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1	Hyperspectral remote sensing of cyanobacterial pigments
2	as indicators for cell populations and toxins in eutrophic
3	lakes
4	
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1 Abstract

2 The growth of mass populations of toxin-producing cyanobacteria is a serious concern for the 3 ecological status of inland waterbodies and for human and animal health. In this study we 4 examine the performance of four semi-analytical algorithms for the retrieval of chlorophyll a 5 (Chl *a*) and phycocyanin (C-PC) from data acquired by the Compact Airborne Spectrographic 6 Imager-2 (CASI-2) and the Airborne Imaging Spectrometer for Applications (AISA) Eagle 7 Sensor. The retrieval accuracies are compared to those returned by semi-empirical band-ratio 8 algorithms optimised for these datasets. The best-performing algorithm for the retrieval of Chl *a* was a semi-empirical band-ratio model ($R^2 = 0.832$; RMSE = 29.8%). The best 9 performing model for the retrieval of C-PC was a semi-analytical nested band ratio model (R² 10 = 0.984; RMSE = 3.98 mg m^{-3}). The concentrations of C-PC retrieved using the semi-11 analytical model were also correlated with those of cyanobacterial cells ($R^2 = 0.380$) and of 12 the particulate and total (particulate plus dissolved) pools of microcystins ($R^2 = 0.858$ and 13 14 0.896 respectively). Importantly, both the semi-empirical and semi-analytical algorithms 15 were able to retrieve the concentration of C-PC at cyanobacterial cell concentrations below 16 current warning thresholds for cyanobacteria in waterbodies. This demonstrates the potential 17 of remote sensing to contribute to early-warning detection and monitoring of cyanobacterial 18 blooms for human health protection, at regional and global scales.

19

Key words: Cyanobacteria; human health; lakes; microcystin; imaging spectrometry; risk
assessment

1 Introduction

2 1.1 Mass populations of toxic cyanobacteria

3 Cyanobacteria (blue-green algae) are natural and cosmopolitan inhabitants of fresh, brackish 4 and marine waters. They are particularly well adapted for growth in nutrient-enriched lakes 5 and other slow-flowing inland waters and often form mass populations (as blooms, scums or 6 biofilms) during summer and autumn in temperate latitudes. These mass populations are a 7 serious concern because they can have profound and far-reaching adverse environmental and 8 economic impacts on natural and controlled waterbodies (Pearson et al., 2001; Havens, 9 2008). Cyanobacterial populations can also pose significant risks to animal and human health 10 because several species can produce potent toxins (cyanotoxins). These toxins constitute 11 some of the most hazardous of all waterborne biological substances and include agents with 12 (examples in parentheses): neurotoxic (anatoxin-a, anatoxin-a[S], beta-methylamino alanine, 13 saxitoxins); hepatotoxic and tumour-promoting (microcystins, nodularins); cytotoxic and 14 genotoxic (cylindrospermopsins), and endotoxic (lipopolysaccharides) properties (Codd et 15 al., 2005b; Metcalf et al., 2008). The microcystins (MC), of which over 90 structural variants 16 are known, are among the most common and potent of the cyanotoxins.

17

18 Blooms of toxigenic cyanobacteria in waterbodies used for drinking water, aquaculture, crop 19 irrigation and recreation are increasing in abundance and spread due to eutrophication. 20 Human exposure to bloom-forming cyanobacteria and their toxins most commonly occurs via 21 drinking water or incidental ingestion, dermal contact and inhalation (Codd et al., 2005b). 22 However, there is also evidence that cyanotoxins can be transferred to crops intended for 23 human consumption via spray-irrigation with contaminated water (Codd et al., 1999; Tyler et 24 al., 2009). Cyanotoxins have been linked with numerous incidences of ill health in humans, 25 ranging from cases of mild skin irritation and gastrointestinal illnesses to acute (and

occasionally fatal) poisonings (Pilotto et al., 1997; Carmichael et al., 2001). These health
 risks may be further exacerbated if incidences of blooms continue to increase under a
 warming climate (Johnk et al., 2008; Paerl & Huisman, 2008).

4

5 Cyanobacterial abundance is a useful indicator of inland and coastal water quality and is one 6 of the biological quality metrics being considered for the assessment of lake ecological status 7 under the European Union's Water Framework Directive (EU WFD) (2000/60/EC). The 8 monitoring of cyanobacteria and cyanotoxins is also necessary so that timely warnings can be 9 provided to safeguard animal and human health. Cyanotoxin concentrations are rarely 10 routinely monitored in waterbodies as the analyses are costly, and require specialist 11 laboratory facilities and technical expertise. Consequently, existing monitoring programmes 12 tend to focus on cyanobacterial cell numbers (or biovolume) rather than cyanotoxin 13 identification and quantification. The resource costs involved in both sampling lakes and 14 analysing cyanobacterial densities, limits agencies across Europe to generally only monitor a 15 single location, from a small proportion of lakes at a monthly frequency at best. 16 Consequently, conventional monitoring programmes are only aimed at providing a general 17 assessment of ecological quality at these lakes and are insufficient to provide timely warnings 18 of bloom development across whole lake areas or across large geographical regions. 19

There is a clear need to develop techniques that can complement and extend current approaches to monitoring and managing cyanobacteria and their toxins in inland, transitional and coastal waters. Indeed, this has recently been acknowledged by a number of international bodies (WHO, 2003, 2004; Codd et al., 2005a). Techniques including flow cytometry and spectrofluorometry (with specificity for diagnostic cyanobacterial pigments) offer possibilities for real-time monitoring of cyanobacteria in the field. However, such techniques

can be less effective where monitoring needs to be undertaken routinely across numerous
 lakes at a regional or national scale. Moreover, as cyanobacterial bloom and scum
 distribution can change rapidly in space and time, e.g. according to wind speed and direction,
 observations at fixed stations or along set transects may be unrepresentative of cyanobacterial
 distribution and abundance throughout the entire waterbody (Rantajarvi et al., 1998; Kutser,
 2004).

7

8 In view of this, there is considerable interest in developing remote sensing-based techniques 9 to detect and map cyanobacterial blooms from space. Recent advances in ocean colour 10 remote sensing have demonstrated that it is possible to develop bespoke inversion algorithms 11 for mapping different phytoplankton functional types (PFTs) and size classes (Nair et al., 12 2008) and to recognise important bloom-forming species including coccolithophores (Smyth 13 et al., 2002), diatoms (Sathyendranath et al., 2004) red tide dinoflagellates (Tomlinson et al., 14 2009) and toxigenic cyanobacteria (Ramos et al., 2005; Westberry et al., 2005). Similarly, in 15 inland waters there is a small but growing body of evidence to suggest that the identification 16 and quantification of cyanobacteria is possible from measurements of remote-sensing-17 reflectance $(R_{rs}(0^+,\lambda))$ because of their unique bio-optical traits (Hunter et al., 2008a; Kutser et al., 2006; Reinart & Kutser, 2006). A comprehensive review of recent progress in this 18 19 field can be found in Kutser (2009).

20

21 1.2 Remote sensing of cyanobacterial blooms

22 Chl *a* concentration is used as a standard measure of phytoplankton biomass in fresh and

23 marine waters and numerous inversion algorithms for the retrieval of Chl *a* from

24 measurements of $R_{rs}(0^+,\lambda)$ have been developed for both Case 1 waters (those waters whose

25 optical properties are determined primarily by phytoplankton and covarying coloured

1 dissolved organic matter and detrital material (CDOM)), and Case 2 waters (whose optical 2 properties are significantly influenced by other constituents such as mineral particles and 3 CDOM that do not covary with phytoplankton concentration) (Gons, 1999; Tyler et al., 4 2006). Because Chl *a* is almost ubiquitous across phytoplankton, the retrieval of Chl *a* alone 5 does not provide any information on the presence and abundance of specific taxa. However, 6 the development of new inversion algorithms for the retrieval of the cyanobacterial biomarker 7 pigment C-phycocyanin (C-PC) from $R_{rs}(0^+,\lambda)$ offers a route to detect and quantify 8 cyanobacterial biomass in inland and coastal waters. To date, three different types of 9 inversion algorithms have been developed for the retrieval of C-PC over turbid inland waters. 10 These include the baseline algorithm proposed by Dekker (1993), the band-ratio algorithms 11 for C-PC retrieval of Schalles et al. (2000) and Hunter et al. (2008b; 2009), and the semi-12 empirical nested band-ratio of Simis et al. (2005) (hereafter referred to as the SIMIS 13 algorithm).

14

15 The SIMIS semi-analytical nested band-ratio algorithm retrieves the value of C-PC 16 absorption coefficient at 620 nm. It includes a correction for absorption by Chl a at the 620 17 nm C-PC absorption maximum. The algorithm is based on an earlier model for Chl a 18 retrieval developed by Gons (1999) for MERIS (Medium Resolution Imaging Spectrometer) 19 onboard the European Space Agency's Envisat (hereafter referred to as the GONS 20 algorithm). These algorithms exploit the relationship between subsurface irradiance 21 reflectance and the absorption and backscattering properties of optically-active constituents in 22 the water column as defined by Gordon et al. (1975):

23

24

$$R(0-,\lambda) = f \frac{b_b(\lambda)}{a(\lambda) + b_b(\lambda)}$$
(1)

2 Where f is an empirically determined scale factor dependent on the ambient light field and 3 $a(\lambda)$ and $b_b(\lambda)$ are the coefficients for total absorption and backscattering respectively. If f 4 and b_b are assumed to be spectrally-independent, and b_b is known for a particular wavelength, 5 then absorption by water and its main constituents can be solved from the ratio $R(\lambda_2):R(\lambda_1)$ 6 and a known value for *a* at wavelength λ_2 :

8

$$a(\lambda_1) = \left\{ [a(\lambda_2) + b_b] \times \left[\frac{R(\lambda_2)}{R(\lambda_1)} \right] \right\} - b_b$$

9

10 The GONS algorithm uses the ratio of reflectance for $\lambda_1 = 665$ and $\lambda_2 = 709$ nm to estimate 11 the Chl *a* absorption coefficient at 665 nm. It assumes that $a(\lambda_1)$ can be attributed to 12 absorption by water $[a_w(\lambda_1)]$ and phytoplankton pigments $[a_{ph}(\lambda_1)]$, and that $a(\lambda_2)$ can be 13 attributed entirely to absorption by water $[a_w(\lambda_2)]$. This ignores the possible contributions of 14 non-phytoplankton particulates (inorganic and organic suspended particulate matter) and 15 CDOM to $a(\lambda_1)$ and $a(\lambda_2)$. It also assumes that absorption by phytoplankton pigments is 16 negligible at $a(\lambda_2)$. The empirical correction factor γ is introduced to relate absorption at λ_1 to that measured at the same wavelength. The following relationship for the retrieval of Chl a 17 absorption at 665 nm $[a_{chl}(665)]$ is then obtained: 18

19

20
$$a_{chl}(665) = \left(\left\{\left[a_w(709) + b_b\right] \times \left[\frac{R(709)}{R(665)}\right]\right\} - b_b - a_w(665)\right) \times ?^{-1}$$
21 (3)

21

22

(2)

1 The value of b_b is estimated according to Gons (1999; 2005) by using a wavelength further 2 into the NIR and inverting Eq. (1) using an estimate of f based upon an empirical relation 3 incorporating the average cosine of downward irradiance: 4 $b_b(779) = \frac{1.61 \times R(779)}{0.082 - 0.6 \times R(779)}$ 5 (4) 6 7 The concentration of Chl *a* is then returned by dividing $a_{chl}(665)$ by the specific absorption coefficient of Chl *a* at 665 nm $(a_{chl}^*(665))$ as shown below: 8 9 Chl $a = a_{chl}(665)/a_{chl}^{*}(665)$ 10 (5) 11 12 The above algorithm was modified in Simis et al. (2005) for the retrieval of C-PC through the introduction of the ratio of reflectance at $\lambda_1 = 620$ and $\lambda_2 = 709$ nm and an empirically 13 14 derived factor δ for the correction of a(620) that mimics the use of the constant γ used in Eq. 3. To account for co-absorption by Chl a and C-PC at 620 nm it is necessary firstly to 15 16 estimate absorption by Chl a at 620 nm [$a_{chl}(620)$] and then subtract this from a(620) to yield 17 the absorption by C-PC [$a_{CPC}(620)$]. This is achieved by the introduction of the conversion 18 factor ε that relates *in vivo* absorption by Chl *a* at 665 nm to that at 620 nm. Subsequently, 19 $a_{\rm CPC}(620)$ can be solved from: 20

21
$$a_{CPC} = \left(\left\{\left[a_w(709) + b_b\right] \times \left[\frac{R(709)}{R(620)}\right]\right\} - b_b - a_w(620)\right) \times ?^{-1} - [? \times a_{chl}(665)]$$
(6)

1 by substitution of $a_w(620) = 0.281$, $a_w(665) = 0.401$ and $a_w(709) = 0.727$ from Buiteveld et al. 2 (1994) and $\delta = 0.84$, $\gamma = 0.68$ and $\varepsilon = 0.24$ from Simis et al. (2005). The concentration of C-3 PC is retrieved by dividing by the specific absorption coefficient for C-PC at 620 nm: 4 $CPC = a_{CPC} (620) / a_{CPC}^{*} (620)$ 5 (7) 6 7 The inversion algorithm developed by Simis et al. (2005) has recently been tested using 8 extensive in-situ radiometric measurements at a series of lakes and reservoirs in the 9 Netherlands, Spain and the United States (c.f., Randolph et al., 2008; Ruiz-Verdu et al., 2008; 10 Simis et al., 2007) and was found to outperform other algorithms for C-PC retrieval. The 11 inversion algorithm was also recently successfully used to retrieve C-PC estimates from 12 MERIS data {Guanter et al. 2010}. The wider testing of this algorithm for the retrieval of C-PC from $R_{rs}(0^+,\lambda)$ spectra measured by airborne and spaceborne sensors is necessary because 13 14 the incursion of errors due to atmospheric and adjacency effects can have a significant effect 15 on the estimation of pigment concentrations from remotely sensed data (Alikas & Reinart, 16 2008). 17 18 In addition to the bespoke algorithm for the retrieval of C-PC of Simis et al. (2005), a 19 medium-independent model to retrieve pigment concentrations has been developed by Dall'Olmo et al. (2003). This model uses the value of $R_{rs}(0^+,\lambda)$ in three bands to estimate the 20 21 pigment concentrations: 22 Pigment concentration $\propto [R_{rs}^{-1}(\lambda_1) - R_{rs}^{-1}(\lambda_2)] \times R_{rs}(\lambda_3)$ 23 (8) 24

1 The specific wavelengths used in the model must be optimised for the pigment of interest 2 through empirical iteration. Dall'Olmo et al. (2003) developed the algorithm originally for 3 the retrieval of Chl *a* in terrestrial vegetation and turbid inland waters. The theory behind the 4 model is that the reciprocal of reflectance at wavelength λ_1 should be maximally sensitive to 5 absorption by the pigment of interest. Dall'Olmo and Gitelson (2005) suggest a wavelength 6 between 660 to 690 nm for the retrieval of Chl a. However, as reflectance in this spectral 7 region is also affected to varying degrees by absorption by mineral particles, organic detritus 8 and CDOM, a second wavelength, λ_2 , that is minimally sensitive to absorption by the 9 pigment of interest, and is close to λ_1 , is needed to correct for co-absorption by other optically-active constituents. It has been shown that λ_2 should occur between 710 to 730 nm 10 11 for the retrieval of Chl a (Dall'Olmo & Gitelson, 2006; Gitelson et al., 2006). 12 13 To account for potential differences in backscattering, potentially caused by non-14 phytoplankton particulates, a third wavelength λ_3 is introduced into the model at which 15 absorption is dominated by water (i.e. λ_3 should be minimally sensitive to absorption by pigments, mineral particles, organic detritus and CDOM). Dall'Olmo and Gitelson (2005) 16 located λ_3 between 740 and 760 nm. 17 18 19 The semi-analytical algorithm derived by Dall'Olmo (2003) has been well-validated for the 20 retrieval of Chl a using empirical datasets and radiative transfer modelling (Dall'Olmo & 21 Gitelson, 2005; Dall'Olmo & Gitelson, 2006; Gitelson et al., 2008). Hunter et al. (2008a)

22 previously adapted this algorithm to estimate C-PC concentrations in experimental cultures of

23 cyanobacteria. This was achieved by setting $\lambda_1 = 630$, $\lambda_2 = 660$ and $\lambda_3 = 750$ nm after

24 iterative tuning of the algorithm. However, the ability of the algorithm to estimate C-PC in

25 natural inland waters has yet to be examined.

2 In the present study, we test the performance of the semi-analytical algorithms of Gons 3 (1999), Simis et al. (2005) and Dall'Olmo et al (2003) for the retrieval of Chl a and C-PC in 4 inland waters using imagery from the Compact Airborne Spectrographic Imager-2 (CASI-2) 5 and the hyperspectral Airborne Imaging Spectrometer for Applications (AISA) Eagle and 6 Hawk sensors. We compare their performance to that of several semi-empirical algorithms 7 optimised for our specific dataset. We also analyse the relationships between the estimated 8 C-PC concentrations and the measured concentrations of cyanobacterial cells and toxins 9 (microcystins). The potential utility of remote sensing as a means of monitoring 10 cyanobacterial blooms in inland waters is discussed and some directions for future research 11 are outlined.

- 12
- 13 **2. Methods**

14 2.1 Study sites

15 The study was undertaken at two shallow, eutrophic lakes in the UK (Fig. 1). Loch Leven is the largest lake (13.3 km²) in lowland Scotland (56°12'N, 3°22'W) with a mean depth of 3.9 16 17 m. Loch Leven water quality has been adversely affected by nutrients from domestic, 18 agricultural and industrial sources and, while there is some evidence for recovery (Carvalho 19 & Kirika, 2003; Ferguson et al., 2008), cyanobacterial blooms continue to occur during 20 summer months. These blooms are typically dominated by genera such as Anabaena, 21 Microcystis, Snowella and Woronichinia. Esthwaite Water is a small lake in the English 22 Lake District (54°21'N, 3°0'W) with a mean depth of 6.4 m. The lake receives nutrients from 23 a local sewage treatment works and a fish farm. Cyanobacterial blooms dominated by genera 24 such as Anabaena, Aphanizomenon, Microcystis, Planktothrix and Woronichinia occur

annually during the summer. Table 1 provides a summary of the physical and
 physicochemical characteristics of the two lakes.

3

4 2.2 Water sampling and analysis

5 2.2.1. Lake sampling for algorithm development and testing

6 Water samples were collected from both lakes concurrent with the airborne remote sensing 7 flights to provide ground reference data for algorithm development and testing. Five litre 8 samples were collected from the surface using a large bucket at 5 stations across each lake. 9 The 5L water samples were thoroughly mixed to uniformly disperse any buoyant 10 cyanobacterial cells, and then split into replicate aliquots for phytoplankton cell counts and 11 pigment and microcystin analysis. The samples intended for phytoplankton counts were 12 immediately fixed with Lugol's Iodine solution (1% v/v). Samples for pigment analysis were 13 stored on ice in the dark for <6 h before they were filtered by passing the sample through 14 replicate Whatman GF/F filters under vacuum pressure and freezing the filters at -80°C until 15 further analysis could be performed. Samples for microcystin analysis were stored in the 16 dark at 4°C and processed within 48 h.

17

18 2.2.2. Phytoplankton cell counts

The concentration of cyanobacterial cells was determined in all the samples collected during the lake cruises. Cells were counted under an inverted microscope according to standard procedures (Brierley et al., 2007) with taxonomy following John et al. (2003). Biovolumes were calculated for the Loch Leven samples using appropriate geometric models for each taxon (Hillebrand et al., 1999). In addition, cell counts from routine fortnightly surveys were provided for both lakes by the Centre for Ecology & Hydrology.

1 2.2.3. Pigment analysis

2 Chl a was extracted by manual grinding of filter discs for 1 min and soaking in 90% acetone 3 in the dark for 20 h at -20°C {Hunter et al., 2008}. The extracts were clarified by 4 centrifugation (10 min, 5000 x g) and the concentration of Chl a was determined by 5 spectrophotometry using the equations of Lorenzen (1967). C-PC was extracted into a 50 mmol L^{-1} sodium phosphate buffer (pH 7.0) using probe ultrasonication (1 min in 15 s bursts) 6 7 and 9 consecutive freeze-thaw cycles (-80 to 4°C). This is a modification of the method 8 presented in Sarada et al. (1999) and was used here to increase extraction efficiency. The 9 extracts were clarified by centrifugation (10 min, 5000 x g) and C-PC was determined by 10 spectrophotometry using the equations of Bennett and Bogorad (1973). 11 12 2.2.4. Microcystin (MC) analysis 13 MCs were analysed by enzyme-linked immunosorbent assay (ELISA) according to Metcalf et 14 al. (2000). The procedure included polyclonal antibodies raised against the commonly-15 occurring MC structural variant, MC-LR, which show similar cross-reactivities with all other 16 MC variants tested, e.g. MC-LY, -LA and -LW. The ELISA was calibrated with purified 17 MC-LR and MC concentrations expressed as microcystin-LR equivalents. Environmental 18 samples were analysed by ELISA either directly to determine extracellular (dissolved) MCs 19 only, or after boiling in a waterbath at 100°C for one minute to release MCs from the 20 cyanobacterial cells (Metcalf & Codd, 2000). In the latter case, the MCs determined 21 comprised the soluble (extracellular) plus particulate (intracellular) toxin pools (Metcalf and 22 Codd, 2000), and are termed "total microcystin". Analysis of all environmental samples for 23 MCs was performed in triplicate.

24

25 2.3 Radiometric measurements

Radiometric measurements were made concurrent to the airborne overflights so that the
 accuracy of the atmospheric correction applied to the CASI-2 and AISA Eagle data could be
 assessed. Shipboard measurements of downwelling (*E*_d(0⁺,λ)) and water-leaving radiance
 (*L*_w(0+,λ)) just above the water surface were collected at the sampling stations using an ASD
 FieldSpec HandHeld VNIR (Analytical Spectral Devices Inc., Boulder, CO) spectrometer.
 The ASD FieldSpec instrument has a spectral interval of 1.4 nm and a full-width half
 maximum of 3 nm over the sampling range of 348-1074 nm.

8

9 The instrument was mounted on a telescopic pole that was extended over the side of the 10 vessel at 0.5 m above the water surface to ensure that measurements were not affected by 11 shadows cast by the boat. The measurement geometry followed Mobley (1999) with spectra 12 acquired at an approximate viewing zenith angle = 42° and azimuth angle = 135° over solar 13 zenith angles = $42.1-58.1^{\circ}$. This measurement geometry was used to minimise the 14 contribution of sky radiance $(L_{sky}(0+,\lambda))$ to $L_w(0+,\lambda)$ and because the spectra were collected 15 under clear skies and over turbid water the skylight effect would have been largely limited to 16 the blue wavelengths that were not utilised this study. We therefore did not invoke a 17 correction for the skylight term. $E_d(0^+,\lambda)$ was obtained from a calibrated Spectralon reflectance panel. $R_{rs}(0^+,\lambda)$ was calculated as the ratio of $L_w(0^+,\lambda)$ to $E_d(0^+,\lambda)$. 18

19

20 2.4 Remote sensing

21 2.4.1. Data acquisition

The AISA Eagle and Hawk sensors were flown in tandem over Esthwaite Water and Loch Leven on the 13 and 26 April, 2007. CASI-2 was flown over Loch Leven on 22 August 2007. The overflights occurred between 10:00 and 14:00 h Greenwich Mean Time under clear skies. The AISA Eagle is a 12-bit pushbroom sensor with a 1024 pixel swath width (field of view (FOV) = 39.7°) across 126 spectral pixels over the range 400-970 nm and a 2.9
nm spectral resolution. The AISA Hawk is a 14-bit pushbroom hyperspectral sensor with a
320 pixel swath width (FOV = 24.0°) across 237 spectral pixels over the range 970-2450 nm
and an 8 nm spectral resolution. CASI-2 is a fully-customisable 12-bit pushbroom sensor.
CASI-2 was operated in Spatial Mode providing a 512 pixel swath width (FOV = 54.4°) and
12 spectral pixels. The channel configuration used for CASI-2 was optimised for the imaging
of Case 2 waters as described by Hunter et al. (2008b).

8

9 The sensors were flown on a Dornier 228-101 fixed wing airplane at an altitude between 10 2900-3200 m (above ground level) and at a ground speed of 135 knots (250 kph). The 11 resulting AISA Eagle and Hawk imagery had spatial resolutions (at nadir) of between 2.0-2.1 12 m and 3.6-3.9 m respectively and the CASI-2 imagery had a spatial resolution of 13 approximately 6 m. The imagery was flown in a south-to-north direction over Esthwaite and 14 a southeast-to-northwest direction over Loch Leven to minimise sun glint effects. Four 15 separate flightlines with an overlap of approximately 30% were flown over Loch Leven and 16 mosaicked together to achieve full lake coverage. The aircraft turn time between flightlines 17 was typically less than 5 min.

18

19 2.4.2. Geometric and atmospheric correction

The airborne imagery was geometrically corrected for pitch, roll and yaw effects using the *azgcorr* programme and geolocated using navigation data obtained by a differentiation global position system. The individual AISA Eagle and Hawk bandsets were mosaicked together using coordinate based layer stacking. The AISA Eagle and Hawk at-sensor radiances were atmospherically corrected to $R_{rs}(0^+,\lambda)$ using the Fast Line-of-Sight Atmospheric Analysis of Spectral Hypercubes (FLAASH) module in RSI ENVI (v.4.5). FLAASH is a physics-based

atmospheric correction model developed by Spectral Sciences Incorporated (Alder-Golden et
al., 1999) and based upon the widely-used and well-validated MODTRAN4 (MODerate
resolution atmospheric TRANsmission) radiative transfer code. FLAASH has been used
successfully in previous research for the atmospheric correction of hyperspectral data
{Kutser, 2004; Cooley et al. 2002}.

6

7 The inputs into FLAASH comprised the altitude of the sensor, the ground elevation above sea 8 level, the pixel size of the imagery and the flight date and time. The atmospheric model used 9 was the MODTRAN4 Sub-Arctic. The concentration of aerosols was retrieved on a pixel-by-10 pixel basis according to the 2-band (K-T) method of Kaufman et al. (1997) using a rural 11 model and the AISA Hawk channels located at 660 and 2100 nm, and 420 and 800 nm. 12 Water vapour was retrieved pixel-by-pixel using the water absorption feature located at 1135 13 nm. The AISA Eagle spectra were interpolated to a final resolution of 0.5 nm and the 14 accuracy of the atmospheric correction was assessed by checking for quantitative agreement 15 with the $R_{rs}(0^+,\lambda)$ spectra calculated from the in-situ radiometry. The accuracy of the 16 atmospheric correction was assessed on the basis of the root mean square error (RMSE). 17 18 It was not possible to correct the CASI-2 data using FLAASH because the sensor does not 19 possess channels in the near- and mid-infrared needed for aerosol and water vapour retrieval. 20 In addition, because FLAASH only models the atmosphere for a single nadir looking 21 direction, it does not account for the increase in path length that occurs in the cross-track 22 direction for wide FOV sensors such as CASI-2. We therefore used the empirical line 23 technique {Karpouzli and Malthus, 2003} to convert the CASI-2 at-sensor radiances to 24 approximate $R_{rs}(0^+,\lambda)$.

1 2.5 Development and testing of inversion algorithms

2	We tested the semi-analytical algorithms of Gons (1999) and Dall'Olmo et al. (2003) for the
3	retrieval of Chl a and of Simis et al. (2005), and a modified version of Dall'Olmo et al.
4	(2003), for the retrieval of C-PC. The algorithms of Gons (1999) and Simis et al. (2005)
5	estimate the values of $a_{chl}(665)$ and $a_{CPC}(620)$ respectively; the values for $a_{chl}(665)$ and
6	$a_{CPC}(620)$ were converted to pigment concentrations by dividing by the specific absorption
7	coefficients for Chl <i>a</i> at 665 nm (0.0153 m ⁻¹) and C-PC at 620 nm (0.007 m ⁻¹) as given in
8	Simis et al. (2007).
9	
10	The wavelengths used in the medium-independent algorithm developed by Dall'Olmo et al
11	(2003) can be spectrally-tuned to optimise Chl a retrieval. In this study, we adopted the
12	parameterisation of the algorithm used by Gitelson et al. (2008) for MERIS-like sensors:
13	
14	Chl $a = 23.1 + 117.4 \times \{ [R_{rs}^{-1}(600 - 670) - R_{rs}^{-1}(700 - 730)] \times R_{rs}(740 - 760) \} $ (9)
15	
16	We did not re-optimise the algorithm for the retrieval of $Chl a$ so that we could test its ability
17	for generalisation.
18	
19	In addition, we adapted the algorithm of Dall'Olmo et al. (2003) for estimating C-PC. Here,
20	it was necessary to optimise the wavelengths and coefficients used for C-PC retrieval. This
21	was achieved by initially setting $\lambda_1 = 620$ nm (C-PC absorption maximum) and $\lambda_3 = 730$ nm
22	and then regressing the model $[R_r^+]_{620} - R_r^+]_{\lambda_2} \ge R_r (730)$ against the measured
23	concentration of C-PC for values of λ_2 between 560 and 660 nm. In the second iteration, λ_2
24	was set to 600 nm and then the model $[R_r^{\dagger} \langle 620 - R_r^{\dagger} \langle 600 \rangle] = R_r \langle \lambda_3 \rangle$ was regressed against
25	the measured concentrations of Chl <i>a</i> for values of λ_3 between 700 and 775 nm. In the final

1	iteration, λ_3 was set to 725 nm and λ_1 was optimised by regressing the model
2	$[R_r, \lambda_1] - R_r, \lambda_0 = R_r, \lambda_1 = 0$ against the measured concentrations of Chl <i>a</i> for values of λ_1
3	between 610 and 630 nm. This resulted in the following algorithm for C-PC retrieval:
4	
5	$CPC \propto [R_{rs}^{-1}(615) - R_{rs}^{-1}(600)] \times R_{rs}(725) $ (10)
6	
7	The wavelengths used in Eq. 10 differ somewhat from those originally employed for the
8	retrieval of C-PC by Hunter et al. (2008a) ($\lambda_1 = 630$; $\lambda_2 = 660$; $\lambda_3 = 750$). This is mostly
9	likely because the $R_{rs}(0_+,\lambda)$ made in Hunter et al. (2008a) were made from laboratory
10	cultures of cyanobacteria.
11	
12	To evaluate the performance of the algorithms proposed by Gons (1999), Simis et al. (2005)
13	and Dall'Olmo et al. (2003), we developed a number of semi-empirical algorithms for the
14	retrieval of Chl a and C-PC that were optimised for our study lakes. We used the band-ratios
15	$[R_{rs}(705):R_{rs}(670)]$ and $[R_{rs}(705):R_{rs}(620)]$ for the estimation of Chl <i>a</i> and C-PC respectively
16	because these indices have been used effectively in previous research (Hunter et al., 2008b,
17	2009). The semi-empirical algorithms were derived by regressing the band-ratios against the
18	measured concentrations of Chl a and C-PC. In addition, we derived a further algorithm for
19	the retrieval of C-PC by incorporating both the $[R_{rs}(705):R_{rs}(620)]$ and $[R_{rs}(705):R_{rs}(670)]$
20	band-ratios in a multivariate regression model. It was anticipated that this might provide a
21	rudimentary correction for the effect of Chl <i>a</i> absorption on the retrieval of C-PC.
22	
23	The performance of the inversion algorithms was assessed on the basis of the coefficient of
24	determination (R^2) and the root mean square error (RMSE) derived comparison with the
25	measured pigment concentrations. Subsequently, the retrieved concentrations of Chl a and

C-PC from the best-performing algorithms were compared to cyanobacterial cell counts and
 measured MC concentrations by regression analysis.

3

4 **3. Results and discussion**

5 3.1 Cyanobacterial cells, pigments and toxins

The seasonal change in the abundance of cyanobacteria in Esthwaite Water and Loch Leven is shown in Fig. 2 and a summary of the pigment and cell concentrations measured in the lakes concurrent to the airborne overflights is presented in Table 2. The AISA Eagle data collected over Loch Leven on 13 Apr coincided with the end of the spring diatom bloom and the beginning of the clear water phase. The abundance of cyanobacteria at this time was very low (approximately 26 cells cm⁻³). The mean concentrations of Chl *a* and C-PC were consequently also low at 6.58 and 1.15 mg m⁻³ respectively and C-PC:Chl *a* ratio was 0.175.

The AISA Eagle data acquired on 26 Apr over Esthwaite Water coincided with the early stages of a bloom of the potentially-toxic cyanobacterium *Anabaena circinalis*. The mean concentration of cyanobacteria measured in the lake on 26 Apr was approximately 18 767 cells cm⁻³, but this varied considerably among the various sampling stations. The mean concentrations of Chl *a* and C-PC were 10.4 and 16.8 mg m⁻³ respectively; the C-PC:Chl *a* ratio was 1.62 which suggests cyanobacterial dominance (Simis et al., 2005).

20

The CASI-2 flights over Loch Leven on 22 Aug also coincided with a cyanobacterial bloom: in this instance co-dominated by *Aphanothece* spp., *Woronichinia naegeliana* and, to a lesser extent, *Microcystis aeruginosa*. The mean concentration of cyanobacteria on 22 Aug was 25 238 cells ml⁻¹, but again this varied considerably between sampling stations. The mean

- concentrations of Chl *a* and C-PC were 36.3 and 68.9 mg m⁻³ respectively and the C-PC:Chl
 a ratio was 1.90, which again strongly indicates cyanobacterial dominance.
- 3

4 Microcystins were detected by ELISA in both lakes (Fig. 3). Confirmation of the toxins was 5 provided by high performance liquid chromatography with photodiode array detection 6 (Metcalf et al., 2000). This yielded characteristic peaks (abs. max. 238 nm) of multiple MC 7 variants (data not shown). In all instances, the total concentration of MCs measured in the two waterbodies exceeded the current guideline value of 1 mg m⁻³ MC-LR in drinking water 8 9 established by the World Health Organisation (WHO) (WHO, 2002, 2003) and were thus of 10 significance for human health. The lowest mean concentration of total MCs was measured in 11 Loch Leven on 13 Apr and the highest mean concentration was measured in the same lake 12 during the cyanobacterial bloom on 22 Aug. The MCs occurred in dissolved form in the 13 water phase and in the cyanobacteria-containing particulate fraction. The % distribution 14 between these pools varied widely (Fig. 3) from being almost entirely soluble (mean 94% : 15 Loch Leven, April) to being almost entirely particulate (mean 96% : Loch Leven, August). 16 17 This variability in MC compartmentation has implications for the use of remote sensing as a 18 tool for aiding risk assessment activities for human health protection. It is not possible to

19 detect dissolved MCs directly by remote sensing. The concentration of cyanobacterial cells

20 within a MC-producing bloom can, however, be used as an indicator of potential MC

21 concentrations. The implicit assumptions are that (i) the MC pool is predominantly

22 intracellular and (ii) MC concentrations per cell are relatively uniform within and between

- 23 blooms. If a substantial proportion of the total MC pool occurs in the soluble phase, or where
- 24 environmental conditions influence intracellular MC pool sizes, then it cannot necessarily be

assumed that remotely sensed estimates of cyanobacterial cell biomass will provide a reliable
 measure of bloom MC concentrations.

3

4 The largely extracellular proportion (ca. 94%) of the total MC pool detected in Loch Leven 5 may have included an over-winter residual of the MCs released during the decline of the 6 cyanobacterial bloom in the previous autumn. This possibility is raised because: (i) the 7 residence time in Loch Leven is ca. 180 days (Table 1); (ii) although MCs are susceptible to 8 microbial degradation in water, biodegradation rates are temperature-dependent and minimal 9 in winter (Ward, 1999). The very low concentrations of cyanobacterial cells and C-PC 10 measured in Esthwaite on 13 Apr would therefore not have provided a sentinel indication of 11 potential risks to health because of this dissolved MC pool.

12

13 3.2 Atmospheric correction

14 The AISA Eagle $R_{rs}(0^+,\lambda)$ spectra calculated by the FLAASH atmospheric correction model 15 were realistic for Case 2 waters in terms of their shape and magnitude and they demonstrated 16 a reasonable level of correspondence with the in-situ $R_{rs}(0^+,\lambda)$ spectra. The $R_{rs}(0^+,\lambda)$ 17 spectrum calculated by FLAASH and that measured near to Station 01 on Esthwaite Water 18 during the overflights on 26 Apr is shown in Fig. 4. The shape of the $R_{rs}(0^+,\lambda)$ spectrum 19 calculated by FLAASH closely resembles that measured in-situ and the reflectance troughs 20 associated with the Chl a and C-PC absorption peaks have been clearly preserved. Moreover, 21 the FLAASH calculated spectrum mainly lies within the 95% confidence interval for mean 22 in-situ $R_{\rm rs}(0^+,\lambda)$.

23

24 The RMSE of $R_{rs}(0^+,\lambda)$ calculated by FLAASH from the AISA Eagle data for all stations on

Loch Leven on 13 Apr ranged from 0.0048 to 0.0168 (400 to 800 nm) (equating to a

1 RMSE(%) of between 26.4 and 77.7%). The RMSE on the values of $R_{rs}(0^+,\lambda)$ calculated by 2 FLAASH from the AISA Eagle data from Esthwaite Water on 26 Apr ranged from 0.00369 3 to 0.0114 (400 to 800 nm) (equating to a RMSE(%) of between 19.7 and 141.9%).

4

5 Correspondence between reflectance measured in-situ and from the aircraft was strongly 6 wavelength-dependent and generally strongest in the green, red and on the NIR shoulder 7 (520-720 nm) and poorest in the blue (<500 nm) and at NIR wavelengths beyond 750 nm. 8 The relatively large uncertainties observed in the blue region were partly due to the poorer 9 radiometric calibration of the AISA Eagle sensor and strong aerosol and water vapour 10 scattering. The degree of correspondence also deteriorated significantly with the time 11 elapsed between the overflights and the in-situ sampling (Fig. 5).

12

The empirical line correction applied to the CASI-2 data yielded coefficients of determination for the regression equations between $R^2 = 0.76$ and 0.99 for the 13 channels (see Table 3). The accuracy of empirical line correction was generally better for the channels centred at red and near-infrared wavelengths used in the semi-empirical and semi-analytical algorithms pigment retrieval ($R^2 = 0.86-0.98$).

18

19 3.3 Retrieval of Chl a

The relationship between the measured concentrations of Chl *a* and the $[R_{rs}(710):R_{rs}(670)]$ band-ratio was strongly non-linear. The two best-performing semi-empirical algorithms for the retrieval of Chl *a* were derived using quadratic functions of the $[R_{rs}(710):R_{rs}(670)]$ bandratio. The inversion algorithm employing the decadic log of Chl *a* as the response variable provided the marginally better estimation ($R^2 = 0.856$; RMSE = 5.29 mg m⁻³) (see Table 4). This algorithm markedly outperformed the semi-empirical algorithms based upon linear 1 functions of $[R_{rs}(710):R_{rs}(670)]$ and also performed better than the semi-analytical algorithm 2 of Dall'Olmo et al. (2003).

3

4 The Dall'Olmo et al. (2003) algorithm did provide reasonable estimates of Chl a when 5 applied to the data acquired over Esthwaite Water on 26 Apr and the CASI-2 data acquired over Loch Leven on 22 Aug ($R^2 = 0.796$; RMSE = 9.92 mg m⁻³ (44.5%)). However, the 6 7 algorithm greatly underestimated the concentrations of Chl a measured in Loch Leven on 13 Apr ($R^2 = 0.019$; RMSE = 35.6 mg m⁻³ (541%)). The reasons for this are not entirely clear. 8 9 The algorithm is not based upon a ratio of $R_{rs}(\lambda,0+)$ and is thus likely to be more sensitive to 10 shifts in the amplitude of the water-leaving signal. The comparatively low phytoplankton 11 biomass in Loch Leven in early April would certainly have enhanced the contributions from 12 non-phytoplankton particles and bottom reflectance to the water-leaving signal. In addition, 13 the atmospheric correction of the AISA Eagle data from 13 Apr was poorer than that 14 achieved for the other datasets. These factors could have produced shifts in spectral 15 amplitude sufficient to impair algorithm performance.

16

The values retrieved using the algorithm of Gons (1999) were very strongly 17 correlated with the measured concentrations of Chl a (R² = 0.879) (see Fig. 6). However, the 18 19 algorithm tended to overestimate the actual concentrations of Chl a and consequently the RMSE was larger (22.6 mg m⁻³) than that returned by the best-performing semi-empirical 20 21 algorithm. The over-estimation of Chl *a* by the Gons (1999) algorithm might have been the result of using an inappropriate value of $a_{a}^{*}(\mathbf{G}_{b})$, rather than errors in the estimation of 22 a^*_{α} We did not determine a^*_{α} in this study. Instead, we used the value measured 23 by Simis et al. (2005). However, a_{c}^{*} ($\frac{2}{b}$ can vary in response to changes in pigment 24 25 composition and packaging, which is similarly dependent on the physiological status, size

spectra and floristic composition of the phytoplankton community (Bricaud et al., 1995, ,
 2004).

3

Simis et al. (2005) found that $\overset{*}{\mathcal{A}}$ (G,) varied between 0.0138 and 0.0203 m⁻¹ in Lake 4 Ijsselmeer in the Netherlands. The substitution of a^*_{α} (G, $c = 0.0203 \text{ m}^{-1}$ into the Gons 5 (1999) algorithm was sufficient to reduce the RMSE for the estimation of Chl a in Eshtwaite 6 Water and Loch Leven to 11.9 mg m^{-3} (41.4%). This not only emphasises the extent to which 7 a_c^* () can vary, but also why it is important to make rigorous measurements of the specific 8 9 inherent optical properties of waterbodies so that analytically-based algorithms can be 10 properly parameterised for constituent retrieval. There is a clear need for more detailed 11 studies of the inherent optical properties of inland waterbodies to support algorithm 12 development and testing.

13

14 This issue notwithstanding, the coefficients estimated by the semi-analytical 15 algorithm of Gons (1999) were as strongly correlated with the measured concentrations of 16 Chl a as the band-ratios employed in the semi-empirical inversion algorithms. This is 17 perhaps somewhat surprising given that the semi-empirical algorithms were optimised using 18 these specific datasets. It is considered that semi-analytical algorithms are more transferable 19 across different water types. The fact that the accuracy of the semi-analytical algorithms is 20 shown here to be comparable to that achievable with optimised semi-empirical models 21 merely strengthens the case for the wider use of analytical approaches for the retrieval of in-22 water constituents. However, while analytical algorithms are widely used for Chl a retrieval 23 in coastal and ocean waters, they have yet to be widely tested in inland waters. This is a clear 24 priority for future research.

1 3.4 Retrieval of C-PC

2 The best performing semi-empirical algorithm for the retrieval of C-PC was developed using a quadratic model based on the $[R_{rs}(710):R_{rs}(620)]$ band-ratio ($R^2 = 0.982$; RMSE = 4.00 mg 3 m^{-3} (13.8%)) (Table 5). However, all the semi-empirical algorithms for the retrieval of C-PC 4 returned estimates of C-PC with R^2 values >0.86 and RMSEs <10.5 mg m⁻³. Interestingly, 5 6 the multivariate regression model incorporating both the $[R_{rs}(710):R_{rs}(620)]$ and $[R_{rs}(710):R_{rs}(670)]$ band-ratios provided a better estimation of C-PC (R² adj. = 0.935; RMSE 7 8 = 7.13 (24.6)) than the model using only the $[R_{rs}(710):R_{rs}(620)]$ band-ratio ($R^2 = 0.907$; 9 RMSE = 9.22 (31.8)). This suggests that the incorporation of the $[R_{rs}(710):R_{rs}(670)]$ band-10 ratio was partly effective in providing a rudimentary correction for Chl a absorption at 620 11 nm. However, clearly it might not be possible to generalise this type of empirical correction 12 to other waterbodies beyond those considered here. 13

The values of $a_{CPC}(620)$ estimated by the algorithm of Simis et al. (2005) were strongly correlated with the measured concentration of C-PC ($R^2 = 0.984$) and the resulting concentrations of C-PC derived from these coefficients also demonstrated a near 1:1 relationship with measured C-PC values (RMSE = 3.98 mg m⁻³ (13.7%)) (Fig. 6). The accuracy of C-PC retrieval provided by the Simis et al. (2005) algorithm was superior to that achieved by the semi-empirical inversion algorithms.

20

The semi-analytical algorithm based upon the model proposed by Dall'Olmo et al. (2003) for the retrieval of Chl *a* also performed strongly ($R^2 = 0.921$; RMSE = 2.65 mg m⁻³ (9.15%)) when applied to the dataset collected over Loch Leven on 13 Apr and Esthwaite Water on 26 Apr. We could not apply this algorithm to the CASI-2 dataset acquired over Loch Leven on 22 Aug because of the incompatible channel configuration. The Simis et al. (2005) algorithm was not particularly sensitive to the uncertainties in atmospheric correction in spite of the fact
that it uses an absolute value of *R*_{rs}(0+,λ) to estimate the *b_b*(779) coefficient and has
previously been shown to be sensitive to shifts in spectral amplitude (Ruiz-Verdu et al.,
2008). However, it is likely that the any errors in the estimation of *b_b*(779) are relatively
insignificant at high C-PC concentrations.

6

7 The superior performance of the Simis et al. (2005) algorithm is almost certainly attributable 8 to the fact that it corrects for the contribution of Chl *a* to absorption at 620 nm. The relative 9 contribution of Chl a to absorption at 620 nm is obviously strongly dependent on the floristic 10 composition of the phytoplankton community and, more specifically, on the abundance of 11 cyanobacteria containing C-PC. The relative contribution of Chl a to total pigment 12 absorption at 620 nm can thus be expected to be inversely proportional to C-PC:Chl a. The 13 contribution of Chl *a* is relatively insignificant in waters with high C-PC:Chl *a* ratios. 14 However, in waters with low C-PC:Chl a ratios, the increased contribution of Chl a to total 15 absorption at 620 nm is known to lead to errors on the estimation of C-PC (Simis et al., 16 2007). We found that the value of the relative residuals increases significantly at C-PC:Chl a 17 ≤ 0.5 (Fig. 7), in agreement with findings reported elsewhere (Ruiz-Verdu et al., 2008; 18 Randolph et al., 2008; Hunter et al., 2008b).

19

The cell counts made at Esthwaite Water and Loch Leven suggest that C-PC:Chl a = 0.5 is approximately equivalent to 6500 cells ml⁻¹. Importantly, this cell concentration is below the WHO health guidance values for cyanobacteria cell numbers in recreational waters. This is significant because it suggests that this algorithm (and the best-performing semi-empirical algorithms) can detect cyanobacterial cells at concentrations significantly below those at which they would typically pose risks to human health by recreational contact and ingestion.

- This emphasises the fact that remote sensing has the potential to provide early-warnings of
 cyanobacterial bloom development (Hunter et al., 2009).
- 3

4 3.5 Comparisons between cyanobacterial cells, pigments and microcystins

5 The C-PC estimates retrieved using the algorithm of Simis et al. (2005) were weakly, but 6 significantly, related to the concentration of cyanobacterial cells ($R^2 = 0.380$; p = 0.014) (Fig. 7 8a). The retrieved concentrations of C-PC were also significantly correlated with 8 cyanobacterial biovolume as measured in the samples from Loch Leven ($R^2 = 0.483$; p =9 0.026) (no biovolume estimates were available for Esthwaite Water). The retrieved estimates 10 of Chl *a* were not significantly correlated with either cyanobacterial cell numbers or

11 biovolume.

12

24

25

13 The relationship between cyanobacterial cell concentration and that of the C-PC biomarker 14 pigment was notably weaker than those previously reported elsewhere (e.g. Brient et al., 15 2008; Randolph et al., 2008). It is possible that some of the scatter in this relationship might 16 be simply explained by the uncertainties inherent in microscopic cell counts. However, the 17 intracellular concentration of C-PC is also known to be strongly dependent upon irradiance 18 and the supply of fixed nitrogen (Fogg et al., 1973; Jiang & Qiu, 2005; Raps et al., 1983). 19 The relationship between cell and C-PC concentrations measured in the water column is 20 therefore likely to vary in response to seasonal patterns in both light and nutrient supply. 21 22 Intriguingly, in our case the majority of the scatter observed in the relationship between cell 23 numbers and C-PC concentration was the result of within-lake spatial variation rather than

within the individual blooms. This variability might reflect differences in water depth and

seasonal (or between-lake) variation. This implies that C-PC cell quotas were highly variable

circulation among the sampling stations which in turn is likely to have influenced the light
 history (in terms of quality and quantity) of the cyanobacterial cells. That is, positively
 buoyant cells stationed in shallow waters might have experienced significantly higher
 irradiances compared to cells entrained within deeper vertically mixing waters.

5

Heterogeneity in the species composition of the blooms might also have contributed to scatter
in the cell-pigment relationship. Phycobiliprotein cell quotas can vary markedly across
different cyanobacteria (e.g. Patel et al., 2005). This is likely to cause variability in the
association between the concentrations of cells and pigments in the water column where
spatial distribution of species is not uniform. This is particularly likely to occur where
blooms are composed of both buoyancy-regulating and non-buoyancy-regulating species.

12

The Loch Leven bloom on 22 Aug contained both buoyancy-regulating (*Microcystis*, *Woronichinia*) and non-buoyancy-regulating genera (*Aphanothece*). These differing traits might have led to variability in their distribution throughout Loch Leven and consequently influenced the correlation between cell numbers and pigment concentrations. That said, the bloom on Esthwaite Water on 22 Apr was almost entirely dominated by *Anabaena*: thus variability in species composition cannot be used to explain the lack of a strong correlation between cells and pigments in this instance.

20

The relationship between the concentration of cyanobacterial cells and total MCs was also weak and only just significant ($R^2 = 0.251$; p = 0.057). This can be partly explained by the fact that MC production is influenced to some extent by nutrient supply and environmental conditions. Cellular MC content has been shown to correlate positively with nutrient supply and to decrease at high irradiances (Rapala et al., 1997; Wiedner et al., 2003). The accessory

1 phycobiliproteins and MCs thus appear to respond in similar ways to variations in resource 2 supply. Therefore, it is perhaps not surprising that the retrieved concentrations of C-PC in 3 Esthwaite Water and Loch Leven were very strongly correlated with the concentrations of particulate ($R^2 = 0.906$; p = <0.0005) and total MCs ($R^2 = 0.896$; p = <0.0005) (Fig. 8b). 4 5 Strong correlations between the phycobiliprotein and MC content of cyanobacteria have also 6 been reported elsewhere (e.g. Izydorczyk et al., 2005). Chl a was less strongly correlated with total MCs ($R^2 = 0.665$; p = <0.0005). These observations imply that the C-PC content 7 8 of cells might provide a crude measure of bloom MC content (assuming that MC-producing 9 strains are present).

10

11 3.6 Monitoring cyanobacterial populations in lakes

12 Information on the presence and abundance of cyanobacteria in lakes is needed to (i) feed 13 into local, regional and national monitoring activities such as those required under the EU 14 WFD; (ii) provide intelligence to support risk assessment activities to protect human health 15 from toxin-producing blooms; (iii) provide essential information on ecosystem processes and 16 function. However, standard lake monitoring programmes are currently constrained by an 17 inability to obtain this information at high spatial and temporal resolutions. Sampling at a 18 single or a small number of stations will rarely provide a representative estimate of 19 cyanobacterial abundance where blooms are distributed patchily across lakes. Moreover, 20 since blooms can increase and decrease over relatively short time scales (e.g., days to weeks), 21 samples collected on a fortnightly or monthly basis are unlikely to trace the underlying 22 dynamism of cyanobacterial populations in lakes. 23

24 Mounting concerns about the pressures impacting aquatic ecosystems mean that the

25 development of more effective and efficient monitoring systems is becoming an ever-

1 increasing priority. In Europe, the WFD has placed an increased burden on those agencies 2 responsible for monitoring, assessing and managing the ecological status of Europe's surface 3 waters. The metrics being considered for the assessment of lake ecological status include Chl 4 a concentration and cyanobacterial biomass. It has been widely shown that Chl a can be 5 estimated relatively reliably in inland waters using remotely sensed data (Kutser, 2004; Tyler 6 et al., 2006; Gons et al., 2008). This study and others also now show that cyanobacterial 7 biomass estimation by remote sensing is achievable (Simis et al., 2007; Ruiz-Verdu et al., 8 2008; Randolph et al., 2008). Remote sensing-based estimates of phytoplankton biomass 9 could thus strongly contribute to regulatory monitoring of surface water ecological status 10 under the WFD, although operational capability is currently restricted to larger lake systems 11 observable from ocean colour platforms.

12

13 The remotely-sensed estimation of cyanobacterial biomass could also be used to support 14 hazard recognition and risk assessments activities for the protection of human and animal 15 health. It has been shown that remotely sensed information can be integrated with existing alert monitoring protocols such as the WHO guideline levels (GLs) for recreational waters to 16 17 aid the assessment of human health risks (Hunter et al., 2009). The WHO GLs for 18 recreational waters indicate that cyanobacterial cell concentrations of $\geq 20\ 000$ and ≥ 100 000 cells ml⁻¹ equate to low and moderate risks respectively of adverse health effects in 19 20 humans, while the presence of cyanobacterial scums is considered to pose the highest risks to 21 human health. Significantly, we have demonstrated that cyanobacterial populations can be detected using remote sensing at levels well below those currently used to provide human 22 23 health warnings. The use of remote sensing platforms for the early-warning of cyanobacterial 24 blooms in lakes and other waterbodies for health protection would therefore seem to be feasible. 25

2 The value of using remotely sensed imagery to map cyanobacterial populations and provide 3 intelligence for health protection is clearly shown in Figs. 9 and 10. These mapped C-PC 4 image products show that the distribution of cyanobacterial cells in Esthwaite Water and 5 Loch Leven was very patchy with thick surface scums clearly visible on both lakes. This 6 patchiness was almost certainly caused by buoyant cyanobactria becoming entrained within 7 advective currents and massing as scums on windward shorelines (Hunter et al., 2008b). The 8 heterogeneous and transient nature of these scums complicates their detection in the field and 9 this can affect the assessment of health risks.

10

In spite of the fact that thick scums had formed elsewhere on both lakes, cyanobacterial cell concentrations in the water samples collected from the sampling stations on Esthwaite Water and Loch Leven never exceeded 100 000 cells per ml (i.e. the WHO GL for moderate risk). However, the scums recognised in the AISA Eagle and CASI-2 imagery are evidence of much greater risks to human health. This is also supported by the measured concentrations of MCs in these waterbodies. This highlights the extent to which limited sampling at discrete stations in a heterogeneous waterbody may provide inadequate warning of the risks to health.

Spatially synoptic intelligence provided by remote sensing platforms, particularly with the possibility of near real-time reporting, may overcome the limitations of heterogeneity and point sampling (cloud cover permitting). Indeed, the next generation of high spatial and spectral resolution satellite-based imaging spectrometers, such as the German-led Environmental Mapping and Analysis Program (EnMAP), will offer new opportunities for monitoring phytoplankton populations in much smaller waterbodies.

25

The inversion algorithms currently used for the retrieval of constituent concentrations in Case 2 waters were designed primarily for application with multispectral ocean colour sensors. To 3 exploit fully the hyperspectral capabilities of the new generation of spaceborne imaging 4 spectrometers it will be necessary to develop new and better approaches to the inversion of 5 water-leaving optical signals. In this respect, there is clearly a need for further work on the 6 development and testing of inversion algorithms for inland (and coastal) Case 2 type waters.

7

8 4. Conclusions

9 The accuracy of different semi-empirical and semi-analytical inversion algorithms for the 10 retrieval of Chl a and C-PC was examined using high-resolution and multi-temporal AISA 11 Eagle and CASI-2 imagery from two shallow lakes in the UK. The best performing 12 algorithm for the retrieval of Chl *a* was a nonlinear semi-empirical model. The absorption 13 coefficients returned by the semi-analytical algorithm of Gons (1999) were also strongly 14 correlated with the measured pigment concentrations, but the actual retrieved pigment 15 concentrations were subject to significant errors. These were mostly likely due to variability 16 in the specific absorption coefficient of chlorophyll and as such the algorithm could easily be 17 modified to reduce these uncertainties. The best performing inversion algorithm for the 18 retrieval of C-PC was the semi-analytical nested band ratio algorithm of Simis et al. (2005). 19 The absorption coefficients and actual C-PC concentrations returned by this algorithm were 20 strongly correlated with those measured in the water samples collected from the lake. These 21 results demonstrate that the performance of analytically-based algorithms is equal or superior 22 to that of widely used semi-empirical approaches. The greater transferability of the analytical 23 models also makes them better suited to regional or global scale monitoring of phytoplankton 24 pigments in lakes. These results strongly suggest that remotely sensed data might contribute 25 significantly to the proactive and regulatory monitoring of lakes.

2 The concentrations of C-PC retrieved using the semi-analytical nested band ratio algorithm 3 were correlated (albeit weakly) with cyanobacterial cell numbers and biovolume. Very 4 strong correlations were observed between the estimated C-PC concentrations and the 5 measured concentration of MCs. This implies that C-PC concentrations retrieved from 6 remotely sensed data might provide a useful indication of the potential risks to human health. 7 The ability to map cyanobacterial populations at a synoptic scale and recognise the presence 8 of hazardous surface scums is a clear advantage of utilising remotely sensed imagery in this 9 context. This study demonstrated that cyanobacterial populations can be detected using 10 remote sensing at concentrations well below those of human health significance. This 11 suggests that remote sensing platforms can contribute to the early-warning monitoring of 12 cyanobacterial populations for human health protection.

13

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24

25 References

1	Alder-Golden, S. M., Matthew, M. W., Bernstein, L. S., Levine, R. Y., Berk, A., Richtsmeier,
2	S. C., Acharya, P. K., Anderson, G. P., Felde, G., Gardner, J., Hoke, M., Jeong, L. S.,
3	Pukall, B., Ratkowski, A., & Burke, HH. (1999). Atmospheric correction for short-
4	wave spectral imagery based on MODTRAN4. SPIE Proceedings on Imaging
5	Spectrometry, 3753, 61-69.
6	Alikas, K. & Reinart, A. (2008). Validation of the MERIS products on large European lakes:
7	Peipsi, Vanern and Vattern. Hydrobiologia, 599, 161-168.
8	Bennett, A. & Bogorad, L. (1973). Complementary chromatic adaptation in a filamentous
9	blue-green-alga. Journal of Cell Biology, 58, 419-435.
10	Bricaud, A., Babin, M., Morel, A., & Claustre, H. (1995). Variability in the chlorophyll-
11	specific absorption-coefficients of natural phytoplankton - analysis and
12	parameterization. Journal of Geophysical Research-Oceans, 100, 13321-13332.
13	Bricaud, A., Claustre, H., Ras, J., & Oubelkheir, K. (2004). Natural variability of
14	phytoplanktonic absorption in oceanic waters: Influence of the size structure of algal
15	populations. Journal of Geophysical Research-Oceans, 109.
16	Brient, L., Lengronne, M., Bertrand, E., Rolland, D., Sipel, A., Steinmann, D., Baudin, I.,
17	Legeas, M., Le Rouzic, B., & Bormans, M. (2008). A phycocyanin probe as a tool for
18	monitoring cyanobacteria in freshwater bodies. Journal of Environmental Monitoring,
19	10, 248-255.
20	Brierley, B., Carvalho, L., Davies, S., & Krokowski, J. (2007). Guidance on the quantitative
21	analysis of phytoplankton in freshwater samples. In Phytoplankton Classification Tool
22	(Phase 2). Final Report to SNIFFER (pp. 63-88). Edinburgh: SNIFFER.

1	Buiteveld, H., Hakvoort, J. H. M., & Donze, M. (1994). The optical properties of pure water.
2	Ocean Optics XII, 2258, 174-183.
3	Carmichael, W. W., Azevedo, S. M. F. O., An, J. S., Molica, R. J. R., Jochimsen, E. M., Lau,
4	S., Rinehart, K. L., Shaw, G. R., & Eaglesham, G. K. (2001). Human fatalities from
5	cyanobacteria: Chemical and biological evidence for cyanotoxins. Environmental
6	Health Perspectives, 109, 663-668.
7	Carvalho, L. & Kirika, A. (2003). Changes in shallow lake functioning: response to climate
8	change and nutrient reduction. Hydrobiologia, 506, 789-796.
9	Codd, G. A., Azevedo, S. M. F. O., Bagchi, S. N., Burch, M. D., Carmichael, W. W.,
10	Harding, W. R. et al. (2005a). CyanoNet: A global network for cyanobacterial bloom
11	and toxin risk management. International Hydrologolical Programme. Initial
12	situation assessment and recommendations. (Rep. No. 76). UNESCO, Paris.
13	Codd, G. A., Metcalf, J. S., & Beattie, K. A. (1999). Retention of Microcystis aeruginosa and
14	microcystin by salad lettuce (Lactuca sativa) after spray irrigation with water
15	containing cyanobacteria. Toxicon, 37, 1181-1185.
16	Codd, G. A., Morrison, L. F., & Metcalf, J. S. (2005b). Cyanobacterial toxins: risk
17	management for health protection. Toxicology and Applied Pharmacology, 203, 264-
18	272.
19	Dall'Olmo, G. & Gitelson, A. A. (2005). Effect of bio-optical parameter variability on the
20	remote estimation of chlorophyll-a concentration in turbid productive waters:
21	experimental results. Applied Optics, 44, 412-422.

1	Dall'Olmo, G. & Gitelson, A. A. (2006). Effect of bio-optical parameter variability and
2	uncertainties in reflectance measurements on the remote estimation of chlorophyll-a
3	concentration in turbid productive waters: modeling results. Applied Optics, 45, 3577-
4	3592.
5	Dall'Olmo, G., Gitelson, A. A., & Rundquist, D. C. (2003). Towards a unified approach for
6	remote estimation of chlorophyll-a in both terrestrial vegetation and turbid productive
7	waters. Geophysical Research Letters, 30.
8	Dekker, A. G. (1993). Hyperspectral Remote Sensing of Inland Water Quality. The
9	Netherlands: Vrije University, Amsterdam.
10	Ferguson, C. A., Carvalho, L., Scott, E. M., Bowman, A. W., & Kirika, A. (2008). Assessing
11	ecological responses to environmental change using statistical models. Journal of
12	Applied Ecology, 45, 193-203.
13	Fogg, G. E., Stewart, W. D. P., Fay, P., & Walsby, A. E. (1973). The Blue-Green Algae.
14	London and New York: Academic Press.
15	Gitelson, A. A., Dall'Olmo, G., Moses, W., Rundquist, D. C., Barrow, T., Fisher, T. R.,
16	Gurlin, D., & Holz, J. (2008). A simple semi-analytical model for remote estimation
17	of chlorophyll-a in turbid waters: Validation. Remote Sensing of Environment, 112,
18	3582-3593.
19	Gitelson, A. A., Vina, A., Verma, S. B., Rundquist, D. C., Arkebauer, T. J., Keydan, G.,
20	Leavitt, B., Ciganda, V., Burba, G. G., & Suyker, A. E. (2006). Relationship between
21	gross primary production and chlorophyll content in crops: Implications for the
22	synoptic monitoring of vegetation productivity. Journal of Geophysical Research-
23	Atmospheres, 111.

1	Gons, H. J. (1999). Optical teledetection of chlorophyll a in turbid inland waters.
2	Environmental Science & Technology, 33, 1127-1132.
3	Gons, H. J., Auer, M. T., & Effler, S. W. (2008). MERIS satellite chlorophyll mapping of
4	oligotrophic and eutrophic waters in the Laurentian Great Lakes. Remote Sensing of
5	Environment, 112, 4098-4106.
6	Gordon, H. R., Brown, O. B., & Jacobs, M. M. (1975). Computed relationships between
7	inherent and apparent optical properties of a flat homogeneous ocean. Applied Optics,
8	14, 417-427.
9	Havens, K. E. (2008). Cyanobacterial blooms: effects on aquatic ecosystems. In H.K.Hudnell
10	(Ed.), Cyanobacterial Harmful Algal Blooms: State of the Science and Research
11	Needs (pp. 733-747). Berlin: Springer-Verlag.
12	Hillebrand, H., Durselen, C. D., Kirschtel, D., Pollingher, U., & Zohary, T. (1999).
13	Biovolume calculation for pelagic and benthic microalgae. Journal of Phycology, 35,
14	403-424.
15	Hunter, P. D., Tyler, A. N., Gilvear, D. J., & Willby, N. J. (2009). Using remote sensing to
16	aid the assessment of human health risks from blooms of potentially-toxic
17	cyanobacteria. Environmental Science & Technology, 43, 2627-2633.
18	Hunter, P. D., Tyler, A. N., Presing, M., Kovacs, A. W., & Preston, T. (2008a). Spectral
19	discrimination of phytoplankton colour groups: The effect of suspended particulate
20	matter and sensor spectral resolution. Remote Sensing of Environment, 112, 1527-
21	1544.

1	Hunter, P. D., Tyler, A. N., Willby, N. J., & Gilvear, D. J. (2008b). The spatial dynamics of
2	vertical migration by Microcystis aeruginosa in a eutrophic shallow lake: A case
3	study using high spatial resolution time-series airborne remote sensing. Limnology
4	and Oceanography, 53, 2391-2406.
5	Izydorczyk, K., Tarczynska, M., Jurczak, T., Mrowczynski, J., & Zalewski, M. (2005).
6	Measurement of phycocyanin fluorescence as an online early warning system for
7	cyanobacteria in reservoir intake water. Environmental Toxicology, 20, 425-430.
8	Jiang, H. B. & Qiu, B. S. (2005). Photosynthetic adaptation of a bloom-forming
9	cyanobacterium Microcystis aeruginosa (Cyanophyceae) to prolonged UV-B
10	exposure. Journal of Phycology, 41, 983-992.
11	John, D. M., Whitton, B. A., & Brook, A. J. (2003). The freshwater algal flora of the British
12	Isles: an identification guide to freshwater and terrestrial algae. Cambridge:
13	Cambridge University Press.
14	Johnk, K. D., Huisman, J., Sharples, J., Sommeijer, B., Visser, P. M., & Stroom, J. M.
15	(2008). Summer heatwaves promote blooms of harmful cyanobacteria. Global
16	Change Biology, 14, 495-512.
17	Kaufman, Y. J., Wald, A. E., Remer, L. A., Gao, B. C., Li, R. R., & Flynn, L. (1997). The
18	MODIS 2.1-mu m channel - Correlation with visible reflectance for use in remote
19	sensing of aerosol. IEEE Transactions on Geoscience and Remote Sensing, 35, 1286-
20	1298.
21	Kutser, T. (2004). Quantitative detection of chlorophyll in cyanobacterial blooms by satellite
22	remote sensing. Limnology and Oceanography, 49, 2179-2189.

1	Kutser, T. (2009). Passive optical remote sensing of cyanobacteria and other intense
2	phytoplankton blooms in coastal and inland waters. International Journal of Remote
3	Sensing, 30, 4401-4425.
4	Kutser, T., Metsamaa, L., Strombeck, N., & Vahtmae, E. (2006). Monitoring cyanobacterial
5	blooms by satellite remote sensing. Estuarine Coastal and Shelf Science, 67, 303-312.
6	Lorenzen, C. J. (1967). Determination of chlorophyll and pheo-pigments -
7	spectrophotometric equations. Limnology and Oceanography, 12, 343-346.
8	Metcalf, J. S., Banack, S. A., Lindsay, J., Morrison, L. F., Cox, P. A., & Codd, G. A. (2008).
9	Co-occurrence of beta-N-methylamino-L-alanine, a neurotoxic amino acid with other
10	cyanobacterial toxins in British waterbodies, 1990-2004. Environmental
11	Microbiology, 10, 702-708.
12	Metcalf, J. S., Bell, S. G., & Codd, G. A. (2000). Production of novel polyclonal antibodies
13	against the cyanobacterial toxin microcystin-LR and their application for the detection
14	and quantification of microcystins and nodularin. Water Research, 34, 2761-2769.
15	Metcalf, J. S. & Codd, G. A. (2000). Microwave oven and boiling waterbath extraction of
16	hepatotoxins from cyanobacterial cells. FEMS Microbiology Letters, 184, 241-246.
17	Nair, A., Sathyendranath, S., Platt, T., Morales, J., Stuart, V., Forget, M. H., Devred, E., &
18	Bouman, H. (2008). Remote sensing of phytoplankton functional types. Remote
19	Sensing of Environment, 112, 3366-3375.
20	Paerl, H. W. & Huisman, J. (2008). Climate - Blooms like it hot. Science, 320, 57-58.

1	Patel, A., Mishra, S., Pawar, R., & Ghosh, P. K. (2005). Purification and characterization of
2	C-phycocyanin from cyanobacterial species of marine and freshwater habitat. Protein
3	Expression and Purification, 40, 248-255.
4	Pearson, M. J., Bateman, I. J., & Codd, G. A. (2001). Measuring the recreational and amenity
5	values affected by toxic cyanobacteria: a contingent valuation study of Rutland Water,
6	Leicestershire. In R.K.Turner, I. J. Bateman, & W. N. Adger (Eds.), Economics of
7	Coastal and Water Resources: Valuing Environmental Functions (pp. 67-89).
8	Dordrecht, The Netherlands: Kluwer.
9	Pilotto, L. S., Douglas, R. M., Burch, M. D., Cameron, S., Beers, M., Rouch, G. J., Robinson,
10	P., Kirk, M., Cowie, C. T., Hardiman, S., Moore, C., & Attewell, R. G. (1997). Health
11	effects of exposure to cyanobacteria (blue-green algae) during recreational water-
12	related activities. Australian and New Zealand Journal of Public Health, 21, 562-566.
13	Ramos, A. G., Martel, A., Codd, G. A., Soler, E., Coca, J., Redondo, A., Morrison, L. F.,
14	Metcalf, J. S., Ojeda, A., Suarez, S., & Petit, M. (2005). Bloom of the marine
15	diazotrophic cyanobacterium Trichodesmium erythraeum in the Northwest African
16	upwelling. Marine Ecology-Progress Series, 301, 303-305.
17	Randolph, K., Wilson, J., Tedesco, L., Li, L., Pascual, D. L., & Soyeux, E. (2008).
18	Hyperspectral remote sensing of cyanobacteria in turbid productive water using
19	optically active pigments, chlorophyll a and phycocyanin. Remote Sensing of
20	Environment, 112, 4009-4019.
21	Rantajarvi, E., Olsonen, R., Hallfors, S., Leppanen, J. M., & Raateoja, M. (1998). Effect of
22	sampling frequency on detection of natural variability in phytoplankton: unattended

1	high-frequency measurements on board ferries in the Baltic Sea. Ices Journal of
2	Marine Science, 55, 697-704.
3	Rapala, J., Sivonen, K., Lyra, C., & Niemela, S. I. (1997). Variation of microcystins,
4	cyanobacterial hepatotoxins, in Anabaena spp. as a function of growth stimuli.
5	Applied and Environmental Microbiology, 63, 2206-2212.
6	Raps, S., Wyman, K., Siegelman, H. W., & Falkowski, P. G. (1983). Adaptation of the
7	cyanobacterium Microcystis aeruginosa to light-intensity. Plant Physiology, 72, 829-
8	832.
9	Reinart, A. & Kutser, T. (2006). Comparison of different satellite sensors in detecting
10	cyanobacterial bloom events in the Baltic Sea. Remote Sensing of Environment, 102,
11	74-85.
12	Ruiz-Verdu, A., Simis, S. G. H., de Hoyos, C., Gons, H. J., & Pena-Martinez, R. (2008). An
13	evaluation of algorithms for the remote sensing of cyanobacterial biomass. Remote
14	Sensing of Environment, 112, 3996-4008.
15	Sarada, R., Pillai, M. G., & Ravishankar, G. A. (1999). Phycocyanin from Spirulina sp:
16	influence of processing of biomass on phycocyanin yield, analysis of efficacy of
17	extraction methods and stability studies on phycocyanin. Process Biochemistry, 34,
18	795-801.
19	Sathyendranath, S., Watts, L., Devred, E., Platt, T., Caverhill, C., & Maass, H. (2004).
20	Discrimination of diatoms from other phytoplankton using ocean-colour data. Marine
21	Ecology-Progress Series, 272, 59-68.

1	Schalles, J. F. & Yacobi, Y. Z. (2000). Remote detection and seasonal patterns of
2	phycocyanin, carotenoid and chlorophyll pigments in eutrophic waters. Archiv fur
3	Hydrobiologie Special Issues Advances in Limnology, 55, 153-168.
4	Simis, S. G. H., Peters, S. W. M., & Gons, H. J. (2005). Remote sensing of the cyanobacterial
5	pigment phycocyanin in turbid inland water. Limnology and Oceanography, 50, 237-
6	245.
7	Simis, S. G. H., Ruiz-Verdu, A., Dominguez-Gomez, J. A., Pena-Martinez, R., Peters, S. W.
8	M., & Gons, H. J. (2007). Influence of phytoplankton pigment composition on remote
9	sensing of cyanobacterial biomass. Remote Sensing of Environment, 106, 414-427.
10	Smyth, T. J., Moore, G. F., Groom, S. B., Land, P. E., & Tyrrell, T. (2002). Optical modeling
11	and measurements of a coccolithophore bloom. Applied Optics, 41, 7679-7688.
12	Tomlinson, M. C., Wynne, T. T., & Stumpf, R. P. (2009). An evaluation of remote sensing
13	techniques for enhanced detection of the toxic dinoflagellate, Karenia brevis. Remote
14	Sensing of Environment, 113, 598-609.
15	Tyler, A. N., Hunter, P. D., Carvalho, L., Codd, G. A., Elliott, J. A., Ferguson, C. A., Hanley,
16	N. D., Hopkins, D. W., Maberly, S. C., Mearns, K. J., & Scott, E. M. (2009).
17	Strategies for monitoring and managing mass populations of toxic cyanobacteria in
18	recreational waters: a multi-interdisciplinary approach. Environmental Health, 8
19	(Suppl. 1), S11.
20	Tyler, A. N., Svab, E., Preston, T., Presing, M., & Kovacs, W. A. (2006). Remote sensing of
21	the water quality of shallow lakes: A mixture modelling approach to quantifying
22	phytoplankton in water characterized by high-suspended sediment. International
23	Journal of Remote Sensing, 27, 1521-1537.

1	Ward, C. J. (1999). Investigations into the toxicology and persistence of microcystins. PhD
2	Thesis, University of Dundee, Scotland.
3	Westberry, T. K., Siegel, D. A., & Subramaniam, A. (2005). An improved bio-optical model
4	for the remote sensing of Trichodesmium spp. blooms. Journal of Geophysical
5	Research-Oceans, 110.
6	WHO (2003). Guidelines for safe recreational water environments. Vol. 1, Coastal and fresh
7	waters Geneva: World Health Organisation.
8	WHO (2004). Guidelines for drinking-water quality (Rep. No. Vol. 1). Geneva: WHO.
9	Wiedner, C., Visser, P. M., Fastner, J., Metcalf, J. S., Codd, G. A., & Mur, L. R. (2003).
10	Effects of light on the microcystin content of Microcystis strain PCC 7806. Applied
11	and Environmental Microbiology, 69, 1475-1481.