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PROTECTION BY WR-151327 AGAINST LATE-EFFECT DAMAGE FROM FISSION-SPECTRUM NEUTRONS

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INTRODUCTION

Considerable effort has been expended to develop chemical agents capable of modifying radiation-induced damage to biological systems (1). Most of this work has focused on maximizing the ability of these agents to protect against acute lethal events (2,3). Maisin and others have demonstrated the ability of chemical agents to protect against radiation-induced late-effect damage in rodents (4-6). The endpoint studied was life shortening, a phenomenon mediated in large part by the onset of lethal radiation-induced tumors (7).

Recent reports have demonstrated that nonlethal genotoxic damage from either low- or high-LET radiations can be significantly reduced by aminothiol radioprotector chemicals (8,9). Protection was observed even when the radioprotectors were administered up to three hours following radiation exposure. Under <u>in vitro</u> conditions, WR-1065, the free thiol of WR-2721, can protect against mutations in V79 cells at the hypoxanthine-quanine phosphoribosyl transferase (HGPRT) locus following exposure to either ⁶⁰Co gamma rays (8) or 0.85-MeV fission-spectrum neutrons (9). These data demonstrated that radiation-induced genotoxic damage can be protected against by the administered after irradiation, no protection against cytotoxicity was observed.

Work in our laboratory has focused on the characterization and application of aminothiol compounds as antigenotoxic agents. WR-1065 can protect against the clastogenic effects of neutrons and gamma rays (10) and ⁶⁰Co-induced transformation in C3H10T1/2 cells (11). WR-2721 protects neonatal rats against radiation-induced preneoplastic lesions (12). We have expanded our studies to determine whether the

chemical radioprotector WR-151327 [S-3-(3-methylaminopropylamino) propylphosphorothiotic acid] can be used to protect mice against the life-shortening effects of fission-spectrum neutrons. This agent affords excellent protection against neutron-induced damage to the jejunum of mice (13), and it can be administered orally.

MATERIALS AND METHODS

The B6CF₁, hybrid mouse (i.e., C57BL/6J ANL female x BALB/cJ ANL male) was used in this study. Animals were 110 days of age at the time of irradiation. Four hundred mice (two hundred of each sex) were randomly assigned to each of the two treatment groups. Animals were routinely housed five per cage, maintained in animal rooms of 200-cage capacity, and provided with access to food and water at all times. Temperature and humidity was controlled to 72 ± 4 F and $50\pm 10\%$ relative humidity. A 12-hour light/dark cycle was always used.

Irradiations were performed using the JANUS reactor (14). Neutron doses were 10 cGy whole body. Animals were individually placed in one-pint polyethylene containers for irradiation and then returned to their cages. Controls were sham-irradiated. All operational logistics and cage locations in the holding rooms were controlled by computer and randomized to avoid experimental biases due to human decision making. Death checks were performed daily, and all mice were examined to ascertain cause of death and other incidental findings.

WR-151327 was supplied by Col. David E. Davidson, Jr. A dose of 580 mg/kg was administered I.P. to animals 30 min prior to irradiation.

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All life-shortening data was subjected to rigorous statistical analysis. The productlimit method described by Kaplan and Meier (15) was used to estimate the survival distribution function for each group in the study. Males and females were analyzed separately, and all analyses were performed on the IBM 3033 at Argonne National Laboratory using the LIFETEST procedure in the SAS software package (16). Data are presented as cumulative survival curves for time intervals of 300 to 850 days and 850 to 1300 days following irradiation.

RESULTS

Animals injected I.P. with 580 mg/kg of WR-151327 and later exposed to JANUS fission-spectrum neutrons were afforded protection against the life-shortening effects of the radiation. A break point of 850 days post irradiation was arbitrarily chosen because it approximates the median survival time of neutron-exposed animals.

The magnitude and timing of protection depended on the sex of the animal. From 300 to 850 days following irradiation, no statistical differences were found between the cumulative survival curves of treated and control male animals (P = 0.30; Fig. 1). In contrast, female animals during this interval were protected (Fig. 2), as demonstrated by the longer survival times of the treated animals (P = 0.02).

Analysis of the cumulative survival curves between 850 and 1300 days showed a different relationship. For male animals, treatment with WR-151327 appeared to enhance survival beyond the 1000-day point (Fig. 3); a comparison of these curves yielded a P value of 0.14. The cumulative survival curve of the protected animals was shifted to longer survival times. No significant difference was observed for female animals

(Fig. 4; P = 0.37), but the cumulative survival curve for the WR-151327 exposed animals was shifted to shorter survival times.

DISCUSSION

Life shortening has been used as a measure of chronic radiation injury because it is an efficient integrator of cumulative multisystemic injury over a lifetime (7). In the B6CF₁ mouse system, 85% of the accelerated deaths due to radiation exposure are related to tumors (7). We have verified this finding in selected groups of animals by histopathological analysis following routine necropsy procedures. A detailed account of these data will be presented elsewhere.

We recently reported that the radioprotectors WR-2721 and WR-151327 can protect against neutron-induced life shortening in the B6CF₁ hybrid mouse system (17). A major mechanism of action implicated in these protective effects is free-radical scavenging (18,19). However, neutrons are believed to cause injury directly, in contrast to low-LET photons, which act indirectly through free-radical formation. While the role of free-radical scavenging can not be ruled out as an important mechanism in the action of these radioprotectors, these agents are known to be effective in binding to DNA and stabilizing chromatin (20,21), affecting DNA synthesis (22), and affecting cell cycle progression (23). Each of these properties can affect the subsequent expression of radiation-induced genotoxic damage.

We describe here differences in the radioprotective effect of WR-151327, depending on the sex of the animal and the post-irradiation time interval considered. The greatest effect in female animals is prior to 850 days post irradiation. The greater

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protection in male animals is seen during the time increment following 850 days after irradiation. While it is difficult at present to ascribe these effects to a particular model, it is suggestive that hormonal factors may play a role in aminothiol protection against radiation-induced life shortening and concomitant tumor induction in the B6CF₁ hybrid mouse system. With respect to subsequent tumor induction, our preliminary findings to be published elsewhere suggest that tumors of lymphoreticular origin are the class of tumors most affected by the administration of a radioprotector prior to irradiation.

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Fig. 1. Effect of WR-151327 on life shortening caused by fissionspectrum neutrons in male mice between 300 and 850 days following irradiation.



Fig. 2. Effect of WR-151327 on life shortening caused by fissionspectrum neutrons in female mice between 300 and 850 days following irradiation.



Fig. 3. Effect of WR-151327 on life shortening caused by fissionspectrum neutrons in male mice between 850 and 1300 days following irradiation.



Fig. 4. Effect of WR-151327 on life shortening caused by fissionspectrum neutrons in female mice between 850 and 1300 days following irradiation.