester, 1990; Dai and Martin, 1995; Lohan and Bruland, 2006; Sholkovitz, 1978). We apply a removal rate of 99% to our gross Fe flux, and obtained a net input of riverine dissolved Fe to the coastal ocean of 0.02 Tg Fe  $y^{-1}$ .

#### 8.6.4 Particulate (POC) and Dissolved Organic (DOC) and Inorganic (DIC) Carbon

The predicted river carbon fluxes are based on models relating river carbon fluxes to their major controlling factors (Ludwig and Probst, 1998; Ludwig, 1996). For POC, sediment flux is the dominant controlling parameter. For DOC, runoff intensity, basin slope, and the amount of soil OC in the basin are the controlling parameters (Ludwig, 1996). We applied this model to the DDM30 data set, and we estimate a gross discharge of 148 Tg C  $y^{-1}$  and 189 Tg C  $y^{-1}$  for POC and DOC, respectively. We assume that DOC has a conservative behavior in estuaries. These values are in agreement with recent modeled values of 170 Tg C  $y^{-1}$  as DOC (Harrison et al., 2005), and 197 Tg C  $y^{-1}$  as POC (Beusen et al., 2005; Seitzinger et al., 2005). We used a C:N:P:Fe ratio of 122:16:1:2.44  $10^{-4}$ , thus riverine DOC and POC, when they are remineralized, are also N, P and Fe sources to the ocean. Inorganic carbon is mainly transported by rivers in the dissolved form. For DIC inputs, drainage intensity and river basin lithology are the controlling parameters (Ludwig et al., 1996). We applied this model to the DDM30 data set, and we estimate a DIC and alkalinity discharge of 385 Tg C  $y^{-1}$  (32.12 Tmol C  $y^{-1}$ ).

Table 18: List of Parameters used in river input

Term	Variable	Description	Defined in
	rn_rivdic	river input of DIC	namelist.trc.sms
	rn₋rivdoc	river input of DOC	namelist.trc.sms
	rn₋rivfer	river input of Fe	namelist.trc.sms
	rn_rivpoc	river input of POC	namelist.trc.sms
	rn_rivnit	river input of nitrate	namelist.trc.sms
	rn_rivpo4	river input of phosphate	namelist.trc.sms
	rn₋rivsil	river input of silica	namelist.trc.sms
	rn₋sedfer	coastal release of Fe	namelist.trc.sms

# 8.7 The namelist.trc.sms file

Values used for the parameters defined in *namelist.trc.sms* are given in the following tables.

Table 19: List of Parameters defined in namelist.trc.sms

Parameter	(optimised) value	Units	Description
	(and range)		•
rn_ag1poc	1.2e4	$L \text{ s (mol d)}^{-1} \text{ m}^{-2}$	small POC (POC <sub>s</sub> aggregation
rn_ag2poc	1e4	$L \text{ s (mol d)}^{-1} \text{ m}^{-2}$	$POC_s$ - large $POC$ ( $POC_l$ ) aggregation
rn_ag3poc	140	$L \pmod{d}^{-1}$	$POC_s$ - $POC_l$ aggregation
rn₋ag4poc	150	$L \pmod{d}^{-1}$	POC <sub>s</sub> aggregation
rn₋ag5doc	180	$L \text{ s (mol d)}^{-1} \text{ m}^{-2}$	$DOC - POC_s$ aggregation
rn_ag6doc	3.9e3	L s (mol d) $^{-1}$ m $^{-2}$	$DOC - POC_l$ aggregation
rn_alpphy	1.e-6	mol C m <sup>2</sup> (g Chl	initial slope of photsyntheses vs light intensity curve
,		mol photons) <sup>-1</sup>	
rn₋aoun2o	1.23e-4	mol N2O (mol NH4) <sup>-1</sup>	N2O yield nitrification
	(0.37e-4 - 2.53e-4)		
rn₋aoun2s	1.06e-5	mol N2O (mol O2) <sup>-1</sup>	oxic N2S yield
	(0.33e-5 - 2.26e-5)	(	
rn_betn2s	1.7e-3	mol N2O (mol O2) <sup>-1</sup>	suboxic N2S yield
	(1.7e-3 - 10.18e-3)		
rn_coccal	0.433	_	ratio of CaCO <sub>3</sub> to organic carbon
rn_degn2o	0	mol N2O (mol NO3) <sup>-1</sup>	yield N2O consumption
	(0 - 9.65e-2)		J · · · · · · · · · · · · · · · · · · ·
rn₋denn2o	3.4e-3	mol N2O (mol NO3) <sup>-1</sup>	N2O yield denitrification
	(3.4e-3 - 80.8e-3)		
rn_domphy	0.45	_	maximum DOC excretion ratio for all phyto
rn₋discal	0.75	_	fraction of CaCO <sub>3</sub> dissolved during coccolithophore
			mortality
rn_docphy	0.05	_	excretion ratio for all phyto
rn₋ekwgrn	0.0232	$m^{-1}$	green light absorption coefficient of H <sub>2</sub> O
rn_ekwred	0.225	$\begin{array}{c c} m \\ m^{-1} \end{array}$	red light absorption coefficient of H <sub>2</sub> O
rn_etomax	80.	$ m W~m^{-2}$	maximum surface insolation
rn_faco18	0.98	-	bacterial fractionation for O <sub>18</sub>
rn_fersol	0.01	_	solubility of iron in dust
rn_gbadoc	0.088	_	relative preference of BAC grazing for DOC
rn_gbagoc	8.76	_	relative preference of BAC grazing for GOC
rn_gbapoc	8.76	_	relative preference of BAC grazing for POC
rn_ggebac	.21	_	growth efficiency BAC
rn_ggemac	0.3	_	growth efficiency MAC
rn_ggemes	0.25	_	growth efficiency MES
rn_ggemic	0.29	_	growth efficiency PRO
rn_gmabac	0.186	_	relative preference of MAC grazing for BAC
rn_gmagoc	0.186	_	relative preference of MAC grazing for GOC
rn_gmames	1.860	_	relative preference of MAC grazing for MES
rn_gmamic	1.860	_	relative preference of MAC grazing for PRO
rn_gmaphy	1.860	_	relative preference of MAC for DIA
<u>.</u> gapy	1.860	_	relative preference of MAC for MIX
	1.860	_	relative preference of MAC for COC
	.930	_	relative preference of MAC for PIC
	1.860	_	relative preference of MAC for PHA
	.186	_	relative preference of MAC for FIX
rn_gmapoc	0.186	_	relative preference of MES grazing for POC
rn_gmebac	.165	_	relative preference of MES grazing for BAC
rn_gmegoc	0.165	_	relative preference of MES grazing for GOC
g2g00	1 3.100	<u> </u>	Continued on next page
			Continued on next page

29

Table 19 – continued from previous page

		Table 19 – continued fr	
Parameter	Value	Units	Description
rn₋gmemic	3.302	-	relative preference of MES grazing for PRO
rn_gmephy	1.651	-	relative preference of MES for DIA
	1.238	-	relative preference of MES for MIX
	1.238	-	relative preference of MES for COC
	1.238	-	relative preference of MES for PIC
	1.238	-	relative preference of MES for PHA
	0.165	-	relative preference of MES for FIX
rn_gmepoc	0.165	_	relative preference of MES grazing for POC
rn_gmibac	2.480	_	relative preference of PRO grazing for BAC
rn_gmigoc	0.062	_	relative preference of PRO grazing for GOC
rn_gmiphy	0.620	_	relative preference of MIC for DIA
	1.240	_	relative preference of MIC for MIX
	1.240	_	relative preference of MIC for COC
	1.240	_	relative preference of MIC for PIC
	1.240	_	relative preference of MIC for PHA
	1.240	_	relative preference of MIC for FIX
rn_gmipoc	0.062		relative preference of PRO grazing for POC
	3.15	$d^{-1}$	
rn_grabac		$\begin{pmatrix} \mathbf{d} \\ \mathbf{d}^{-1} \end{pmatrix}$	maximum BAC uptake rate
rn_gramac	0.106	$d$ $d^{-1}$	maximum MAC grazing rate
rn_grames	1.22		maximum MES grazing rate
rn_gramic	1.59	$d^{-1}$	maximum PRO grazing rate
rn_grkmac	9.e-6	$\mod L^{-1}$	$K_m$ for MAC grazing
rn_grkmes	10.e-6	$\mod L^{-1}$	$K_m$ for MES grazing
rn_grkmic	10.e-6	$\mod L^{-1}$	$K_m$ for PRO grazing
rn_icemac	100.0	%	MAC enhanced recruitment under ice
rn_kgrphy	.0118	$L (m g Chl)^{-1}$	light absorption in blue-green for DIA
	.0257	$L (m g Chl)^{-1}$	light absorption in blue-green for MIX
	.0257	$L (m g Chl)^{-1}$	light absorption in blue-green for COC
	.0696	$L (m g Chl)^{-1}$	light absorption in blue-green for PIC
	.0257	$L (m g Chl)^{-1}$	light absorption in blue-green for PHA
	.0657	$L (m g Chl)^{-1}$	light absorption in blue-green for FIX
rn₋kmfbac	0.025e-9	$\mod L^{-1}$	$K_m$ for Fe in DOC remineralisation by bacteria
rn_kmfphy	40.e-9	$\mod L^{-1}$	${\sf K}_m^{Fe}$ for DIA ${\sf K}_m^{Fe}$ for MIX
	25.e-9	$\mod L^{-1}$	$K_{m}^{\widetilde{F}e}$ for MIX
	25.e-9	$\mod L^{-1}$	$K_m^{\widetilde{r}_e}$ for COC
	10.e-9	$\mod L^{-1}$	
	25.e-9	$\mathrm{mol}\ \mathrm{L}^{-1}$	$K^{m}_{Fe}$ for PHA
	40.e-9	$\mod L^{-1}$	$K^{Fe}$ for FIX
rn_kmhnit	0.1e-6	$\mod L^{-1}$	$K^{NH4}$ nitrification
rn_kmhphy	5.e-6	$\mod L$	$K^{m}$ intrinduction $K^{NH4}$ for DIA
TIT_KITITIPITY	0.5e-6	$\mod L$ $\mod L^{-1}$	$K_m$ for DIA $K^{NH4}$ for MIY
	0.5e-6	$\mod L$ $\mod L^{-1}$	K <sub>m</sub> for WIX
	0.3e-6	$\mod L$ $\mod L^{-1}$	N <sub>m</sub> for COC
		$\mod L$ $\mod L^{-1}$	$N_m$ 101 PIC
	1.5e-6	$\begin{array}{c c} \operatorname{mol} L & \\ \operatorname{mol} L^{-1} \end{array}$	$K_m^{Fe}$ for PIC $K_m^{Fe}$ for PHA $K_m^{Fe}$ for FIX $K_m^{NH4}$ nitrification $K_m^{NH4}$ for DIA $K_m^{NH4}$ for MIX $K_m^{NH4}$ for PIC $K_m^{NH4}$ for PIC $K_m^{NH4}$ for PHA $K_m^{NH4}$ for PIX $K_m^{NH4}$ for PHA $K_m^{NH4}$ for FIX $K_m^{NO3}$ for DIA $K_m^{NO3}$ for DIA $K_m^{NO3}$ for DIA $K_m^{NO3}$ for PIC $K_m^{NO3}$ for PIX $K_m^{NO3}$ for PIA $K_m^{NO3}$ for PIA $K_m^{NO3}$ for PIOC in DOC remineralisation by bacteria
	0.3e-6		$N_{m}^{-1}$ 10f PIA
rn_kmnphy	2.e-6	$\mod L^{-1}$	$K_m^{\sim}$ Tor DIA
	2.0e-6	$\mod L^{-1}$	$K_m^{NO3}$ for MIX
	2.0e-6	$\mod L^{-1}$	$K_m^{NO3}$ for COC
	2.0e-6	$\mod L^{-1}$	$K_m^{AVO3}$ for PIC
	3.0e-6	$\mod L^{-1}$	$K_m^{NO3}$ for PHA
	13.0e-6	$\mod L^{-1}$	$K_m^{NO3}$ for FIX
rn₋kmobac	1e-7	$\mod L^{-1}$	···
rn₋kmpbac	1e-7	$\mod L^{-1}$	$K_m$ for $PO_4$
rn_kmpphy	7.6e-6	$\mod L^{-1}$	$K_m^{PO_4}$ for DIA
			Continued on next page

30

Table 19 – continued from previous page				
Parameter	Value	Units	Description	
	12.2e-6	$mol L^{-1}$	$\mathbf{K}_{m}^{PO_{4}}$ for MIX $\mathbf{K}_{m}^{PO_{4}}$ for COC $\mathbf{K}_{m}^{PO_{4}}$ for PIC $\mathbf{K}_{m}^{PO_{4}}$ for PHA $\mathbf{K}_{m}^{PO_{4}}$ for FIX	
	15.9e-6	$\mod L^{-1}$	$K_m^{PO_4}$ for COC	
	15.9e-6	$\mod L^{-1}$	$K_m^{PO_4}$ for PIC	
	97.6e-6	$\mod L^{-1}$	$K_{m}^{PO_{4}}$ for PHA	
	24.4e-6	$\mod L^{-1}$	$K_{m}^{PO_{4}}$ for FIX	
rn_kmsbsi	20e-6	$\mod L^{-1}$	$K_m$ for the Si/C ratio of DIA	
rn_krdphy	.0056	$L (m g Chl)^{-1}$	light absorption in red for DIA	
	.0098	$L (m g Chl)^{-1}$	light absorption in red for MIX	
	.0098	$L \text{ (m g Chl)}^{-1}$	light absorption in red for COC	
	.0197	$L (m g Chl)^{-1}$	light absorption in red for PIC	
	.0098	$L (m g Chl)^{-1}$	light absorption in red for PHA	
	.0181	$L (m g Chl)^{-1}$	light absorption in red for FIX	
rn₋lyscal	10e-5	$\mod L^{-1}$	inertia conc. for CaCO <sub>3</sub> dissolution	
rn₋mormac	0.020	$d^{-1}$	MAC mortality rate	
rn₋mormes	0.040	$d^{-1}$	MES mortality rate	
rn_motmac	1.0481	-	temp. dependence of MAC mortality	
rn_motmes	1.1161	-	temp. dependence of MES mortality	
rn_mumpft	0.44	$d^{-1}$	maximum growth rate DIA	
	0.35	$d^{-1}$	maximum growth rate MIX	
	0.70	$d^{-1}$	maximum growth rate COC	
	0.26	$d^{-1}$	maximum growth rate PIC	
	0.68	$d^{-1}$	maximum growth rate PHA	
	0.046	$d^{-1}$	maximum growth rate FIX	
rn_munfix	0.56	-	fraction of growth rate during N2fix relative to	
			growth on NO3	
rn_mutpft	1.0400	-	temp. dependence of proto-zooplankton	
	1.0242	-	temp. dependence of meso-zooplankton	
	1.1165	-	temp. dependence of macro-zooplankton	
	1.0680	-	temp. dependence of DIA	
	1.0461	-	temp. dependence of MIX	
	1.0132	-	temp. dependence of COC	
	1.0611	-	temp. dependence of PIC	
	1.0520	-	temp. dependence of PHA	
	1.0623	-	temp. dependence of FIX	
	1.0379	-	temp. dependence of BAC	
rn_nitnh4	0.79	$d^{-1}$	maximum nitrification rate	
rn₋qmaphy	2.e-7	-	maximum quota for Fe for all phyto	
rn_qmiphy	4.0e-6	-	minimum quota for Fe for all phyto	
rn_qopphy	8.6e-6	- ,	optimal quota for Fe for all phyto	
rn₋resbac	0.10	$d^{-1}$	BAC respiration at 0°C	
rn₋resmac	0.018	$d^{-1}$	MAC respiration at 0°C	
rn₋resmes	0.028	$d^{-1}$	MES respiration at 0°C	
rn₋resmic	0.010	$d^{-1}$	PRO respiration at 0°C	
rn_resphy	0.012	-	fractional phytoplankton loss rate: DIA	
	0.15	-	fractional phytoplankton loss rate: MIX	
	0.15	-	fractional phytoplankton loss rate: COC	
	0.15	-	fractional phytoplankton loss rate: PIC	
	0.15	-	fractional phytoplankton loss rate: PHA	
	0.15	-	fractional phytoplankton loss rate: FIX	
rn₋retbac	1.0494	-	temp. dependence of BAC respiration	
rn₋retmac	1.0942	-	temp. dependence of MAC respiration	
rn_retmes	1.0887	-	temp. dependence of MES respiration	
rn_retmic	1.0897	-	temp. dependence of PRO respiration	
rn_rhfphy	29.	-	maximum/minimum Fe uptake rate	
			Continued on next page	

Table 19 – continued from previous page

Parameter	Value	Units	Description
rn_rivdic	1.	-	(1 - estuarine retention fraction) of river DIC
rn₋rivdoc	1.	-	(1 - estuarine retention fraction) of river DOC
rn₋rivpoc	0.55	-	(1 - estuarine retention fraction) of river POC
rn_rivpo4	1.	-	(1 - estuarine retention fraction) of river PO <sub>4</sub>
rn_rivsil	1.	-	(1 - estuarine retention fraction) of river SIL
rn_rivfer	0.25	-	(1 - estuarine retention fraction) of river FER
rn_scofer	1.e-3	$\pmod{L^{-1}}^{-0.6} d^{-1}$	scavenging of Fe
rn_scmfer	1.e-3	$\pmod{L^{-1}}^{-0.6} d^{-1}$	minimum scavenging of Fe
rn_sedfer	1e-11	$\mod L^{-1}$	coastal release of Fe
rn₋sigmac	0.70	-	fraction of MAC excretion as PO <sub>4</sub>
rn₋sigmes	0.68	-	fraction of MES excretion as PO <sub>4</sub>
rn₋sigmic	0.66	-	fraction of PRO excretion as DOM
rn₋sildia	0.42e-6	$\mod L^{-1}$	$K_m^{SiO_3}$ for diatoms
rn₋singoc	0.0303	$m^2 (kg d)^{-1}$	Sinking rate parameter of $POC_l$ , $CaCO_3$ and $DSi$
rn_snkgoc	0.6923	-	sinking rate parameter of $POC_l$ , $CaCO_3$ and $SiO_2$
rn_snkpoc	3.0	$m d^{-1}$	sinking speed of $POC_s$
rn_thmphy	0.7	$  g \text{ mol}^{-1}  $	maximum CHL:C ratio for DIA
	0.4	$\mid$ g mol <sup>-1</sup>	maximum CHL:C ratio for MIX
	0.4	$g \text{ mol}^{-1}$	maximum CHL:C ratio for COC
	0.4	$\mid$ g mol <sup>-1</sup>	maximum CHL:C ratio for PIC
	0.5	$\mid$ g mol <sup>-1</sup>	maximum CHL:C ratio for PHA
	0.3	$ g \text{ mol}^{-1} $	maximum CHL:C ratio for FIX
rn₋unamac	0.18	-	unassimilated fraction of phyto during MAC grazing
rn_unames	0.3	-	unassimilated fraction of phyto during MES grazing
rn₋unamic	0.13	-	unassimilated fraction of phyto during PRO grazing

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