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- CARBON-11, NITROGEN-13 :AND OXYGEN-15 : LABELLED MOLECULES

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ABSTRACT

The development of positron emission tomography (PET) has entailed the preparation of radio-pharmaceuticals labelled with 2^4 emitting isotopes such as ^{11}C ^{13}F and ^{15}O .

The use of these isotopes to label molecules for biological applications has led to the development of specific synthesis methods.

After considerations on the general characteristics of 11 C 13 S 15 O, on the choice of molecule to be labelled and on the labelling plan, some examples of labelled molecules are given: 15 O - gas and 15 O-butanol, 11 C-amino-acids and different 11 C-ligands for receptors studies, enzymatic labelling with 13 N.

CARRON-11, HITHOCEN-13 AND OFFICER-15 - LANGLED HOLECULES

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INTRODUCTION

The development of positron emission temography (PET), which gives an image of the distribution of a labelled tracer in a transverse section of the body, her entailed the preparation of radiopharmscenticals labelled with $\mathfrak g+$ emitting isotopes such as $^{11}\mathrm{C},$ $^{13}\mathrm{H}$ and $^{15}\mathrm{O}.$

The use of these radioisotopes to label molecules for biological applications has led to the development of specific synthesis methods suited to the characteristics of the muclides themselves : short half-life, small mass and high radioactivity required.

CHURAL CHARACTERISTICS OF 11c. 13g AND 150

These radioisotopes are produced by particle accelerators, usually cyclotrons, and decay by emission of positrons which on contact with matter are annihilated giving off two 511 keV γ rays at 180°.

Their short half-life, while an advantage in that the radiation dose received by the patient is relatively small, confines their place of use to the production site itself.

These products, resulting from nuclear transmutation and being so short-lived, may be obtained with a high specific activity (table 1), even if the theoretical value cannot actually be reached because of contamination by stable elements.

Buclide	Half-life	Nuclear reaction most commonly used	Theoretical specific radioactivity (Ci/pHole)
Carbon-11	20.4	14p (p. e) 11c	9.22.10 ³
Witrogen-13	9.% 🚥	14 _H (p, a) 11 _C 12 _C (d, n) 13 _H 14 _H (d, n) 15 _O	1.89.104
Ozygen-15	2.05 🚥	14 _B (d, n) 15 ₀	9.18.104

Table 1

They may consequently be used to label molecules of some toxicity or present in very small quantities in the body.

CHOICE OF HOLECULE TO BE LABELLED AND LABELLING PLAN

The labelling of a new molecule with a short-lived isotope may require some weeks or months according to the difficulty of the synthesis considered, which means that the molecule must be chosen with special care. This choice relies on the collaboration of biologists, physiologists, physicians, chemists... aware that the process which is to be measured must last no longer than three or four times the half-life of the radioelement.

Moreover the tracer must have a high enough uptake rate inside the organ studied, compared with the injected dose (the total radioactivity injected being limited by the dose delivered to the patient), for the number of disintegrations recorded on the organ to give a statistically significant result.

Once the molecule has been chosen the chemist must plan the labelling procedure.

The synthesis with purification must last no longer than 2 to 2.5 times the half-life of the radioelement, i.e. 40 to 50 minutes for Carbon-11, which means optimising the reaction speeds and finding rapid purification methods.

In addition the synthesis, which will need to be repeated several times a week over some months for injection to patients, must be suited to automation in shielded cells.

The position where the radioelement will be incorporated in the molecule must be examined attentively.

Let us take the example of pratosin, a molecule with a strong affinity for peripheral α_1 receptors. The chemist can introduce a carbon-ll atom in one of two positions: one of the methoxy groups CB₃O- or the - C - group (fig. 1) :

Fig. 1 - Possible positions for labelling in Prazosin

Bowever a study of the pharmacokinetics of this molecule in man has shown that O-demethylation occurs more quickly than cut on the amide. It is obviously better therefore to contemplate labeling on the - CO - rather than on the methoxy groups.

These general remarks will now be followed by a few examples of 11 C, 13 H and 15 O-labelled molecules. Let us start with oxygen-15 which, given its helf-life, is suitable only for very simple synthesis.

LABELLING WITH OXYGEN-15

Most 15 O-labelled molecules are produced on line: $|^{15}$ O| - 0 O₂, $|^{15}$ O| - 0 O₂, $|^{15}$ O| - 0 O₃, $|^{15}$ O| - 0 O₄, $|^{15}$ O| - $|^{1$

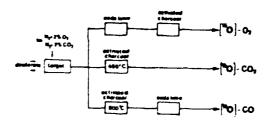


Fig. 2 - On line production of 15 O-labelled molecules

Hitrogen protoxide $|^{15}$ Ol - 12 O has been obtained by catalytic oxidation of amonia (2) (figure 4).

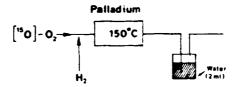


Fig. 3 - Production of 150-labelled water

$$4 \text{ NH}_3 + 4 ^{15}\text{O}_2 \xrightarrow{\text{Pt}} 2 \text{ N}_2^{15}\text{O} + 6 \text{ H}_2\text{O}$$

Fig. 4 - $^{15}0-820$ production from $^{15}0-02$

More recently $|^{15}$ 0| - butanol was synthesized by reaction of oxygen-15 on tributylborane in tetrahydrofurane (3).

 $|CB_3-(CB_2)_3|_3 B + |^{15}0|-0_2 |^{15}0|-CB_3-(CB_2)_3 -08$

After labelling, the butanol is separated out by HPLC in two minutes. Several tens of mCi of $|^{15}$ Ol - butanol have been prepared in this way.

LABELLING WITH CARBON-11

Carbon-11 is incontestably the + emitter most widely used to label molecules for medical applications (4). Before its incorporation in the structure of a molecule, the carbon 11 must be obtained in a highly reactive form known as labelled precursor. The most commonly used of these are listed in table 2.

Carbon-11 is generally recovered from the target in one of two forms: $|^{11}C|$ - $^{11}C|$ - 1

It is most important at this stage to keep the high specific activity of the $\{^{11}C\}-CO_2$ and $\{^{11}C\}-CB_4$ produced in the target, in other words to avoid as far as possible any isotopic dilution with stable carbon. A Labelled molecule will only be a good tracer in fact if the system to be measured is not perturbed, which means that its specific activity must be high.

Very many radiopharmaceuticals have been labelled over the last fifteen years for metabolic or pharmacokinetic research, or to measure certain physiological parameters (flow rate, intracellular pH, membrane permeability... etc). A few illustrations only will be given here.

The first synthesis involved the methylation of demethylated molecules by $|^{11}{\rm C}|$ -methyl iodide. Hethyl iodide is the most widely used radioactive precursor to date.

Examples are the synthesis of $|^{11}\text{Cl}|$ -methyl quinuclidinyl benzylate (5) (fig. 5) or of $|^{11}\text{Cl}|$ methyl 80 15 1788 (6) (fig. 6), ligands for muscarinic receptors and benzodiazepine receptors respectively.

"C. IndeHed procursors upofull for organic synthesis

Table 2 - Precursors for 11C-labelling

$$\begin{array}{c} \text{OH} & \text{OH} &$$

Fig. 5 - | 111C|methyl QNB synthesis

Fig. 6 - | 11 C| methyl RO 15 1788 synthesis

Another kind of reaction with $I^{11}CICS_2I$ is the synthesis of an amino acid, methionine (6), such used to study protein synthesis in a given organ (fig. 7).

Fig. 7 - [11C|methyl methionine synthesis

 $f^{11}C$ accrone has been used to label ligand of β -adrenergic receptors : propranolol, practolol, pindolol (8) (fig. 8).

Fig. 8 - |11c| Pindolol synthesis

 $|^{11}\text{Cl-phosgene}\>$ allows a carbonyl function to be inserted between two stereochemically similar amino functions. This bas been applied to the labelling of different ligands such as pimozide, betanserine and more recently CGP 12 177 (9), a β blocker of very high specificity (fig. 9).

Fig. 9 - | 11c| CGP 12 177 synthesis

Many amino acids have been carbon ll-labelled on the carboxyl group by the Bucherer Strecker method, whereby a hydantolo is prepared by action of KCN on an aldehyde or a ketone then hydrolysed to the corresponding amino acid (fig. 10).

Fig. 10 - Bucherer strecker synthesis

Leucine, valine, tryptophane, phenylalanine have been prepared in this way.

Another more recently reported method involves the action of $\mathbb{H}^{11}\mathbb{C}\mathbb{B}$ on an amino sulphite to give an aminonitrile which is hydrolysed (10) (fig. 11).

$$\begin{array}{c} R_1 \\ C \\ R_2 \end{array} \xrightarrow{\text{NH}_2} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{NH}_2} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} R_1 \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}} \begin{array}{c} H & C \\ R_2 \\ \end{array} \xrightarrow{\text{COOH}$$

Fig. 11 - Amino acid synthesis from aminosulfite

Many other molecules belonging to the sugar, fatty acid and steroid groups have also been synthesized with $^{11}\mathrm{C}.$

LABELLING WITH MITROGEN-13

fitrogen-13 has been used for labelling, though its use is somewhat limited by its 10-minute half-life. However some enzymatic synthesis and one or two organic synthesis have been carried out. ^{13}N may be obtained chiefly in two forms suitable for the purpose: $_{13}\text{N}_{13}$ and $_{13}\text{N}_{13}\text{N}_{23}$. $^{13}\text{N}_{33}$ -may be obtained chiefly in two forms suitable for the purpose: $_{13}\text{N}_{13}$ MH₃ and $_{13}\text{N}_{13}\text{N}_{23}$. $^{13}\text{N}_{33}$ -may be obtained chiefly in two forms suitable for the purpose: $_{13}\text{N}_{13}$ MH₃ and $_{13}\text{N}_{13}\text{N}_{23}$ - $_{13}\text{N}_{33}$ -may be obtained chiefly in two forms suitable for the purpose: $_{13}\text{N}_{13}$ MH₃ and $_{13}\text{N}_{13}$ - $_{13}\text{N}_{13}$ -may be obtained chiefly in two forms suitable for the purpose: $_{13}\text{N}_{13}$ - $_{13}\text{N}_{13}$ -

The general reaction involves the formation of a $|^{13}\rm M|-L$ amino cid from $|^{13}\rm M|-MB_3$ and an a keto acid (fig. 12).

Fig. 12 - Enzymatic synthesis of L amino acid

Certain 13M-amides may also be obtained (fig. 13).

Fig. 13 - Enzymatic synthesis of [13K] smide-asperagine

Of the organic synthesis carried out with 13 M (less than ten compounds altogether) one of the latest is 13 M[phenethylamine (12) (fig. 14).

Fig. 14 - Synthesis of | 13N| phenethylamine

 $^{13}\mathrm{NH}_3$ reacts on phenylproprionyl chloride for 30 seconds then the $^{13}\mathrm{N}\}$ phenylproprionamide undergoes a 5-minute reaction with sodium hypobromite.

GENERAL CONSIDERATIONS ON ^{11}C AND ^{13}N LABELLING

Once the molecules are labelled, an absolutely pure product ready for human injection, must be obtained. The best method of purifying such molecules is high-performance liquid chromatography on an analytical or semi-preparative scale. The separations take less than 10 minutes and the specific activity of the product may be inferred from the resulting chromatogram. The chromatographic solvents must be highly volatile in order to evaporate very quickly after separation.

Separations of products in two forms D and L bave been performed by "Chiral" chromatography. Enzymatic resolutions have also been achieved.

In the case of enzymatic labellings the L form is obtained directly. The increasingly frequent use of enzymes fixed on a solid support reduces the risk of pyrogenicity.

A problem inherent in this type of synthesis is that, because of the short balflife of the radioisotope used it is essential to begin the operation with wery high activities (1 to 1.5 Ci in the case of carbon-il for example). This means that the work must be carried out in shielded cells by means of automatic or at least remoted control systems.

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