



Supplement of

All about nitrite: exploring nitrite sources and sinks in the eastern tropical North Pacific oxygen minimum zone

John C. Tracey et al.

Correspondence to: John C. Tracey (jt16@princeton.edu)

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Correspondence to: John C. Tracey (jt16@alumni.princeton.edu)

Supplementary Material:

Additional Information on Anammox and Denitrification Profiles (SR1805 and FK180624)

In order to calculate the denitrification and anammox rates using Eq. (7 – 9) , the amount of 29 and 30 N₂ in nmol for each vial must first be calculated using the equations below:

$${}^{29}\text{N}_{2\text{nmol}} = (\text{Slope}_{\text{STD Curve}}) * \frac{29}{28_{AM}} * \widehat{28}_{\text{N}_2\text{Peak Area}} \quad (\text{S1})$$

$${}^{30}\text{N}_{2\text{nmol}} = (\text{Slope}_{\text{STD Curve}}) * \frac{30}{28_{AM}} * \widehat{28}_{\text{N}_2\text{Peak Area}} \quad (\text{S2})$$

Where:

$\frac{30}{28_{AM}}$ or $\frac{29}{28_{AM}}$ is the measured ratio in peak area units of the specific sample,

$\widehat{28}_{\text{N}_2\text{Peak Area}}$ is the average 28 peak area across the time course (without outliers), and

$Slope_{STD\ Curve}$ is the standard curve slope in nmol / peak area units.

Outliers were defined as values more than three standard deviations away from the raw time course mean. Outlier points were not used to calculate the average 28 N₂ peak used in Eqs. (S1-2). The average 28 peak area across the time course was used as a data smoothing technique to remove variation due to small differences in the water volume aliquoted into each exetainer or to small amounts of N₂ that escaped purging.

After removing outlying 28 N₂ points, drift corrections, if necessary, were performed as follows. First, the 29 / 28 and 30 / 28 ratios of the standards from each mass spectrometer (MS) run were regressed using both linear and logarithmic models against the total N of each sample. If the regressions returned slopes significantly different from zero via a Student's T test (p values <0.05), a drift correction was performed using the best regression (as determined by R² and the residual mean square error (RMSE) values). Corrections were performed by using the best significant regression to calculate a theoretical value for each total N point. After this the difference between an "endpoint" (the endpoint varied from run to run - the highest or lowest theoretical value for one of the standards was used) and each theoretical value was calculated for every point. This difference was then added to the raw sample and standard values to correct for drift. After performing total N drift corrections, corrected ratios were re-plotted against total N to examine if the correction was successful. After performing a total N correction, samples were examined for drift due to the time in the mass spectrometer run. This was performed in an analogous manner to the total N drift correction.

Drift corrected 29 and 30 N₂ produced values were then checked for outliers across each time course. As above, points more than three standard deviations away from the time course

mean were discarded. After this, the drift corrected, outlier screened 29 and 30 N₂ produced values were used to calculate rates via Eq. (7 – 9). Importantly, the rates reported here were not all calculated using regressions from t₀ – t₄. Instead, regressions from t₀ – t₂, t₀ – t₃, and t₀ – t₄ were compared and the best regression was selected based on a combination of the R² and RMSE value. Rates due to a bottle effect (i.e., exponential growth of an N cycling microbe at the end of the incubation because incubation conditions changed over the course of the experiment to favor exponential growth) were removed by comparing linear and exponential models for any suspected time courses. If the exponential model was a better fit, the rate from that time course was not reported and the next shorter significant time course (t₀ – t₂, or t₀ – t₃) for that incubation was used.

For the SR1805 cruise, anammox rates calculated from the ¹⁵NH₄⁺ and ¹⁵NO₂⁻ tracers were similar in magnitude (Supplementary Table S3). As a result, the SR1805 anammox rates reported in Figs. 2, 3, 6, 7, 8, and 9 represent either the average of the rates calculated from the ¹⁵NH₄⁺ and ¹⁵NO₂⁻ tracers (if significant rates were returned for both tracers) or the rate from the tracer that returned a significant or positive rate with error bars above zero. The FK180624 anammox rates reported in Fig. S1 are from the ¹⁵NO₂⁻ tracer alone.

Additional Information for NO₂⁻ Oxidation Rate Measurements

Methods Details for Depth Profiles

The denitrifying bacterium culture seed (*Pseudomonas chloroaphis*) was prepared aerobically and frozen until needed. The denitrifiers were allowed to grow to high biomass for approximately five days so all NO₃⁻ in the growth medium was consumed. The denitrifiers were then concentrated through centrifugation and resuspended in a medium that lacked NO₃⁻. The

concentrate was distributed in 20 mL Grace vials (Manufacturer: Thermo Fisher Scientific, Bremen, Germany), sealed, and purged of O₂ with He for 3 – 5 hours. Grace vials were chosen because their straight shape allows for minimum O₂ contamination while purging. After removing N₂, exetainers were prepared for the denitrifier method by subsampling 1 mL and then adding 30 μL sulfamic acid to consume any residual ambient or tracer NO₂⁻ present. After this, 30 μL of 5 N NaOH was added to return each sample to its original pH (Granger, J., & Sigman, 2009). The Grace vials were then injected with 1 mL of the sulfamic acid treated seawater samples and incubated overnight in the dark to convert NO₃⁻ to N₂O. Five to six drops of 10 N NaOH was added to terminate the reactions in the vials, and 5-6 drops of Antifoam B were added so the solution would not foam when purged with helium by the mass spectrometer. Drift corrections, mechanism for selecting the best time course from t₀ – t₂, t₀ – t₃, and t₀ – t₄, and checks for bottle effects were conducted in similar ways to those described above for anammox and denitrification depth profiles.

Method Details for Dismutation Experiments

Measurements of anammox, denitrification, and NO₂⁻ oxidation for the dismutation tests were conducted as above except for the following changes:

- (1) For the ¹⁵NO₃⁻ measurements of the dismutation experiments, 200 μL of acidified seawater sample was injected into each Grace vial instead of the 1 mL used in the depth profiles.
- (2) For the anammox and denitrification measurements of the dismutation experiments, unlike in the depth profile experiments, each replicate within the 5 points of our 15 member time course was run on separate MS runs (i.e. t₀ point 1, t₁ point 1, t₂ point 1, t₃ point 1, and t₄ point 1, were run together while t₀ point 2, t₁ point 2, t₂ point 2, t₃ point 2, and t₄ point 2 and t₀ point 3, t₁ point 3, t₂ point 3, t₃ point 3, and t₄ point 3

were run on separate runs. Samples were corrected for time and linearity in each separate MS run as in the depth profile experiments. After performing these corrections, the fifteen points from each time course were combined; however, the resulting time courses often had widely disparate ranges. In order to reduce the error of the rate regression, the range between mini time courses (i.e., $t_{0 \text{ point } 1}$, $t_{1 \text{ point } 1}$, $t_{2 \text{ point } 1}$, $t_{3 \text{ point } 1}$, and $t_{4 \text{ point } 1}$ is one “mini” course out of three per depth), was condensed using the following method:

- (a) Linear regressions for each of the mini time courses were calculated.
- (b) Using these regressions, theoretical values at each point in the mini time courses were calculated.
- (c) The average of these theoretical values for each of the three mini time courses was calculated.
- (d) A linear regression for the whole time course was calculated.
- (e) Using the whole time course regression, theoretical values at each of the five time points in the full length time course was calculated.
- (f) The average of these five theoretical values was calculated.
- (g) The difference between each mini time course’s average and the full length average was calculated.
- (h) This difference was added to each point in the run to condense the mini time courses’ range.

The remaining calculations were performed as in the depth profile experiments.

Station	OMZ region	Bottom Z (m)	Coordinates	Depths (m)	Feature
PS1	Offshore	4100	10N 113W	30	Oxic surface
				60	Oxycline
				70	Oxycline
				90	ODZ top
				100	ODZ top
				110	ODZ top
				150	ODZ top
				260	ODZ core
				500	ODZ core
				1000	Oxycline
PS2	Open ocean	3000	15.76N 105W	60	Oxic surface
				75	Oxycline
				95	ODZ top
				120	ODZ top
				150	ODZ top
				200	ODZ top/SNM
				250	ODZ core/SNM
				300	ODZ core
				500	ODZ core
				850	ODZ core
PS3	Coastal	1030	17.68N 102.35W	16	Oxycline
				25	Oxycline
				35	Oxycline
				45	ODZ top
				60	ODZ top
				70	ODZ top
				100	ODZ top
				160	ODZ top/SNM
				250	ODZ core
			800	ODZ core	

Supplementary Table S1: Sampling depths and geography for NH_4^+ oxidation, NO_3^- reduction, NO_2^- oxidation, anammox, and denitrification profiles (SR1805). Features are classified based on their potential density and O_2 concentrations. ODZ core samples have a $\sigma_\theta > 26.4$ (Babbin et al., 2020). ODZ top, oxycline, and oxic surface samples have a $\sigma_\theta < 26.4$ (Babbin et al., 2020).

Station	OMZ region	Bottom Z (m)	Coordinates	Depths (m)	Feature
2	Open ocean	3450	14N, 103W	102	Oxycline
				105	ODZ top
				120	ODZ top
				130	ODZ top
				200	ODZ top/SNM
				395	ODZ core
3	Open ocean	3000	14N, 104W	120	ODZ top
				140	ODZ top
				170	ODZ top
				250	ODZ core
9	Open ocean	3250	14N, 110W	96	Oxycline
				100	Oxycline
				105	ODZ top
				115	ODZ top
				130	ODZ top
				160	ODZ top/SNM
				168	ODZ core/SNM
				175	ODZ core/SNM
				200	ODZ core/SNM
				250	ODZ core/SNM
				300	ODZ core
				375	ODZ core
				475	ODZ core
				730	ODZ core
11	Open ocean	4000	14N, 112W	116	ODZ top
				320	ODZ core
12	Open ocean	4100	14N, 113W	103	ODZ top
				120	ODZ top
				150	ODZ top/SNM
				610	ODZ core
14	Open ocean	4100	14N, 115W	125	ODZ top/SNM
				323	ODZ core
15	Open ocean	4200	14N, 116W	135	ODZ top
				160	ODZ top
18	OMZ boundary	4100	17N, 119W	130	Oxycline
				160	Oxycline
				200	ODZ core

Supplementary Table S2: Sampling depths and geography for anammox, and denitrification profiles (FK180624). Features are classified based on their potential density and O₂ concentrations. ODZ core samples have a $\sigma_{\theta} > 26.4$ (Babbin et al., 2020). ODZ top, oxycline, and oxic surface samples have a $\sigma_{\theta} < 26.4$ (Babbin et al., 2020).

Depth (m)	Denitrification		Anammox ($^{15}\text{NH}_4^+$)		Anammox ($^{15}\text{NO}_2^-$)		Consensus anammox	
	Rate	Error	Rate	Error	Rate	Error	Rate	Error
PS1								
30	0	0	0	0	0	0	0	0
60	3.9	3.2	0	0	0	0	0	0
70	4.8	2.8	0	0	3.2	3.0	3.2	3.0
100	0	0	0.71	0.53	2.6	0.98	2.6	0.98
150	0	0	2.2	1.0	0.36	0.11	0.36	0.11
260	0	0	3.8	1.3	0.36	0.16	2.1	0.74
500	2.5	0.98	0	0	5.0	2.2	5.0	2.2
1000	0	0	0.74	0.7	0	0	0.74	0.7
PS2								
120	0	0	1.2	0.86	0	0	1.2	0.86
150	0	0	0	0	2.9	2.3	2.9	2.3
200	7.2	2.5	1.6	0.46	0	0	1.6	0.46
250	3.3	2.0	4.3	1.1	4.1	1.2	4.2	1.2
300	1.6	0.58	0	0	0	0	0	0
500	0	0	0	0	0.19	0.11	0.19	0.11
850	0	0	4.5	1.4	0	0	4.5	1.4
PS3								
16	0.8	0.53	0	0	0.52	0.41	0.52	0.41
25	1.2	0.33	0	0	1.0	0.35	1.0	0.35
35	0	0	0	0	0.53	0.27	0.53	0.27
45	0	0	0.24	0.21	0	0	0.24	0.21
60	0.33	0.28	1.2	0.32	0.89	0.29	1.1	0.31
70	0.7	0.33	0.83	0.29	1.0	0.18	0.93	0.23
100	0	0	1.7	0.31	1.3	0.4	1.5	0.35
160	1.9	0.89	0.69	0.21	2.4	0.8	1.5	0.5
250	0	0	1.2	0.35	0.94	0.71	1.2	0.35
800	0.47	0.33	0.58	0.16	0.34	0.26	0.58	0.16

Supplementary Table S3: Comparison of anammox rates calculated from $^{15}\text{NH}_4^+$ and $^{15}\text{NO}_2^-$ and denitrification rates from SR1805 cruise. Rates in bold are significantly different from zero. The consensus anammox rate was determined by (1) averaging the rates from the two tracers if both tracers returned a significant positive rate or (2) taking the rate from the tracer that returned a significant rate if the other tracer returned an insignificant rate or (3) taking the rate from the tracer that returned an insignificant but positive rate with error bars above zero if the other tracer returned a rate of zero.

Cruise	Station	Target O ₂ (nM)	Depth (m)	Cast	Ambient O ₂ (μM) Unnormalized
SR1805	PS1	1	93	4	-0.87
SR1805	PS1	10	110	10	-0.833
SR1805	PS1	100	110	14	-0.257
SR1805	PS1	1000	110	18	-0.549
SR1805	PS1	10000	100	23	-0.026
SR1805	PS2	1	100	31	-0.76
SR1805	PS2	10	113	37	-0.096
SR1805	PS2	100	130	42	-0.878
SR1805	PS2	1000	120	47	-0.016
SR1805	PS2	10000	110	50	-0.777
SR1805	PS3	1	60	70	0.274
SR1805	PS3	10	45	76	0.042
SR1805	PS3	100	50	81	-0.806
SR1805	PS3	1000	46	85	-0.795
SR1805	PS3	10000	60	88	-0.859
FK180624	2	10	120	2	1.329
FK180624	2	1000	120	2	1.329
FK180624	2	100	120	4	1.174
FK180624	2	3000	120	4	1.174
FK180624	9	1000	110	Aft 67	2.0
FK180624	9	1000	105	Frank 197	2.2
FK180624	9	3000	120	9	1.09
FK180624	15	100	110	1	1.633
FK180624	18	10000	105	1	12.138
FK180624	2	10	120	2	1.329
FK180624	2	1000	120	2	1.329
FK180624	2	100	120	4	1.174
FK180624	2	3000	120	4	1.174
FK180624	9	1000	110	Aft 67	2.0
FK180624	9	1000	105	Frank 197	2.2
FK180624	9	3000	120	9	1.09
FK180624	15	100	110	1	1.633
FK180624	18	10000	105	1	12.138

Supplementary Table S4: Ambient O₂ concentrations for the sample water used in the NO₂⁻ oxidation O₂ manipulation experiments on the SR1805 and FK180624 cruises.

Station	Depth (m)	NH ₄ ⁺ oxidation	Maximum N loss from NH ₄ ⁺ oxidation	Anammox	Denitrification	Total N loss	Percent N loss due to NH ₄ ⁺ oxidation	O ₂
PS1	30	0.0010	0.0005	0.00	0.00	0.00	100	199.2
PS1	60	0.8392	0.4196	0.00	3.9	3.9	10.7	108.7
PS1	70	1.0803	0.5402	3.2	4.8	8.0	6.7	21.9
PS1	90	0.2605	0.1302	0.0	0.0	0.0	100	0.2
PS1	100	0.3070	0.1535	2.6	0.0	2.6	5.9	0.5
PS1	110	0.0280	0.0140	0.00	0.00	0.00	100	-0.1
PS1	150	0.0014	0.0007	0.36	0.00	0.36	0.2	0.4
PS1	260	0.0005	0.0003	2.1	0.0	2.1	0.0	2.9
PS1	500	0.0002	0.0001	5.0	2.5	7.6	0.0	-0.1
PS1	1000	0.0002	0.0001	0.7	0.0	0.7	0.0	30.1
PS2	60	0.0293	0.0146	0.0	0.0	0.0	100	174.5
PS2	75	2.7476	1.3738	0.0	0.0	0.0	100	47.0
PS2	95	0.9789	0.4894	0.0	0.0	0.0	100	3.2
PS2	120	0.0093	0.0046	1.2	0.0	1.2	0.4	-0.1
PS2	150	0.0024	0.0012	2.9	0.0	2.9	0.0	-0.1
PS2	200	0.0025	0.0012	1.6	7.2	8.7	0.0	0.0
PS2	250	0.0	0.0	4.2	3.3	7.5	0.0	1.2
PS2	300	0.0	0.0	0.00	1.6	1.6	0.0	-0.2
PS2	500	0.0005	0.0003	0.19	0.00	0.19	0.1	0.1
PS2	850	0.0	0.0	4.5	0.0	4.5	0.0	1.5
PS3	16	0.0	0.0	0.52	0.80	1.3	0.0	90.2
PS3	25	11.7272	5.8636	1.0	1.2	2.2	262	27.7
PS3	35	6.3850	3.1925	0.53	0.00	0.53	601	1.9
PS3	45	0.0	0.0	0.24	0.00	0.24	0.0	-0.2

Station	Depth (m)	NH ₄ ⁺ oxidation	Maximum N loss from NH ₄ ⁺ oxidation	Anammox	Denitrification	Total N loss	Percent N loss due to NH ₄ ⁺ oxidation	O ₂
PS3	60	0.1832	0.0916	1.1	0.33	1.4	6.6	-0.3
PS3	70	0.00	0.0	0.93	0.70	1.6	0.0	0.0
PS3	100	0.0294	0.0147	1.5	0.0	1.5	1.0	0.1
PS3	160	0.0061	0.0031	1.5	1.9	3.4	0.1	0.0
PS3	250	0.0000	0.0000	1.2	0.0	1.2	0.0	0.0
PS3	800	0.0345	0.0173	0.58	0.47	1.1	1.6	0.3

Supplementary Table S5: N loss due to NH₄⁺ oxidation does not contribute a large fraction (> 10% of total N loss) of the total N loss except in oxic depths or depths where denitrification and anammox are low (< 1 nM N₂ d⁻¹). NH₄⁺ oxidation rates and the maximum N loss from NH₄⁺ oxidation rates are shown in nmol N d⁻¹ and nmol N₂ d⁻¹, respectively. Maximum N loss from NH₄⁺ oxidation rates were calculated by unrealistically assuming that all ¹⁵N-NO₂⁻ produced in NH₄⁺ oxidation was converted to N₂. Anammox, denitrification, and total N loss are shown in nmol N₂ d⁻¹ and O₂ is shown in μM. Raw NH₄⁺ oxidation, anammox, and denitrification rates that are significantly different from zero are shown in bold.

Regression	Slope	Error	R²	P value	Line type	Color
Shallow boundary – all data	0.56	0.059	0.55	1.4e-14	Solid	Orange
ODZ core – all data	0.45	0.053	0.54	9.5e-12	Solid	Teal
Shallow boundary – 2018 only	0.29	0.10	0.16	0.009	Dashed	Orange
ODZ core – 2018 only	0.35	0.038	0.79	2.5e-9	Dashed	Teal
Shallow boundary – 2012-13 only	0.64	0.073	0.69	3.1e-10	Dotted	Orange
ODZ core – 2012-13 only	1.1	0.13	0.68	2.3e-10	Dotted	Teal

Supplementary Table S6: Regression statistics for Fig. 9A (plot of denitrification vs. anammox rates).

Station	Depth (m)	AMX Rate (nM N ₂ d ⁻¹)	DN Rate (nM N ₂ d ⁻¹)	NO ₂ ⁻ (μM)	NH ₄ ⁺ (nM)	Added ¹⁵ NO ₂ ⁻ (μM)	Added ¹⁵ NH ₄ ⁺ (nM)	NH ₄ ⁺ Enrichment	NO ₂ ⁻ Enrichment
PS1	30	0.00	0.00	0.02	3.15	3	3000	953.1	188.5
PS1	60	0.00	0.00	1.37	31.36	3	3000	96.7	3.2
PS1	70	0.00	0.00	0.30	4.46	3	3000	673.0	11.1
PS1	90	0.00	0.00	0.01	45.93	3	3000	66.3	380.7
PS1	100	2.61	0.00	0.01	32.99	3	3000	91.9	268.9
PS1	110	0.00	0.00	0.02	2.98	3	3000	1008.9	188.5
PS1	150	0.36	0.00	0.00	2.11	3	3000	1422.9	2001.0
PS1	260	2.07	0.00	0.06	1.98	3	3000	1513.9	53.8
PS1	500	0.00	2.53	0.01	3.78	3	3000	794.3	527.3
PS1	1000	0.00	0.00	0.02	5.56	3	3000	540.2	159.7
PS2	60	0.00	0.00	0.05	35.82	3	3000	84.7	63.6
PS2	75	0.00	0.00	0.13	13.60	3	3000	221.5	24.8
PS2	95	0.00	0.00	0.06	13.65	3	3000	220.8	51.1
PS2	120	0.00	0.00	0.28	23.53	3	3000	128.5	11.7
PS2	150	0.00	0.00	1.16	2.13	3	3000	1409.4	3.6
PS2	200	1.56	7.15	1.82	1.81	3	3000	1657.7	2.6
PS2	250	4.17	0.00	1.92	5.03	3	3000	597.0	2.6
PS2	300	0.00	1.60	1.45	6.86	3	3000	438.6	3.1
PS2	500	0.00	0.00	0.00	3.65	3	3000	823.4	*
PS2	850	4.49	0.00	0.00	1.46	3	3000	2052.6	*
PS3	16	0.00	0.00	0.08	302.03	3	3000	10.9	37.4
PS3	25	1.05	1.19	0.58	250.57	3	3000	13.0	6.2
PS3	35	0.00	0.00	0.27	9.49	3	3000	317.0	12.1
PS3	45	0.00	0.00	0.23	8.85	3	3000	340.1	14.2
PS3	60	1.06	0.00	0.25	1.37	3	3000	2196.9	13.0
PS3	70	0.93	0.00	0.16	4.26	3	3000	704.8	19.4
PS3	100	1.49	0.00	2.23	5.02	3	3000	598.2	2.3
PS3	160	1.52	0.00	2.60	4.70	3	3000	638.8	2.2

Station	Depth (m)	AMX Rate (nM N ₂ d ⁻¹)	DN Rate (nM N ₂ d ⁻¹)	NO ₂ ⁻ (μM)	NH ₄ ⁺ (nM)	Added ¹⁵ NO ₂ ⁻ (μM)	Added ¹⁵ NH ₄ ⁺ (nM)	NH ₄ ⁺ Enrichment	NO ₂ ⁻ Enrichment
PS3	250	1.15	0.00	1.96	5.73	3	3000	524.5	2.5
PS3	800	0.58	0.00	0.00	6.35	3	3000	473.1	*
2	102	0.00	0.00	0.05	N/A	2	N/A	N/A	40.5
2	105	2.21	0.00	0.02	N/A	2	N/A	N/A	132.3
2	120	4.98	0.80	0.25	N/A	2	N/A	N/A	9.0
2	130	0.00	3.38	1.91	N/A	2	N/A	N/A	2.0
2	200	0.89	1.42	2.03	N/A	2	N/A	N/A	2.0
2	395	1.49	0.00	0.24	N/A	2	N/A	N/A	9.3
3	120	6.12	0.00	0.17	N/A	2	N/A	N/A	13.1
3	140	2.62	1.31	1.46	N/A	2	N/A	N/A	2.4
3	170	2.30	0.00	1.72	N/A	2	N/A	N/A	2.2
3	250	5.58	2.99	2.02	N/A	2	N/A	N/A	2.0
9 (7/6)	105	2.23	0.00	0.03	N/A	2	N/A	N/A	76.9
9 (7/6)	115	2.74	0.00	0.06	N/A	2	N/A	N/A	34.0
9 (7/6)	130	4.75	0.00	1.41	N/A	2	N/A	N/A	2.4
9 (7/6)	160	0.00	2.08	1.91	N/A	2	N/A	N/A	2.0
9 (7/6)	168	3.67	0.00	1.52	N/A	2	N/A	N/A	2.3
9 (7/6)	175	0.00	1.40	1.57	N/A	2	N/A	N/A	2.3
9 (7/6)	250	2.82	0.00	1.81	N/A	2	N/A	N/A	2.1
9 (7/6)	375	2.69	0.70	0.00	N/A	2	N/A	N/A	*
9 (7/9)	96	1.53	1.52	0.03	N/A	2	N/A	N/A	69.3
9 (7/9)	100	1.83	2.51	0.03	N/A	2	N/A	N/A	67.9
9 (7/9)	200	19.98	7.03	1.28	N/A	2	N/A	N/A	2.6
9 (7/9)	200	3.41	1.78	1.46	N/A	2	N/A	N/A	2.4
9 (7/9)	300	1.34	0.00	0.99	N/A	2	N/A	N/A	3.0
9 (7/9)	475	1.23	0.00	0.00	N/A	2	N/A	N/A	*
9 (7/9)	730	2.42	0.67	0.00	N/A	2	N/A	N/A	*
11	116	1.92	0.00	0.01	N/A	2	N/A	N/A	151.1
11	320	0.00	0.00	0.00	N/A	2	N/A	N/A	*

Station	Depth (m)	AMX Rate (nM N ₂ d ⁻¹)	DN Rate (nM N ₂ d ⁻¹)	NO ₂ ⁻ (μM)	NH ₄ ⁺ (nM)	Added ¹⁵ NO ₂ ⁻ (μM)	Added ¹⁵ NH ₄ ⁺ (nM)	NH ₄ ⁺ Enrichment	NO ₂ ⁻ Enrichment
12	103	2.64	1.26	0.10	N/A	2	N/A	N/A	20.6
12	120	2.64	0.00	1.51	N/A	2	N/A	N/A	2.3
12	150	8.72	1.91	1.18	N/A	2	N/A	N/A	2.7
12	610	0.99	1.42	0.01	N/A	2	N/A	N/A	399.9
14	125	1.77	0.00	1.20	N/A	2	N/A	N/A	2.7
14	323	0.00	0.34	0.00	N/A	2	N/A	N/A	*
15	135	3.11	1.39	0.07	N/A	2	N/A	N/A	31.7
15	160	1.39	0.00	0.67	N/A	2	N/A	N/A	4.0
18	130	2.01	0.67	0.13	N/A	2	N/A	N/A	16.6
18	160	0.70	0.00	0.00	N/A	2	N/A	N/A	*
18	200	0.00	0.00	0.00	N/A	2	N/A	N/A	*

Supplementary Table S7: NO₂⁻ and NH₄⁺ enrichments due to tracer additions in the SR1805 and FK180624 anammox and denitrification rate samples. The columns labeled NO₂⁻ (μM) and NH₄⁺ (nM) refer to the ambient concentrations. NH₄⁺ concentrations, additions, and enrichments are not reported for FK180624 samples because only ¹⁵NO₂⁻ additions were performed. Asterisks indicate samples where the ambient NO₂⁻ was zero and the enrichment calculation could not be carried out.

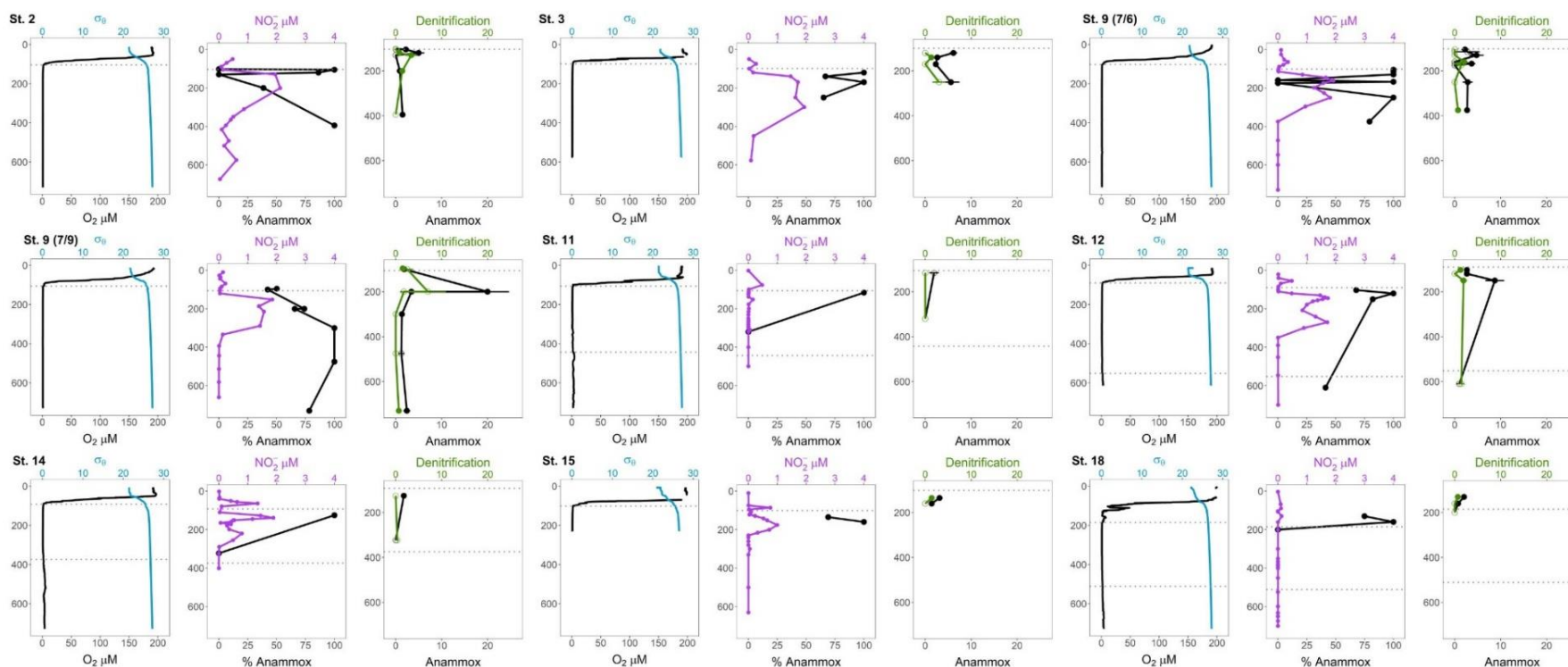


Figure S1: Depth profiles of biogeochemical parameters and N loss rate measurements from cruise FK180624. Measurements were conducted at eight stations along a gradient from waters (St. 2 and 3) similar to OMZ station PS2 (SR1805 cruise) to stations near the border of the OMZ region (St. 15 and 18) (Fig. 1). Station 9 was sampled multiple times, once on 7/6/2018 and once on 7/9/2018. For each station, O_2 (μM) (black) and σ_θ (kg m^{-3}) (cyan) are shown in the first panel, NO_2^- (μM) (purple) and % anammox values (black) are shown in the second panel, and anammox (black) and denitrification rates (green) ($\text{nM N}_2 \text{d}^{-1}$) are shown in the third panel. Grey dotted lines indicate the upper and lower boundaries of the ODZ region at the time of sampling. The bottom boundary was below the edge of the plot in stations 2, 3, and 9 while the CTD did not sample deeply enough to measure this feature for station 15. In the N loss plots, filled circles indicate rates significantly different from zero, while open circles are insignificant rates. Error bars indicate the standard error of the regression.

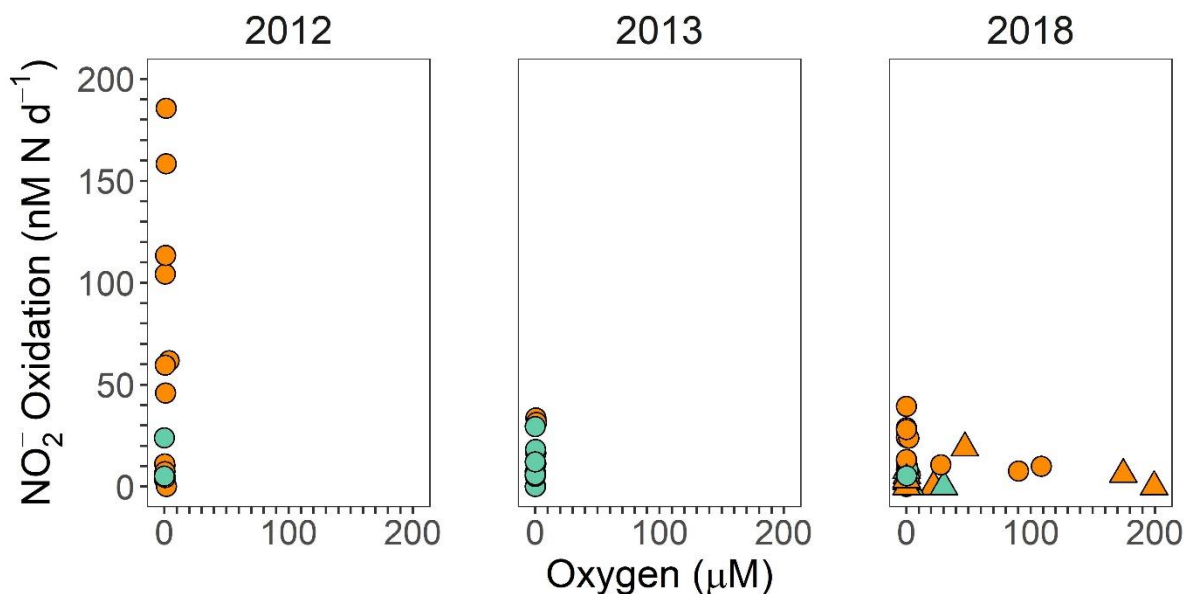


Figure S2: NO₂⁻ oxidation rates (nM N d⁻¹) from the TN278, and NBP1305, and SR1805 cruises vs. O₂ concentration (µM) from the shipboard CTD sensors. O₂ concentrations were normalized across cruises. Rates that are significantly different from zero as assessed via a student T-test (p value < 0.05) are displayed as filled circles, while insignificant rates are displayed as triangles. Rates measured in shallow boundary waters are colored orange while rates from the ODZ core and below are colored teal. 2012 and 2013 data are republished (Babbin et al., 2020).

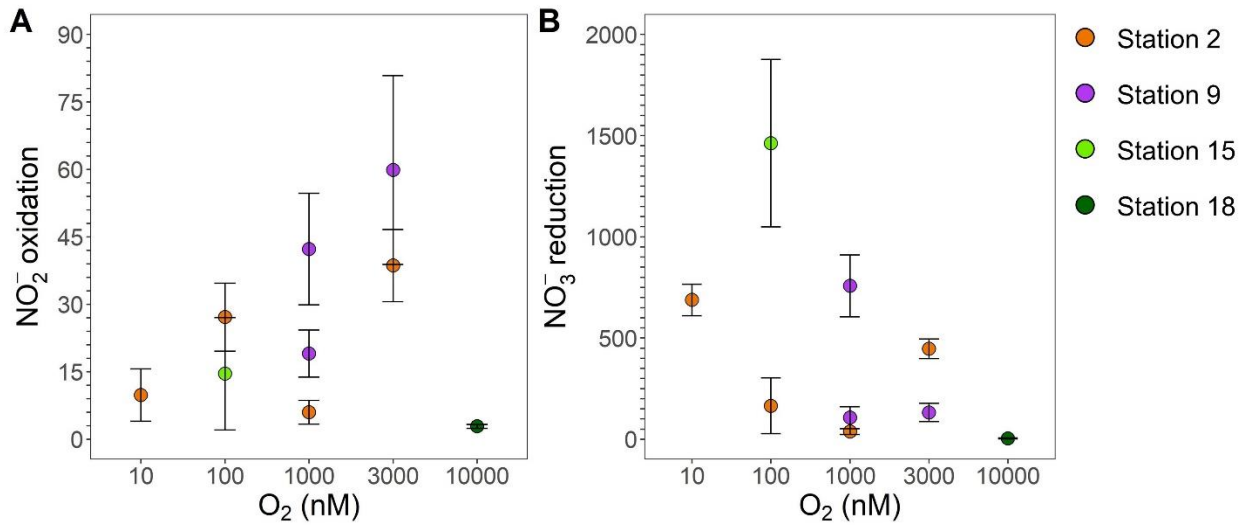


Figure S3: O₂ manipulation experiments from the FK180624 cruise in summer 2018. NO₂⁻ oxidation (**A**) and NO₃⁻ reduction (**B**) (nM N d⁻¹) measured across an O₂ gradient from 10 to 10000 nM. Station 2 (dark orange) is closest to shore and represents a similar region to station PS2 from cruise SR1805 in spring 2018. Station 9 (purple) is an intermediate environment between station 2 and stations 15 (light green) and 18 (dark green) which are farther offshore and represent the boundaries of the OMZ region (Fig. 1). Error bars represent the standard error of the regression.

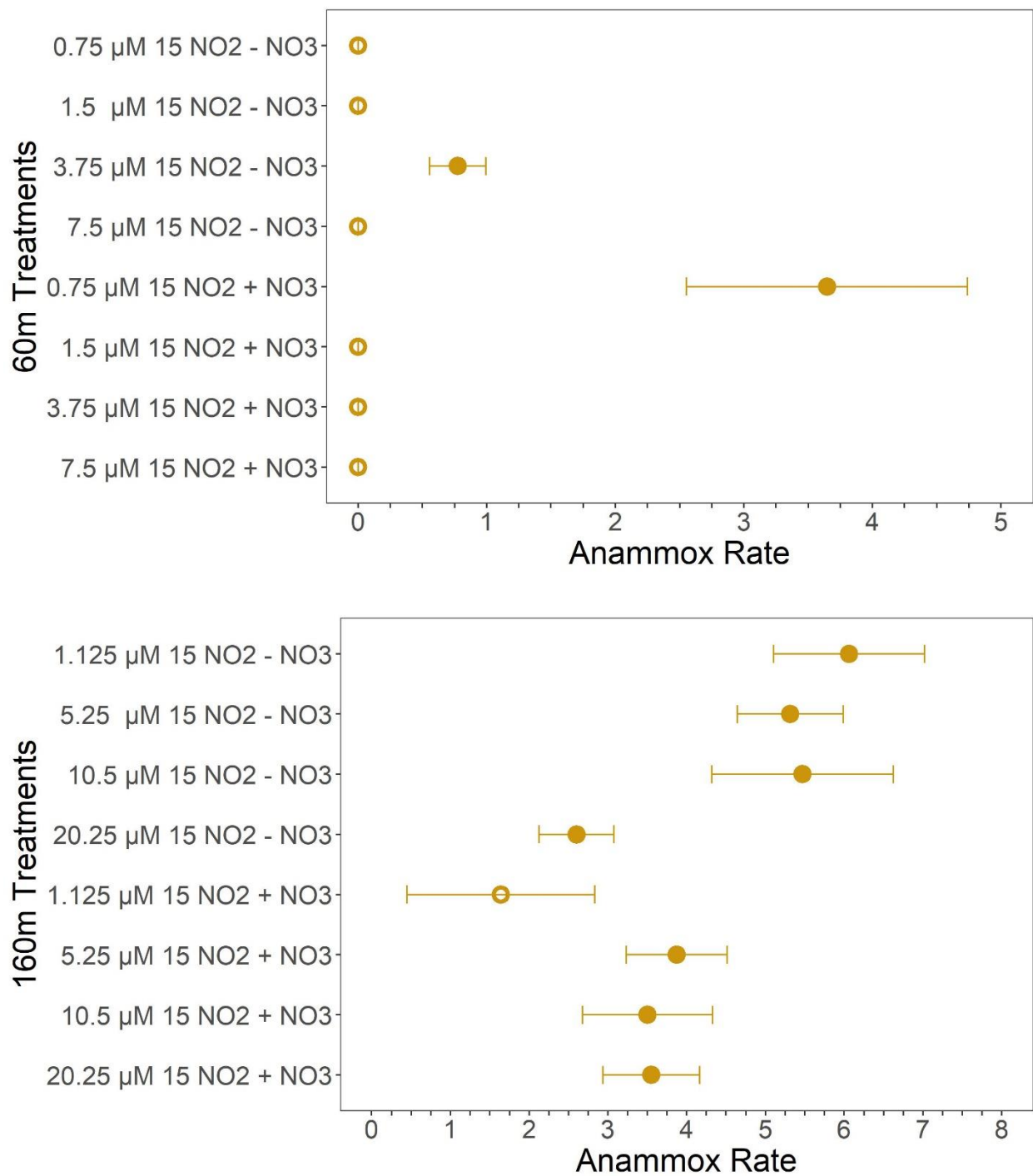


Figure S4: Anammox rates (nM d⁻¹) used to calculate the unexplained NO₂⁻ oxidation rate in Fig. 5. Filled circles indicate significant rates while open circles indicate rates that are not significantly different from zero. Error bars are the standard error of the regression.

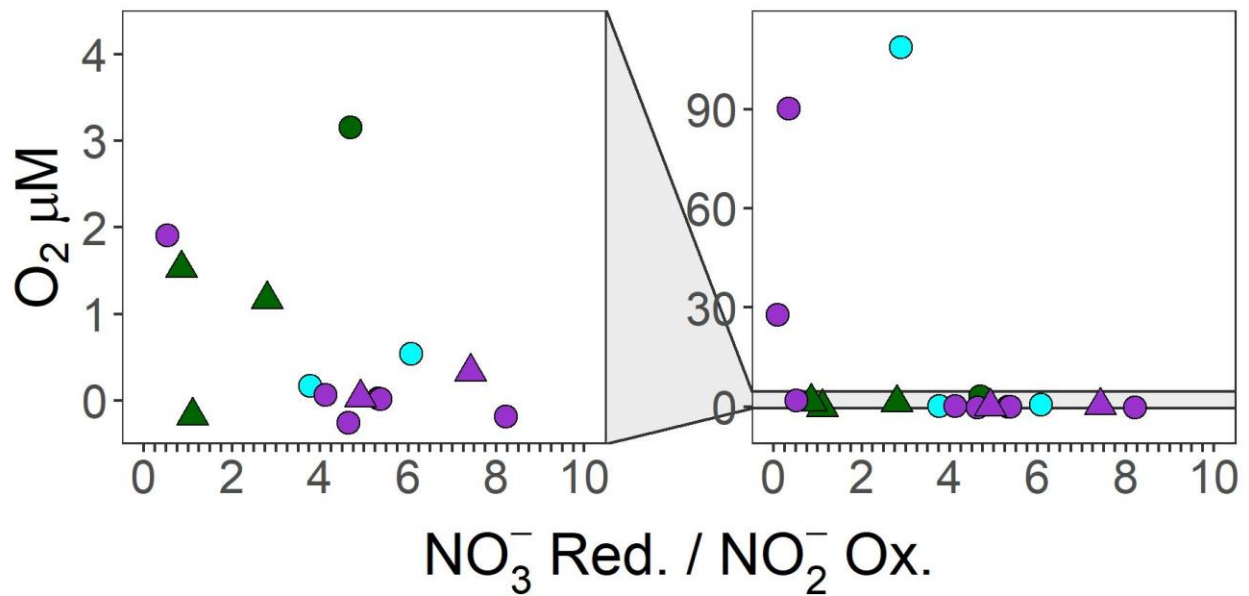


Figure S5: Oxygen does not differentially regulate NO_3^- reduction and NO_2^- oxidation. O_2 (μM) plotted against the ratio between the NO_3^- reduction rate and NO_2^- oxidation rate for all SR1805 depths with significant NO_2^- oxidation rates. Triangles indicate samples from the ODZ core or below ($\sigma_\theta > 26.4$) and circles depict samples from shallow boundary waters ($\sigma_\theta < 26.4$). Colors indicate station: purple (PS3), green (PS2), cyan (PS1).

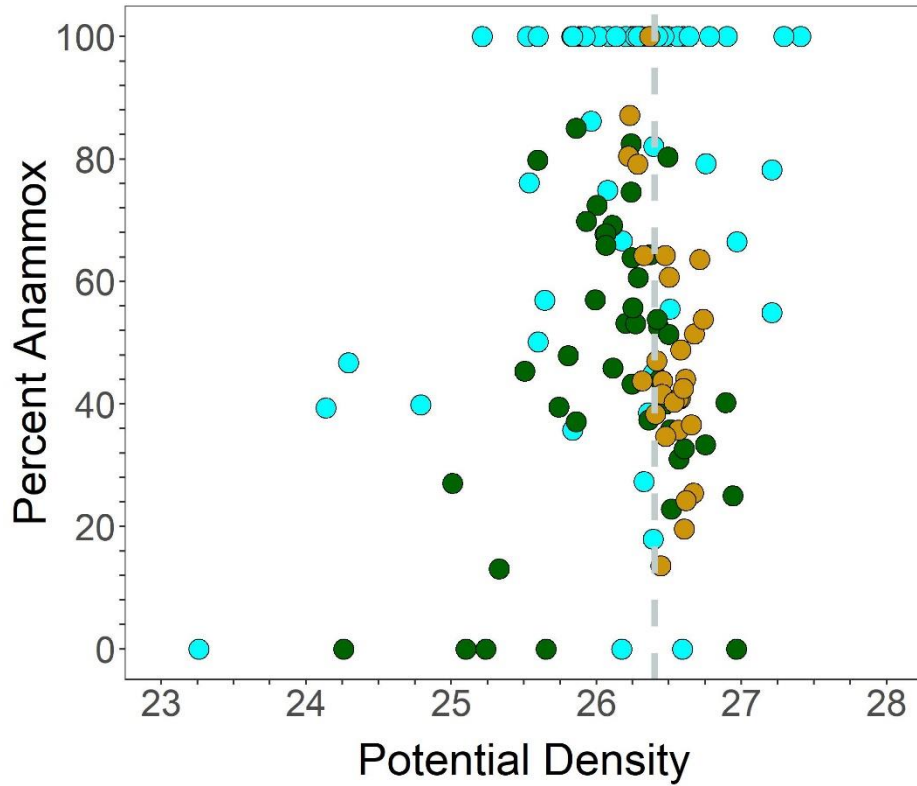


Figure S6: Percent anammox plotted against potential density for all cruises. Data from TN278 (ETNP 2012) is shown in green, NBP1305 (ETSP 2013) is shown in gold, and both 2018 cruises (SR1805 and FK180624) in cyan. The grey dashed line differentiates shallow boundary waters ($\sigma_\theta < 26.4$) and waters from the ODZ core and below ($\sigma_\theta > 26.4$).

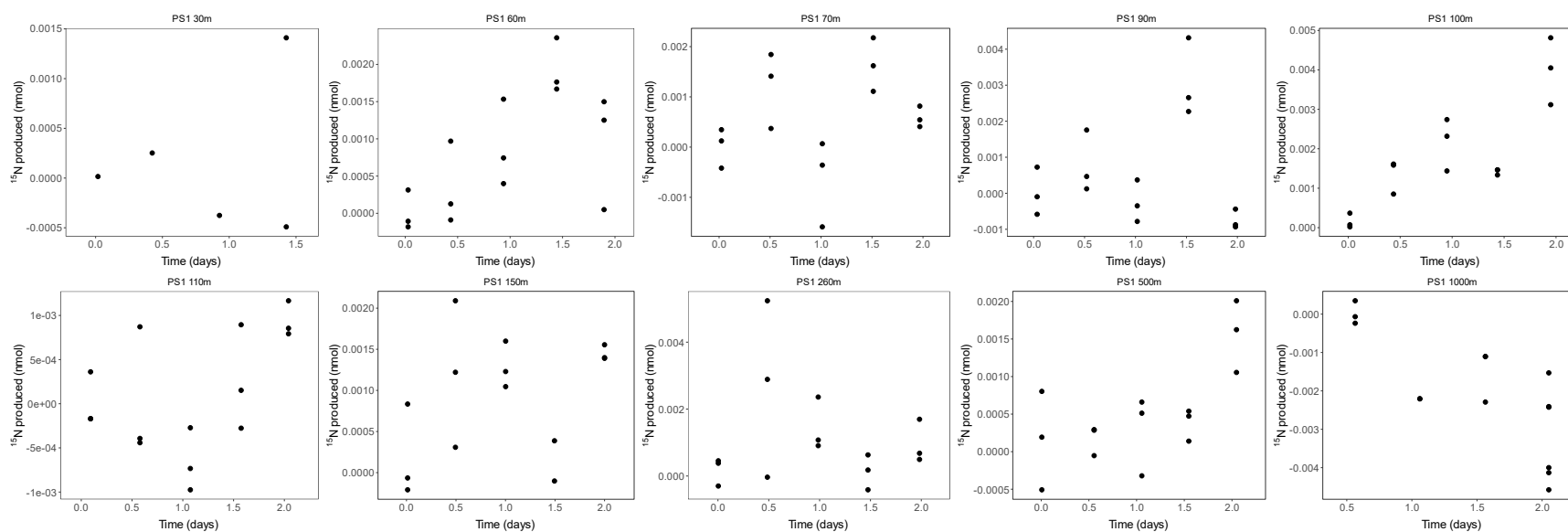


Figure S7: Time-courses from SR1805 NO_2^- oxidation depth profile experiments. All depths from station PS1 are shown. Top row, from left to right: 30, 60, 70, 90, 100 m, bottom row from left to right: 110, 150, 260, 500, 1000m. The y-axis depicts ^{15}N produced in nmol N while the x axis shows time in days. Complete time-courses are shown. For each depth, the best regression from $t_0 - t_2$, $t_0 - t_3$, or $t_0 - t_4$, was used to calculate the final rate plotted in Fig. 2.

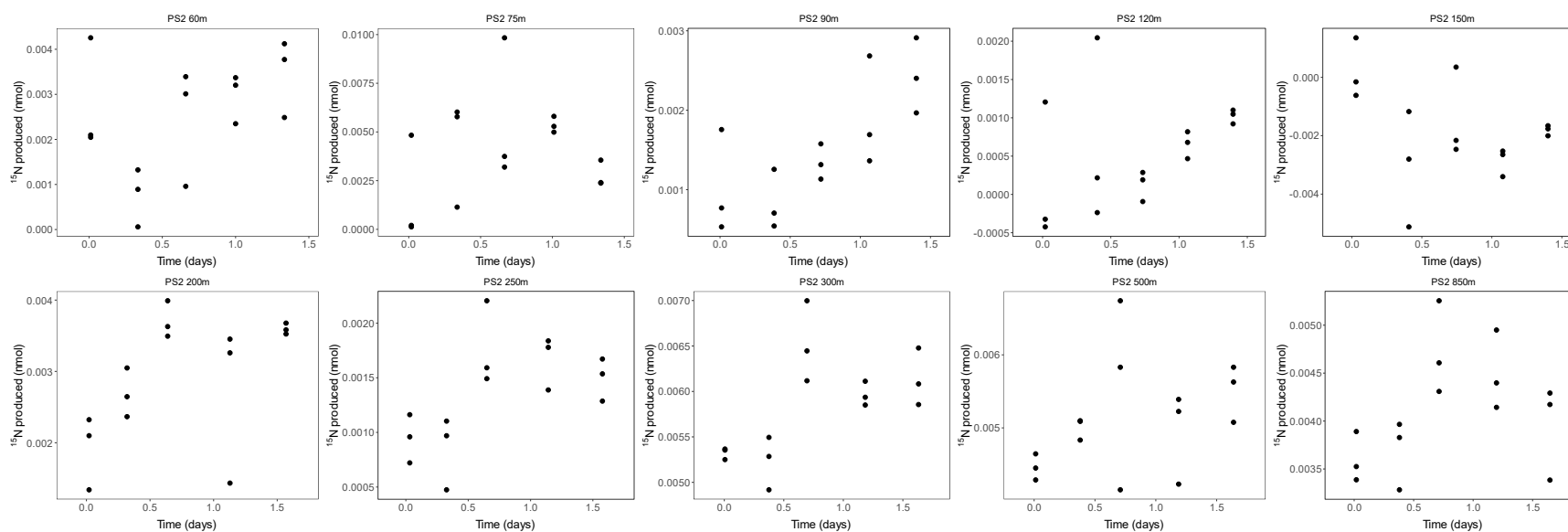


Figure S8: Time-courses from SR1805 NO_2^- oxidation depth profile experiments. All depths from station PS2 are shown. Top row from left to right: 60, 75, 90, 120, 150 m, bottom row from left to right: 200, 250, 300, 500, 850m. The y-axis depicts ^{15}N produced in nmol N while the x axis shows time in days. Complete time-courses are shown. For each depth, the best regression from $t_0 - t_2$, $t_0 - t_3$, or $t_0 - t_4$, was used to calculate the final rate plotted in Fig. 2.

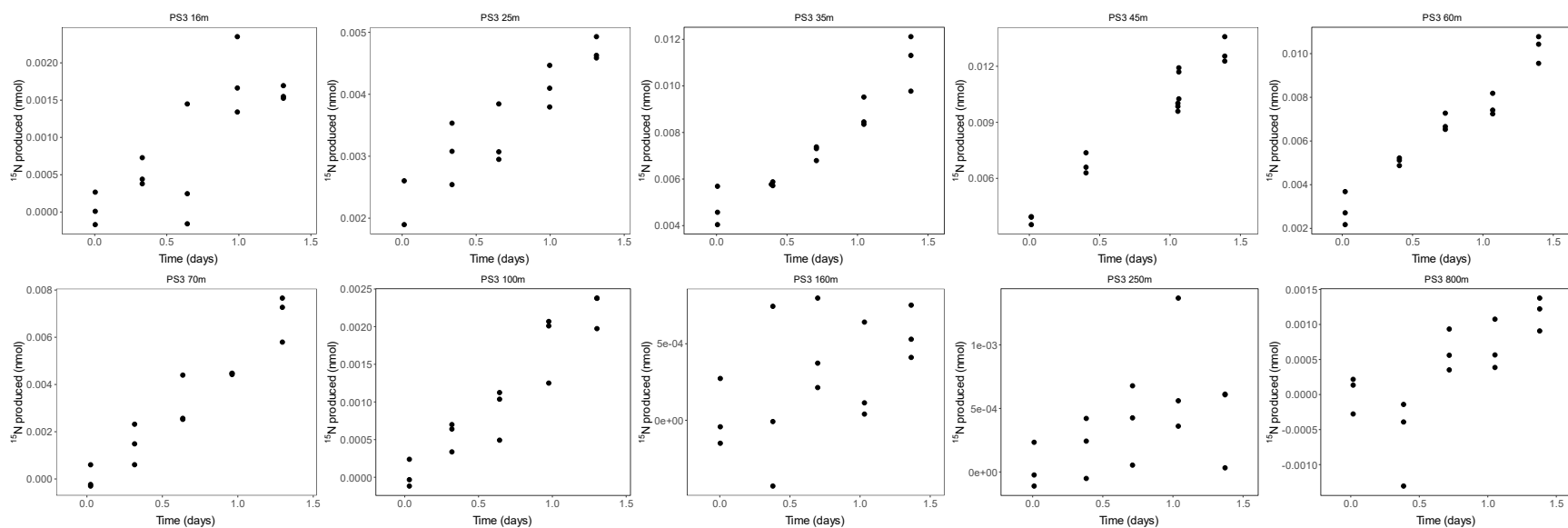


Figure S9: Time-courses from SR1805 NO_2^- oxidation depth profile experiments. All depths from station PS3 are shown. Top row from left to right: 16, 25, 35, 45, 60, bottom row from left to right: 70, 100, 160, 250, 800m. The y-axis depicts ^{15}N produced in nmol N while the x axis shows time in days. Complete time-courses are shown. For each depth, the best regression from $t_0 - t_2$, $t_0 - t_3$, or $t_0 - t_4$, was used to calculate the final rate plotted in Fig. 2.